CHEMICAL COMPOSITION OF ESSENTIAL OIL OBTAINED FROM (ARTEMESIA ABSINTHIUM L.) GROWN UNDER THE CLIMATIC CONDITION OF SKARDU BALTISTAN OF PAKISTAN

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Abstract

The common wormwood *Artemisia absinthium* (L.) is an aromatic perennial herb, which is wildly grown in the Skardu Baltistan. Locally its fresh and dried form is used as both medicinal and insecticidal purposes. This study first time reports the chemical composition of hydro-distilled essential oil obtained from aerial parts of *A. absinthium*. Upon hydro-distillation, it gave dark green colored oil with yield 0.41% (w/w) (based on sheltered dried plant parts). The variation in chemical composition and their relative percentage in essential oil of *A. absinthium* are cited to vary qualitatively and quantitatively according to geographical location and environmental conditions. The GC-MS analysis of *A. absinthium* essential oil obtained from the population collected from Skardu Baltistan also showed variation in chemical composition and their relative percentage. The oil was dominated by Sesquiterpenes (61.01%) and Monoterpenes (17.92%). The major dominating constitutes were Guaiol (19.33%), α -Bisabolol (8.83%), Carveol (6.16%), Chamazulene (5.94%), Limonen-6-ol, Pivalate (5.37%) and Geranyl- α -Terpinene (5.63%), which accounts overall 51.62% of total constitutes identified.

Key words: Artemisia absinthium, Essential oil, GC-MS, Sesquiterpenes, Guaiol.

Introduction

Common wormwood (Artemisia absinthium L.) is an aromatic herb, perennial medicinal plant (Tan et al., 1998; Bora & Sharma, 2010). It is native to temperate region of the world and widely distributed in Afghanistan, China, India, Japan, Kazakhstan, Kyrgyzstan, Pakistan, Russia, North Africa, Europe, North America (Bora & Sharma, 2011). Its extract and oil were traditionally used in ethnopharmacology and ethno-medicine (Abad et al., 2011). From ancient times species of Artemisia i.e., A. absinthium, A. annuaand A. vulgarishave been used in folk medicine (Hayat et al., 2009; Bora et al., 2011b; Bano et al., 2014; Milena et al., 2016). It is also used as antiseptic, stomachic, cardiac stimulant, antispasmodic, anthelmintic and to reducing liver inflammation and restoration of mental function and memory improvement (Howes et al., 2003; Guarrera, 2005). Some recent studies have reported that the genus Artemisia contained potent monoterpenes, sesquiterpenes and volatile acetylenes components and biological properties such as deterrent, antifeedant, insecticidal, acaricidal, antifungal and cytotoxic (antitumor) activities (Bora et al., 2011a; Tatjana et al., 2014; Milena et al., 2016).

Chemical composition and essential oil yield of wormwood is greatly influenced by geographical distribution and climatic conditions for example sabinyl acetate (18.6%) was reported as dominating constituent of *A. absinthium*essential oil from Belgium by Orav *et al.* (2006), *cis*-sabinyl acetate (26.4%) from Canada by Lopes-Lutz *et al.* (2008), dehydrocostus lactone (41.8%) from Ethiopia by Tariku *et al.* (2011), β-pinene (23.8%) from India by Rezaeinodehi & Khangholi (2008), chamazulene (17.8%) from Turkey by Kordali *et al.* (2005), β-thujone (19.8-63.4%) from Serbia and Montenegro by Blagojevica *et al.* (2006), *cis*-epoxyocimene (39.9%) from Spain by Orav *et al.* (2006), *trans*-thujone (33.1%) from USA by Tucker *et al.*, (1993).

In Skardu Baltistan A. absinthium grows wildly on roadsides and barren lands and locally it is used as both medicinal and insecticidal purposes, the sun-dried leaf and flowers powder are used for stomach aches, antiinflammatory, anti- migraine and for high blood pressure and diabetics (Hayat et al., 2009; Bano et al., 2014). Due to ethno-pharmacology, ethno-medicine, insecticidal and antimicrobial properties of A. absinthium, much investigation yet has been performed and the previous literature indicated that geographical distribution greatly influences the chemical composition and essential oil content. Nature has blessed the Skardu Baltistan region with a diversity of wild herbs and shrubs of economic importance. The aim of this study is to analyze the chemical composition of A. absinthium plant grown under the agro-climatic condition of Skardu Baltistan, Pakistan. This study provides the information about the impact of climatic conditions of Skardu Baltistan on chemical composition and oil yield of A. absinthium.

Materials and Methods

Plant materials: The *A. absinthium* leaves and flowers were collected at "Halqa 2 Skardu" (35.18°N 75.37°E) in August 2016 from Skardu Baltistan, Pakistan. The plant was identified by (Hayat *et al.*, 2009), with voucher specimens PUP, PH004 (ART004) submitted to the Herbarium, University of Peshawar, Pakistan.

Extraction of essential oil: The oven dried plant samples (500g) were put in a Clevenger-type apparatusfor 5 hours for hydro-distillation. The plant material was divided into 2 samples of each 250g, and 400ml of distilled water was added to each sample. The oil was collected and dried over anhydrous sodium sulfate and stored in transparent glass vials (1.5ml) (CNW Technologies (Shanghai) Inc.) and was kept at 4°C for further analysis.

Gas chromatography-mass spectrometry (GC-MS): The *A. absinthium* essential oil was analyzed by using a GC-MS (Agilent 6890N GC, Agilent 5973N MS). The GC-MS was equipped with capillary column DB-1(30 m x 0.22mm i.d., 0.25 μ m film thickness). Using helium as a carrier gas at 1.0 mL/min, the injector and detector temperatures were 250°C and 280°C. The oven temperature was programmed from 70°C (2 min) to 250°C (2 °C/min and held for 5 min) and then increased to 270°C (3°C/min, held for 5 min). The injection size was 0.1 mL of 1% solution prepared in *n*-hexane; split ratio was 1:50.MS was taken at 70 eV with mass scan range of 50–800 amu.

Identification of components: The chemical constitutes of oilwere identified by retention index (RIs) determined with relative to the homologous series of *n*-alkanes C_7 - C_{40} (Sigma Aldrich), under identicalexperimental conditions (Adams, 2001), the individual peaks were computer accorded with NIST 05 spectral library and their disintegration arrangements were also compared with the previous literature for further confirmation (Li *et al.*, 2010; Joshi *et al.*, 2013; Zhang *et al.*, 2014; Vieira *et al.*, 2017; NCBI, 2017).

Kovats retention index formula:

$$I = 100 \left[Z + \frac{\log t'_{Ri} - \log t'_{Rz}}{\log t'_{(z+1)} - \log t'_{Rz}} \right]$$

where:

I = retention index for isothermal GC analysis.

 t_{Ri} = adjusted retention time of sample peak.

 t_{Rz}' = adjusted retention time of *n*-alkane peak eluting immediately before sample peak.

 $t_{R(z+1)}$ = adjusted retention time of *n*-alkane peak eluting immediately after sample peak.

z = carbon number of *n*-alkane peak eluting immediately before sample peak.

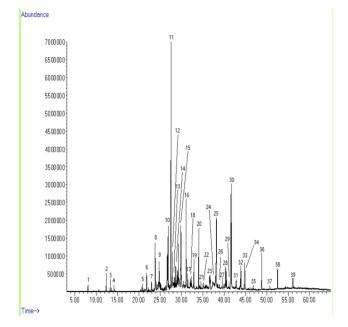
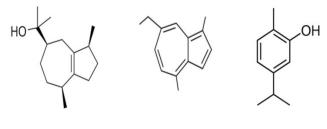


Fig. 1. GC-MS peaks of hydro-distilled *A. absinthium* essential oil obtained from the population grown under the agro-climatic condition of Skardu Baltistan, Pakistan.

Results and Discussion

The hydro-distillation of A. absinthium aerial parts gave dark green-colored oil with a yield of 0.41% (w/w) (based on sheltered dried plant parts). The essential oil chromatogram and composition analyzed by GC-MS are listed in (Table 1, Fig. 1). Analyses revealed that the ecotype is contained of thirty-nine compounds, representing 99.99% of the essential oil. The chemical constitutes were identified by NIST 05 MS spectral library andby retention index (RI) determined with relative to the homologous series of n-alkanes C7-C40 and also compared with previous literature for further confirmation (Joshi, 2014; Zhang et al., 2014; Vieira et al., 2017). The GC-MS analysis revealed that the A. absinthium grown under the climatic condition of Skardu Baltistan was enriched in sesquiterpenes (61.01%) and monoterpenes (17.92%) and their derivatives. The major dominating constitutes were guaiol (19.33%), α-bisabolol (8.83%), carveol (6.16%), chamazulene (5.94%), limonene-6-ol, pivalate (5.37%) and geranyl- α -terpinene (5.63%). Their structures can be seen in Fig. 2. The major six dominating constituent accounts for 51.62% of the total constitutes identified, and their economic importance and uses are presented (Table 2). While other 33 minor components make up the balance are camphor (0.826%), terpinen-4-ol (0.619%), caryophyllene (1.683%), santolinatriene (0.940%), α-copaene (3.505%), germacrene-D (0.450%), β-bisabolene (2.669%), caryophyllene oxide (2.102%), (-)-spathulenol (1.936%), α-santalol (3.481%), cedrol (2.693%), 4-epicubedol (0.676%), cubenol (1.886%), γ-eudesmol (1.187%), 8-epi-γ-eudesmol (1.136%), geranylisobutyrate (2.755%), longifolenaldehyde (0.925%), methyl hinokiate (0.953%), tetrakis (1-methyl)-pyrazine (2.261%), cubedol (1.159%), geranyl-p-cymene (1.629%), nerolidol-epoxyacetate (1.123%), spathulenol (0.734%), heneicosane (1.601%), phytol (1.212%), 1-ethyl-4-methoxy-benzene (0.633%), carvacrol (1.293%), tricosane (1.483%), 1-heptatriacotanol (1.027%), pentacosane (2.102%), heptacosane (1.203%) andnonacosane (0.796%) respectively.



Chamazulene (C14H16)

Guaiol (C15H26O)

Carvacrol (C10H14O)

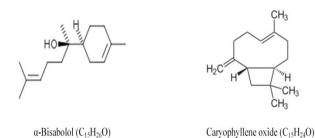


Fig. 2. Chemical structures of some economically important constituents found in *A. absinthium* essential oil.

Table 1. Chemical composition of A. absinthium essential oil.

Peak #	RT ^a	Compounds name ^b	Relative %	KI (Exp) ^c	KI (Lit) ^d	ID ^e
1	12.351	Camphor	0.862	1147	1148	MS,RI
2	13.291	Terpinen-4-ol	0.619	1172	1179	MS,RI
3	21.838	Caryophyllene	1.683	1416	1427	MS,RI
4	22.976	Santolinatriene	0.940	1452		MS
5	23.811	α-Copaene	3.505	1478	1470	MS,RI
6	24.054	Germacrene-D	0.450	1486		MS
7	24.754	β-Bisabolene	2.669	1508	1505	MS,RI
8	26.599	Caryophyllene oxide	2.102	1570		MS,RI
9	26.737	(-)-Spathulenol	1.936	1575	1575	MS,RI
10	26.861	α-Santalol	3.481	1579	1582	MS,RI
11	27.559	Guaiol	19.33	1602	1596	MS,RI
12	27.837	Cedrol	2.693	1612	1610	MS,RI
13	28.356	4-epi-Cubedol	0.676	1631	1627	MS,RI
14	28.64	Cubenol	1.886	1641	1652	MS,RI
15	29.011	γ-Eudesmol	1.187	1654	1652	MS,RI
16	29.113	8- <i>epi</i> -γ-Eudesmol	1.136	1657	1653	MS,RI
17	29.269	Geranylisobutyrate	2.755	1663	1685	MS
18	29.844	α-Bisabolol	8.833	1683	1684	MS,RI
19	31.067	Chamazulene	5.943	1728	1710	MS,RI
20	31.455	Longifolenaldehyde	0.925	1742		MS
21	32.355	Methyl hinokiate	0.953	1776		MS
22	32.92	Tetrakis(1-methyl)-Pyrazine	2.261	1797	1797	MS,RI
24	36.568	Cubedol	1.159	1941	1939	MS,RI
25	36.748	Geranyl-p-Cymene	1.629	1948		MS
26	37.999	Nerolidol-epoxyacetate	1.123	1999		MS
27	38.176	Geranyl-a-terpinene	5.636	2007		MS
28	39.549	Spathulenol	0.734	2066	2071	MS,RI
29	40.341	Heneicosane	1.601	2100	2100	MS,RI
30	40.507	Phytol	1.211	2107		MS
31		Limonen-6-ol, Pivalate	5.371	2154		MS
32		Carveol	6.167	2161		MS
33	43.784	1-ethyl-4-methoxy-benzene	0.633	2256		MS
34	43.922	Carvacrol	1.293	2262	2252	MS,RI
35	44.735	Tricosane	1.483	2300		MS,RI
36	44.931	1-Heptatriacotanol	1.027	2309		MS
37	48.786	Pentacosane	2.102	2500		MS
38	52.539	Heptacosane	1.203	2700		MS
39	56.106	Nonacosane	0.796	2899		MS
	22.100	Total identified	99.9			
		Oil yield (%)	0.41			
		Monoterpenes	17.92			
		Sesquiterpenes	61.01			
		Others	21.06			

^aRetention time

^bCompounds are listed in order of their retention time

^cLinear retention index on DB-1 (30 m x 0.22 mm i.d., 0.25 μ m film thickness), experimentally determined using homologous series of C₇-C₄₀ *n*-alkanes

^dLinear retention index taken from Adams (2007) and/or NIST 08 (2008)

eIdentification methods: based on comparison with authentic compounds, NIST 08 MS databases; RI, based on comparison of calculated RI with those reported in Adams or NIST 08

S. No.	Compounds name		Uses	Source
1.	Guaiol	1.	Toxicity against <i>Plutella xylostella</i> with LD ₅₀ values of 0.07 and 8.9 mg/larva and against <i>Musca domestica</i> , with LC ₅₀ values of 3.5 μ L/L and 16.9 μ L/L	Tao <i>et al.</i> , 2013
		2.	Act as amelanogenesis inhibitor in human skin	Baschong et al., 2008
		3.	Complete feeding inhibition in aphids occurs at concentration of 70 mg/L	Liu et al., 2013
		4.	Showed larvacidal and mosquito biting-deterrent at concentration of 25 $\mu L/L$	Ali et al., 2015
2.	Chamazulene	1.	Anti-inflammatory properties and inhibits the CYP1A2 enzyme.	Safayhi et al., 1997
		2.	Knock down effect against Rhizoctonia solani at 12.5mL/20mL.	Bouzenna et al., 2013
		3.	LC50 Rhysopertha dominica occur at 50 mg/L topical appliaction	Bouzenna et al., 2013
		4.	Phytotoxic activity against <i>Arvense persicaria</i> , <i>Chenopodium album</i> at 100 mg/L	Solymosi, 2000
3.	α-Bisabolol	1.	Glioma cells treated with high concentration of α -bisabolol (10 μ M) resulted in a 100% cell death	Cavalieri et al., 2004
		2.	α -Bisabolol at the dose of 25 g/L caused 100 mortality after 72 hours of application against <i>Bemisia argentifolii</i>	Corpas-Lopez et al., 2015
		3.	α -Bisabolol at oral doses of 50 and 100 mg/kg markedly in rats reduced gastric damage up to 87% and 96%, respectively	de Andrade et al., 2004
	Carvacrol	1.	Used in food industries for their flavoring and preservative properties	Marchese et al., 2016
		2.	Soya sauce containing a combination of thymoland carvacrol (<0.0157%) killed > 7 log CFU/ml of foodborne pathogen	Moon et al., 2016
4.		3	Showed antioxidant, anti-inflammatory, local anesthetic, cicatrizing, antiseptic, and especially antibacterial and antifungal activities	Marchese et al., 2016
		4	LC ₅₀ values 28.52 mg/L against <i>Pochazia shantungensis</i> adults and nymphs using leaf dip bio assay	Park et al., 2017
5.	Caryophyllene oxide	1	It alters several key pathways for cancer development i.e. I3K/AKT/mTOR/S6K1 and STAT3 pathways	Nagappan et al., 2016
		2	Antitumor and apoptotic effects on MG-63 human osteosarcoma cells	Pan et al., 2016

 Table 2. Some economically important constitutes identified in A. absinthium essential oil obtained from the population collected from Skardu Baltistan.

According to the previous literature, the chemical composition and essential oil content within the same species of A. absinthium were greatly influenced by the geographical distribution (Telci et al., 2006; Okut et al., 2017; Fatima et al., 2018), climate conditions and time of harvesting (Joshi, 2013). Similarly, the chemical constituents and oil yield of coriander (Coriandrum sativum L.) collected from two different locations of Tunisian were not similar (Sangwan et al., 2001). According to the previous literature the A. absinthium essential oil yield and its chemical composition collected from different geographical locations of the world showed great variations (Telci et al., 2006), for example, sabinyl acetate (18.6%) was the dominant constitutes of wormwood collected from Belgium (Orav et al., 2006), trans-verbenol (11.6%) from Lithuania (Judzentiene et al., 2009), 1, 8-cineol (3.3%) from Germany (Orav et al., 2006), cis-linalool oxide (34.2%) from Spain (Bailen et al., 2013), and borneol (18.7%) from India (Joshi, 2014). The geographical distribution and climatic condition greatly influenced the chemical composition and their relative percentage in the oil, as our result revealed that the chemical composition of A. absinthium grown under the agro-climatic condition of Skardu Baltistan have variation in chemical composition and their relative percentage as compared to the previous literature.

Conclusions

It was observed in the current investigation that *A. absinthium* grown in the Skardu Baltistan, Pakistan showed considerable variation in chemical composition and in the percentage of essential oil yield. The reported yields for essential oil obtained from *Artemisia* sp. range from 0.1 to 1.46 % (w/w) (Msaada *et al.*, 2015). However, in the current investigation greater amount of oil was present 0.41% (w/w). Its composition resembles a previous report and differs from others suggesting the existence of chemo-types also in this plant. The geographical distribution and climatic conditions of Skardu Baltistan influenced the chemical composition and their relative percentage in essential oil as the oil was dominated by guaiol (19.33%).

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References

Abad, M. J., L.M. Bedoya, L. Apaza and P. Bermejo. 2011. The Artemisia L. genus: A review of bioactive essential oils. Molecules, 17: 2542-2566.

- Ali, A., Tabanca, N.B. Demirci, E.K. Blythe, E.K. Ali, K.H.C. Baser and I.A. Khan. 2015. Chemical composition and biological activity of four salvia essential oils and individual compounds against two species of mosquitoes. J. Agric. Food. Chem., 63: 447-456.
- Adams, R.P. 2007. Identification of essential oil components by GC-MS. Carol Stream (IL): Allured Publication. 456p.
- Bailen, M., L.F. Julio, C. Diaz, J. Sanz, R.A. Martinez-Diaz, R. Cabrera and A. Gonzalez-Coloma. 2013. Chemical composition and biological effects of essential oils from *Artemisia absinthium* L. cultivated under different environmental conditions. *Indust. Crop Product*, 49: 102-107.
- Bano, A., M. Ahmad, T.B. Hadda, A. Saboor, S. Sultana, M. Zafar, M.P.Z. Khan, M. Arshad and M.A. Ashraf. 2014. Quantitative ethno-medicinal study of plants used in the Skardu valley at high altitude of Karakoram-Himalayan range, Pakistan. J. Ethnobio. Ethnomedic, 10: 43-50.
- Baschong, W., G. Heinemann and D. Ochs. 2008. Use of guaiol for treating the skin. U.S. Patent 7: 320-425, issued September 16, 2008.
- Blagojevica, P., N. Radulovica, R. Palica and G. Stojanovica. 2006. Chemical composition of the essential oils of Serbian wild-growing *Artemisia absinthium* and *Artemisia vulgaris*. J. Agric. Food. Chem., 54: 4780-4789.
- Bora, K.S. and A. Sharma. 2011a. the genus *Artemisia*: a comprehensive review. *Pharm. Biol.*, 49: 101-109.
- Bora, K.S. and A. Sharma. 2010. Phytochemical and pharmacological potential of *Artemesia absinthium* L. and *Artemisia asiatica*: a review. J. Pharm. Res., 3: 325-328.
- Bora, K.S and A. Sharma. 2011b. Evaluation of antioxidant and free-radical scavengingpotential of *Artemisia absinthium*. *Pharm. Biol.*, 49: 1216-1223.
- Bouzenna, H. and L. Krichen. 2013. Pelargonium graveolens L. and Artemisia arborescens L. essential oils: chemical composition, antifungal activity against Rhizoctonia solani and insecticidal activity against Rhysopertha dominica. Nat. Prod. Resh., 27: 841-846.
- Cavalieri, E., M. Sofia, F. Cinzia, A.C. Prati, G. Rossella, L. Stefano, B.V. Luigi, L.M. Giuliana, C.R. Anna and S. Hisanori. 2004.α-Bisabolol, a nontoxic natural compound, strongly induces apoptosis in glioma cells. *Biochem. Biophysic. Resh. Comm.*, 315: 589-594.
- Corpas-Lopez, V., M.M. Francisco, M.N.M. Concepción, M.E. Gemma, D.S. Victoriano and M.S. Joaquina. 2015. (-)-α-Bisabolol a promising oral compound for the treatment of visceral *Leishmaniasis*. J. Nat. Prod., 78: 1202-1207.
- de Andrade, I.L., J.N.S. Bezerra, M.A.A. Lima, R.A.P. de Faria, M.A.S. Lima, M. Andrade-Neto and A.L.M. Mesquita. 2004. Chemical composition and insecticidal activity of essential oils from *Vanillosmopsis pohlii* against *Bemisia* argentifolii. J. Agric. Food. Chem., 52: 5879-5881.
- Guarrera, P.M. 2005. Traditional phototherapy in central Italy (Marche, Abruzzo, and Latium). *Fitoterapia*, 76: 1-25.
- Hayat, M. Q., M.A. Khan, M. Ashraf and S. Jabeen. 2009. Ethnobotany of the genus Artemisia L. (Asteraceae) in Pakistan. Ethnobot. Resh. Applic., 7: 147-162.
- Howes, M.R., N.S. Perry and P.J. Houghton. 2003. Plants with traditional uses and activities, relevant to the management of Alzheimer's disease and other cognitive disorders. *Phytother. Res.*, 17: 1-18.
- Joshi, R.K. 2013. Volatile composition and antimicrobial activity of the essential oil of *Artemisia absinthium* growing in Western Ghats region of North West Karnataka, India. *Pharma. Bio.*, 51: 888-892.
- Judzentiene, A, F. Tomi and J.Casanova. 2009. Analysis of essential oils of *Artemisia absinthium* L. from Lithuania by GC-MS and 13C NMR. *Natural Produc. Commun.*, 4: 1113-1118.

- Kordali, S., A. Cakir, A. Mavi, H. Kilic and A. Yildirim. 2005. Screening of chemical composition and antifungal and antioxidant activities of the essential oils from three Turkish Artemisia species. J. Agric. Food. Chem., 5: 1408-1416.
- Li, J. L., X. Dong, F.X.J. Zheng and W. Shan. 2010. Determination of the volatile composition in essential oil of *Descurainia sophia* (L.) Webb ex Prantl (Flixweed) by GC/MS. *Molecules.*, 15: 233-240.
- Liu, T., C.J. Wang, H.Q. Xie and Q. Mu. 2013. Guaiola naturally occurring insecticidal sesquiterpene. *Nat. Prod. Commun.*, 10: 1353-1354.
- Lopes, L.D., D.S. Alviano, C.S. Alviano and P.P. Kolodziej. 2008. Screening of chemical composition, antimicrobial, and antioxidant activities of *Artemisia absinthium* essential oils. *Phytochem.*, 69: 1732-1738.
- Marchese, A., I.E. Orhan, M.D, B. Ramona, D.L. Arianna, F.N. Nabavi, O.G.M. Izadi and S.M. Nabavi. 2016. Antibacterial and antifungal activities of thymol: A brief review of the literature. *Food Chem.*, 210: 402-414.
- Msaada, K., N. Salem, O. Bachrouch, S. Bousselmi, S. Tammar, A. Alfaify, A.K. Sane, A. Ben, S.W. Azeiz and H.A. Brahim. 2015. Chemical composition and antioxidant and antimicrobial activities of wormwood (*Artemisia absinthium* L.) essential oils and phenolics. J. Chem., 5: 1-12.
- Melliou, E., E. Stratis and I. Chinou. 2007. Volatile constituents of propolis from various regions of Greece and its antimicrobial activity. *Food. Chem.*, 103: 375-380.
- Milena. R., G. Janina and B. Barbara. 2016. Effect of water extracts from *Artemisia absinthium* L. on feeding of selected pests and their response to the odor of this plant. J. *Cent. Euro. Agric.*, 17: 188-206.
- Moon, H. and M.S. Rhee. 2016. Synergism between carvacrolandthymol increases the antimicrobial efficacy of soy sauce with no sensory impact. *Int. J. Food. Microbio.*, 217: 35-41.
- Nagappan, T., P. Ramasamy, M.E.A. Wahid, T.C. Segaran and C.S. Vairappan. 2011. Biological activity of carbazole alkaloids and essential oil of *Murraya koenigii* against antibiotic resistant microbes and cancer cell lines. *Molecules*, 16: 9651-9664.
- Okut, N., Y. Mehmet, S. nilufer,and Y. bunyamin. 2017. Chemical composition of essential oil of *Mentha longifolia* L. Longifolia growing wild. *Pak. J. Bot.*, 49: 525-529.
- Orava, A., A. Arakb, M.U. Muiseppa and T. Kailas. 2006. Composition of the essential oil of *Artemisia absinthium* L., of different geographical origin. *Proc. Estonian. Acad. Sci. Chem.*, 55: 155-165.
- Pan, Z., S.K. Wang, X.L. Cheng, X.W. Tian and J. Wang. 2016. Caryophyllene oxide exhibits anti-cancer effects in MG-63 human osteosarcoma cells via the inhibition of cell migration, generation of reactive oxygen species and induction of apoptosis. *Bangladesh J. Pharma.*, 11: 817-823.
- Park, J.H., Y.J. Jeon, C.H. Lee, C. Namhyun and H.S. Lee. 2017. Insecticidal toxicities of carvacrol and thymol derived from *Thymus vulgaris* L. against *Pochazia shantungensis* Chou and Lu. Newly recorded pest. *Sci. Report*, 7: 40902.
- Rezaeinodehi, A, and S. Khangholi. 2008. Chemical composition of the essential oil of *Artemisia absinthium* growing wild in Iran. *Pak. J. Biol. Sci.*, 11: 946-9.
- Fatima, S., F. Ahmed, M. Hameed and A. Rashid. 2018. Ecology and species association of grass species in response to altitudinal gradient in the Potohar region. *Pak. J. Bot.*, 50: 41-49.
- Sangwan, N.S., A.H.A. Farooqi, F. Shabih and R.S. Sangwan. 2001. Regulation of essential oil production in plants. *Plant Growth Reg.*, 34: 13-21.

- Safayhi, H., J. Sabieraj, E.R. Sailer and H.P. Ammon. 1997. Chamazulene an antioxidant type inhibitor of leukotriene B4 formation. *Planta Medica.*, 60: 410-413.
- Solymosi, P. 2000. Herbicidal activity of chamazulene. *Novenyvedelem.*, 36: 119-123.
- Tan, R.X., W.F. Zheng and H.Q. Tang. 1998. Biologically active substances from the genus Artemisia. Planta Med., 64: 295-302.
- Tariku, Y., A. Hymete, A. Hailu and J. Rohloff. 2011. Evaluation of antileishmanial activity and toxicity of essential oils of *Artemisia absinthium* and *Echinops kebericho*. Chem. Biodivers., 8: 614-623.
- Tatjana, M.K. Boris, J. Jovana, I. Budimir, D. Mragoljub, M. Jelena, L.R. Ljubisa, D. Vladimir and Z. Bojan. 2014. Anti-microbial, antioxidative, and insect repellent effects of *Artemisia absinthium. Essential OilPlanta Med.*, 80: 1698-1705.

- Telci, I., O.G. Toncer and N. Sahbaz. 2006. Yield essential oil content and composition of *Coriandrum sativum* grown in two different locations. *J. Essential Oil Res.*, 18: 189-193.
- Tucker, A.O., J.M. Maciarello and S. George. 1993. The essential oils of Artemisia 'Powis Castle' and its putative parents, A. absinthium and A. arborescens. J. Essential Oil Res., 5: 239-242.
- Vieira, T.M., H.J. Dias, T.C. Medeiros, C.O. Grundmann, M.H and E.O. Silva. 2017. Chemical composition and antimicrobial activity of the essential oil of Artemisia absinthium L. J. Essential Oil Bearing Plan, 3: 1-9.
- Zhang, N., T.L. Hu, W. Wang, K. Zhou and Z. Zhang. 2014. Insecticidal, fumigant, and repellent activities of sweet wormwood oil and its individual components against red imported fire ant workers (Hymenoptera: Formicidae). J. Insect. Sci., 14: 241-259.

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