CHEMICAL CONSTITUENTS OF ESSENTIAL OILS OF *BOESENBERGIA ARMENIACA* AND *B. STENOPHYLLA* (ZINGIBERACEAE) ENDEMIC TO BORNEO

NOOR ATIEKAH MD NOR AND HALIJAH IBRAHIM^{*}

University of Malaya, Faculty of Science, Institute of Biological Sciences, Wilayah Persekutuan Kuala Lumpur, 50603 Malaysia *Corresponding author's email: ihalijah@um.edu.my

Abstract

Boesenbergia armeniaca and B. stenophylla are two endemic wild gingers from Borneo. Limited information are available on these wild species. The aim of this study was to identify the leaf essential oil constituents using hydrodistillation and analysed via GC-FID and GC-MS. The essential oil yields of leaves obtained from B. armeniaca and B. stenophylla were 0.01% and 0.08% respectively. The leaf oil of B. armeniaca was dominated by oxygenated sesquiterpenes (47.95%), while the leaf oil of B. stenophylla was dominated by oxygenated monoterpenes (85.12%). The main volatile constituents in the leaf oil of B. armeniaca were Nerolidol (42.55%), Linalool (11.63%) and β -caryophyllene (6.25%) while in the leaf oil of B. stenophylla, methyl cinnamate (83.17%) was most abundant. To date, this is the first report revealing the essential oil components of Boesenbergia armeniaca and B. stenophylla from Sabah, Borneo.

Key words: Methyl cinnamate, Boesenbergia armeniaca, Boesenbergia stenophylla, Essential oil.

Introduction

In recent years, interest in the utilization of essential oils from medicinal plants for scientific research are increasing especially for pharmaceutical, nutritional and cosmeceutical industry (Ksouri *et al.*, 2009). This is due to the fact that the oils exhibit various potent biological activities including antimicrobial (Kamal & Dhruva, 2010), antioxidant (Bellik, 2014) and anti-inflammatory (Alina *et al.*, 2014) activities. Essential oils are secondary metabolites that plants produce for their defence mechanism against herbivory and disease infections (Suthisut *et al.*, 2011). In general, among the important constituents of essential oils are monoterpene and sesquiterpene hydrocarbons, oxygenated monoterpenes and other derivatives (Kamaliroosta *et al.*, 2013).

Zingiberaceae is one of the largest families of the order Zingiberales found in tropical and subtropical forests, with approximately 50 genera and over 1500 species worldwide (Larsen *et al.*, 1999). In the Southeast Asian region, uniqueness of the flavour and medicinal properties of these species lead to their uses as spices, medicines and flavouring agents (Yob *et al.*, 2011). Members of Zingiberaceae are aromatic due to the presence of various essential oil components among others include β zingiberene, linalool and 1,8-cineole (Joseph *et al.*, 2001; Bickers *et al.*, 2003; Tripathi *et al.*, 2013).

Belonging to the tribe Zingibereae, the genus *Boesenbergia* comprises of approximately 80 extant species distributed throughout India to Southeast Asia (Saensouk & Larsen, 2001). The centre of diversity for the genus *Boesenbergia* is mainly in Borneo with more than 21 species followed by Thailand. The habitats of *Boesenbergia* species are mainly the undergrowth areas of tropical and subtropical forests, in particular, damp, humid and shady places in mixed deciduous and evergreen forests and also limestone hills (Gobilik & Limbawang, 2010; Jing *et al.*, 2011). Growing *Boesenbergia* species outside their natural habitats such as a garden may be difficult but possible (Gobilik &

Limbawang, 2010). To date, among the *Boesenbergia* species only *B. rotunda* is widely cultivated commercially (Techaprasan *et al.*, 2006).

Rhizome of *B. rotunda* have been proven to contain panduratin A which was found to inhibit the growth of MCF-7 human breast cancer and HT-29 human colon adenocarcinoma cells, as well as for the treatment of rheumatism, gout and for culinary purposes (Techaprasan *et al.*, 2006: Kirana *et al.*, 2007: Chong *et al.*, 2012). Several species of *Boesenbergia* such as *B. longiflora* and *B. pulchella* have been reported to contain bioactive components such as flavonoids and chalcones which have been reported to have antioxidant and anti-inflammatory properties (Salama *et al.*, 2013; Sudsai *et al.*, 2013; Yuliana *et al.*, 2013).

Boesenbergia stenophylla, a perennial rhizomatous wild ginger is known as 'Jerangau merah' (Malay) and Kaburo adak (Kelabit/Lun Bawang) which is endemic in the highlands of Borneo with altitude of not less than 3000 ft. (Ahmad & Jantan, 2003; Toyat *et al.*, 2015). Ethnobotanically, *B. stenophylla*, has been utilised for various purposes by many ethnic groups, among others include protection against convulsions, prevention of intoxication, cough relief, increase libido, treatment of food poisoning etc. (Ahmad & Jantan 2003; Jing *et al.*, 2010).

Boesenbergia armeniaca is also endemic to Borneo distributed from western Sabah to Brunei. It is not commonly used as a medicinal plant by the local people in Sabah probably due to lack of information with regards to its medicinal properties. However, Jing *et al.*, (2010) and Sarwar Muhammad (2012) reported that the major flavones of luteolin present in the crude extract of *B. armeniaca* might act as a catalyst in contributing to the anticancer activities, especially against MCF-7 cell lines (breast cancer). In addition, the presence of polyphenols (naringin and hesperidin) in the methanol extract of stem and rhizome of *B. armeniaca*, may contribute to the antioxidant activities (Jing *et al.*, 2010). To the knowledge of the authors, there has been no studies on the essential oil of *B. armeniaca*.

Many researchers have demonstrated several pharmacological properties of Zingiberaceae species but studies on biological activities of *Boesenbergia* species are very few including research on the essential oils. Hence, the aim of this study is to analyse and identify the chemical constituents of hydrodistilled leaf oils of *B. armeniaca* and *B. stenophylla* collected from Sabah. Identification of the volatile compounds can provide useful information with regard to their potential applications.

Material and Methods

Plant material: The leaves of *B. armeniaca* and *B. stenophylla*, collected from Serinsim sub-station and Long Pasia, Sabah, were processed at the Institute for Tropical and Conservation Biology of University of Malaysia Sabah, Malaysia (UMS). The leaves were rinsed, cut into small pieces and ground in a blender. All the plants were identified by a taxonomist at the institute and voucher specimens were deposited at BORNEENSIS, Universiti Malaysia Sabah (BORH).

Isolation of essential oils: The fresh samples of ground leaves (1000g) of *B. armeniaca* and *B. stenophylla* were added with distilled water (5L) and subjected to hydrodistillation using modified Clavenger apparatus for about 8 hours. About 5-10 ml of pentane was added through the top of the condenser to trap the condensed oil. Later, the mixtures of water and pentane were dried over anhydrous sodium sulphate and the pentane solution was then evaporated by nitrogen blower to give yellowish essential oils. The oils were stored in dark vials at 4°C until further use.

GC-MS analyses: GC analyses of the volatile constituents were carried out with an Agilent GC model 7890 A, equipped with flame ionization detector (FID) and a CBP-5 capillary column (30 m length x 250 µm internal diameter x 0.25 µm film coating). Helium was used as a carrier gas at the flow rate of 1.3mL/minute. The oven temperature was programmed from 50°C to 250°C at 3°C/min with an initial hold time of 1 minute at 50°C, followed by final hold time of 3 minutes at 230°C. Detector temperature was maintained at 250°C. The sample (1 µL) was injected in split ratio (20:1) at 250°C. GC/MS analyses were performed on Agilent MSD detector 5975C and a CBP-5 capillary column (30 m length x 250 µm internal diameter x 0.25 µm film coating). The operating conditions were as follows: injection and detector temperatures were set at 250°C respectively. A series of n-alkanes, C8-C20 and C21-C40 was subjected to GC-FID to calculate the retention indices (RIs) of samples.

Identification of components: Identification of essential oil constituents was done on the basis of their retention indices (RI) determined with reference to homologous series of *n*-alkanes, matching spectra with MS library search (NIST/Wiley/Adams) and by comparing with the MS literature data (Sivasothy *et al.*, 2011).

Kovats retention index formula:

Kovats Index =
$$\frac{100 \left[\text{Log} \left(\text{Tx} - \text{Tm} \right) - \text{Log} \left(\text{Tn} - \text{Tm} \right) \right]}{\left[\text{Log} \left(\text{Tn} + 1 - \text{Tm} \right) - \text{Log} \left(\text{Tn} - \text{Tm} \right) \right]} + 100 \text{ (N)}$$

where:

Tm = Mobile phase retention time

Tx = Sample component retention time

Tn = Standard hydrocarbon containing carbon retention time

N = Lowest carbon value of n-alkane

Results and Discussion

The leaf oils of *B. armeniaca* and *B. stenophylla* were obtained by hydrodistillation and characterized using GC-FID and GC-MS analyses. Upon hydrodistillation, the yield oil of *B. armeniaca* and *B. stenophylla* were 0.01% and 0.08%, respectively. The yield of the essential oils depends on the extraction type, the duration of the extraction as well as the age of the plants during harvesting (Mejdoub & Katsiotis, 1998; Onyenekwe & Hashimoto, 1999). The identified components of the oil with their Kovat indices (KI) are presented in Table 1 while the chromatograms of *B. armeniaca* and *B. stenophylla* are shown in Figs. 1 and 2, respectively.

The yellowish-coloured oil of B. armeniaca revealed thirty-eight components which represented 99.24% of the total oil composition. Analysis of the oil revealed that the oxygenated sesquiterpenes (47.95%) were found to be the major group in the leaf oil of B. armeniaca, followed by sesquiterpene hydrocarbons (20.69%), oxygenated monoterpenes (14.49%) and monoterpene hydrocarbons 5.99%). The oil mainly comprises of nerolidol (42.55%), linalool (11.63%), and β -caryophyllene (6.25%). Their chemical structures and economic importance are shown in Fig. 3 and Table 2, respectively. Other quantitatively significant constituents include β-pinene (4.83%), βbisabolene (3.96%), phytol (3.15%), α-gurjunene (3.13%), elemol (2.67%) and α -caryophyllene (1.58%). To the best of our knowledge, the chemical profile of the essential oil of B. armeniaca has not been reported before.

A total of twenty-three volatile constituents were identified in the oil of *B. stenophylla* representing 100% of the total oil composition. Oxygenated monoterpenes constitute the major group with 85.12% of the total oils. This is followed by monoterpene hydrocarbons (6.78%), sesquiterpene hydrocarbons (6.44%) and oxygenated sesquiterpenes (0.51%). This oil is highly rich in methyl cinnamate (83.17%). The chemical structure and economic importance of methyl cinnamate are shown in Fig. 4 and Table 2, respectively. Other constituents that are present in appreciable amounts include β -pinene (4.84%), α caryophyllene (3.59%). (R)- α -pinene (1.22%), and linalool (1.18%). A similarly high percentage of methyl cinnamate was reported to be present in *Alpinia malaccensis* var. *nobilis* (60.26%) collected from Pahang (Vejayan *et al.*, 2017).

Ahmad & Jantan (2003) reported that the leaf oil of *B.* stenophylla collected from Sarawak was dominated by (E)methyl cinnamate (49.9%). Several similar constituents were identified in both studies namely β -pinene, borneol, α terpineol, and δ -elemene but present in different compositions. The variations in some of the essential oil constituents may be due to the difference in geographical region, genetic makeup, growth stage and harvest time (Figueiredo *et al.*, 2008; Okut *et al.*, 2017).

No	Table 1. Chemical composition of the leaf oils of B. an Components	rmeniaca a KI ^a	ind <i>B. stenoj</i> KI ^b		D ~
<u>No.</u>	Components			B. a	B. s
1.	Methylcyclohexane	800	740	0.22	0.14
2.	(R)-α-pinene	937 052	-	0.84	1.22
3.	Camphene 9 Diverse	952 070	954 070	-	0.23
4.	β-Pinene	979 002	979 1020	4.83	4.84
5.	β-Phellandrene	993 1022	1029	-	0.11
6.	Limonene	1032	1029	-	0.11
7.	1,8-Cineole	1035	1031	0.78	-
8.	β-Ocimene	1053	-	0.32	0.27
9.	Linalool	1103	1100	11.63	1.18
10.	(E)-2-Butenoic acid, 2-(methylenecyclopropyl)prop-2-yl ester	1120	-	1.15	-
11.	Borneol	1170	1169	0.22	0.55
12.	L-4-terpineol	1181	-	0.42	-
13.	α-Terpineol	1194	1189	0.91	0.22
14.	δ-Elemene	1341	1335	-	0.31
15.	α-Copaene	1378	1377	0.95	0.21
16.	β-Caryophyllene	1394	1417	6.25	0.82
17.	Methyl cinnamate	1392	1379	-	83.17
18.	β-Elemene	1402	1393	0.21	-
19.	α-Gurjunene	1418	1410	3.13	-
20.	γ-Elemene	1426	1434	-	0.21
21.	α-Bergamotene	1439	-	0.66	-
22.	α-Elemene	1446	-	0.32	-
23.	epi-β-Santalen	1451	-	0.48	-
24.	α-caryophyllene	1457	-	1.58	3.59
25.	Isocaryophyllene	1464	-	0.32	-
26.	Germacrene D	1484	1487	0.51	0.58
27.	β-Bisabolene	1507	1505	3.96	-
28.	β-Sesquiphellandrene	1521	1523	1.46	-
29.	δ-Cadinene	1536	1514	0.46	0.22
30.	Elemol	1555	1550	2.67	-
31.	Hotrienol	1539	-	0.53	-
32.	Elixene	1568	-	-	0.5
33.	Nerolidol	1570	1539	42.55	0.19
34.	4-Methyl-1,5-Heptadiene	1582	-	1.54	-
35.	β-Gurjunene	1586	-	0.40	-
36.	Isobutyl cinnamate	1612	1623	0.85	_
37.	Humulene epoxide II	1612	1608	-	0.32
38.	1, 7, 7-Trimethyl-2-vinylbicyclo[2.2.1]hept-2-ene	1634	-	-	0.61
39.	γ-eudesmol	1637	1652	0.70	-
40.	β-eudesmol	1655	1651	0.64	_
41.	α-eudesmol	1658	1645	0.85	-
42.	cis-a-Santalol	1701	-	0.54	_
43.	Hexadecanal	1718	_	1.07	-
44.	Phytol	1915	1943	3.15	-
45.	Isophytol	1913	1948	1.03	_
45. 46.	Kaurene	2033	2042	0.56	0.4
40. 47.	1, 6, 10, 14, 18, 22-Tetracosahexaen-3-ol, 2, 6, 10, 15, 19, 23-	2033		0.50	-
+/.	hexamethyl-, (all-E)-	2037	-	0.55	-
	Total	99.24	100		
	Oil yield	99.24 0.01	0.08		
	•	0.01 5.99			
	Monoterpene hydrocarbons		6.78 85 12		
	Oxygenated monoterpenes	14.49 20.60	85.12		
	Sesquiterpene hydrocarbons	20.69	6.44		
	Oxygenated sesquiterpenes	47.95	0.51		
	Others indices determined on a CBP-5 capillary column	10.12	1.15		

1 Chamical composition of the leaf oils of R1...11.

^aKovats indices determined on a CBP-5 capillary column ^bKovats retention indices taken from Adams (2001) and/or literature

B.a: Boesenbergia armeniaca

B.s: Boesenbergia stenophylla

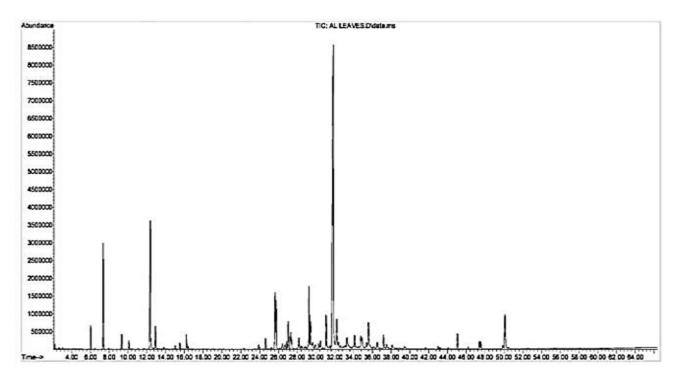


Fig. 1. GC-MS peaks of hydro-distilled leaf oil of Boesenbergia armeniaca collected from Sabah.

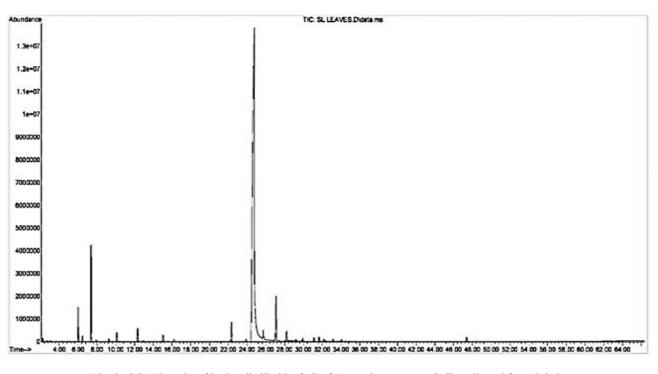
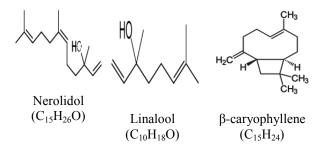
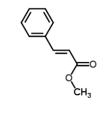


Fig. 2. GC-MS peaks of hydro-distilled leaf oil of Boesenbergia stenophylla collected from Sabah.





Methyl cinnamate (C₁₀H₁₀O₂₎

Fig. 3. Chemical structures of some economically important volatile constituents found in the leaf oil of *B. armeniaca*.

Fig. 4. Chemical structure of methyl cinnamate found in the leaf oil of *B. stenophylla*.

and <i>B. stenophylla</i> obtained from Sabah.							
Compounds	Species	Properties	References				
Nerolidol	B. armeniaca	1. Act as fragrance ingredient in decorative cosmetics, shampoos and	Lapczynski et al., 2008				
		non-cosmetic products such as detergents.					
		2. Exhibited anti-inflammatory, antinociceptive and anti-schistomotal	Silva et al., 2014 &				
		activities.	Fonseca et al., 2015				
		3. Nerolidol displayed gastroprotective activity by inhibiting the	Klopell, 2007				
		formation of ulcers induced by physical and chemical agents in	1				
		dose-dependent manner (50, 250, 500 mg/kg)					
Linalool	B. armeniaca	1. Showed anti-spasmodic activity with 65% of inhibition or	Rekha et al., 2014				
		stimulation and 70% of anti-inflammatory activity.					
		2. Linalool at the IC ₅₀ values f 224 μ M, 222 μ M, and 290 μ M	-				
		exhibited cytotoxic effects against cancer cell of SW 620, T-47D	Chang & Shen, 2014				
		and Hep G2, respectively using WST-1 analysis.					
		3. Linalool gives pleasant odour and also known to elicit	-				
		physiological effects such as inducing calmness and enhancing	Elsharif et al., 2015				
		sleep.					
β-caryophyllene	B. armeniaca	1. Showed significant anti-inflammatory effect on rats at 0.1 ml/kg	Bakir et al., 2008				
		dosage of β -caryophyllene.					
		2. β-caryophyllene demonstrated selective antibacterial activity	Dahham et al., 2015				
		against Staphylococcus aureus (MIC 3 \pm 1.0 μ M) and also	1				
		exhibited selective anti-proliferative effects against colorectal	1				
		cancer cells (IC50 19 µM).					
Methyl cinnamate	B. stenophylla	1. E-isomer of methyl cinnamate from essential oil of Ocimum	Vasconcelos-Silva et al.,				
		micranthum exhibit vasorelaxant effects which tested on rat with	2014				
		IC_{50} value of 877.6 μ mol/L.					
		2. Act as a source of fragrant ingredient that can be found in fine					
		fragrances, shampoos and detergents since methyl cinnamate is	2012				
		reported to have a pleasant fragrance.					
		3. Exhibited antioxidant activity with IC_{50} values of 32.67mg/ml					
		using DPPH free radical scavenging assay.	Vejayan et al., 2017				

Table 2. Some economically important constituents identified in the leaf oils of *B. armeniaca* and *B. stemphylla* obtained from Sabab

Conclusion

This is the first report on the chemical profiling of the leaf oil of B. armeniaca from Borneo. In the case of B. stenophylla, apart from the study on the essential oil constituents from Sarawak, this is the first report for B. stenophylla leaf oil constituents from Sabah. This study reveals that the leaf oil of B. armeniaca constitute three main components namely nerolidol (42.55%), linalool (11.63%), and β -caryophyllene (6.25%). Boesenbergia stenophylla from Sabah is mainly composed of methyl cinnamate (83.17%). The presence of high concentration of nerolidol (42.55%) in the leaf oil of B. armeniaca suggest that it can be potentially utilized as a natural source for nerolidol. This can be a good sample for further studies on biological activities, particularly for the pharmaceutical industry. As for the leaf oil of B. stenophylla, the high percentage composition of methyl cinnamate (83.17%) will be particularly useful for the perfume industry.

References

- Adams, R.P. 2001. Identification of essential oil components by Gas Chromatography/Quadrupole Mass Spectroscopy. Allured Publishing Corporation, Carol Stream (IL), USA.
- Ahmad, F. and I. Jantan. 2003. The essential oils of Boesenbergia stenophylla R. M. Sm. a natural sources of methyl (E)-cinnamate. Flavour Frag. J., 18: 485-486.
- Alina_C.M.C., R.L. Rocío, R.M.M. Aurelio, C.M.M. Margarita, R.G. Angélica and J.A. Rubén. 2014. Chemical composition and in vivo anti-inflammatory activity of *Bursera morelensis* Ramirez essential oil. *Taylor & Francis Group*, 17(5): 758-768.

- Bakır, B., A. Him, H. Özbek, E. Düz and M. Tütüncü. 2008. Investigation of the anti-inflammatory and analgesic activities of β-caryophyllene. *Int. J. Essen. Oil Ther.*, 2: 41-44.
- Bellik, Y. 2014. Total antioxidant activity and antimicrobial potency of the essential oil and oleoresin of *Zingiber* officinale Roscoe. Asian Pac. J. Trop. Dis., 4(1): 40-44.
- Bickers, D., P. Calow, H. Greim, J.M. Hanifin, A.E. Rogers, J.H. Saurat, I.G. Sipes, R.L. Smith and H. Tagami. 2003. A toxicologic and dermatologic assessment of linalool and related esters when used as fragrances ingredients. *Food Chem. Toxicol.*, 41: 919-942.
- Chang, M.Y. and Y.L. Shen. 2014. Linalool exhibits cytotoxic effects by activating antitumor activity. *Molecules*, 19: 6694-6706.
- Chong, T.E., L.Y. Kee, C.C. Fei, H.C. Han, W.S. Ming, C.T.L. Ping, F.G. Teck, N. Khalid, N.A. Rahman, S.A. Karsani, S. Othman, R. Othman and R. Yusof. 2012. *Boesenbergia rotunda*: from ethnomedicine to drug discovery. *Evidbased Compl. Alt. Med.*, 25 pages.
- Dahham, S.S., Y.M. Tabana, M.A. Iqbal, M.B. Ahamed, M.O. Ezzat, A.S. Majid and A.M. Majid. 2015. The anticancer, antioxidant and antimicrobial properties of the sesquiterpene β -caryophyllene from the essential oil of *Aquilaria crassna*. *Molecules*, 20(7): 11808-11829.
- Elsharif, S.A., A. Banerjee and A. Buettner. 2015. Structure-odor relationships of linalool, linalyl acetate and their corresponding oxygenated derivatives. *Front. Chem.*, 3(57): 1-10.
- Figueiredo, A.C., J.G. Barroso, L.G. Pedro and J.J.C. Scheffer. 2008. Factors affecting secondary metabolite production in plants: volatile components and essential oils. *Flavour Frag. J.*, 23(4): 213-226.
- Fonseca, D.V., P.R.R. Salgado, F.L. Carvalho, M.G.S.S. Salvadori, A.R.S. Penha, F.C. Leite, L.C.M. Pordeus and R.N. Almeida. 2015. Nerolidol exhibits antinociceptive and anti-inflammatory involvement of the GABAergic and proinflammatory cytokines. *Fund. Clin. Pharmacol.*, 30(1): 14-22.

- Gobilik, J. and S. Limbawang. 2010. Notes on species composition and ornamental gingers in Tawau Hills Park, Sabah. *J. Trop. Biol. Conserv.*, 7: 31-48.
- Jing, L.J., M. Mohamed, A. Rahmat and M.F. Abu Bakar. 2010. Phytochemicals, antioxidant properties and anticancer investigations of the different parts of several gingers species (*Boesenbergia rotunda*, *Boesenbergia pulchella* var *attenuata* and *Boesenbergia armeniaca*). J. Med. Plants Res., 4(1): 27-32.
- Jing, L.J., M.F. Abu Bakar, M. Mohamed and A. Rahmat. 2011. Effects of selected *Boesenbergia* species on the proliferation of several cancer cell lines. *J. Pharmacol. Toxicol.*, 6: 272-282.
- Joseph, R., T. Joseph and J. Joseph. 2001. Volatile essential oil constituents of *Alpinia smithiae* (Zingiberaceae). *Rev. Biol. Trop*, 49(2).
- Kamal, L.B. and K.J. Dhruva. 2010. Comparative chemical constituents and antimicrobial activity of normal and organic ginger oils (*Zingiber officinale* Roscoe). Int. J. Appl. Biol. Pharm., 4(1): 259-266.
- Kamaliroosta, Z., L. Kamaliroosta and A.H. Elhamiraa. 2013. Isolation and identification of ginger essential oil. J. Food Biosci. Technol., 3: 73-80.
- Kirana, C., G.P. Jones, I.R. Record and G.H. McIntosh. 2007. Anticancer properties of panduratin A isolated from *Boesenbergia pandurata* (Zingiberaceae). J. Nat. Med., 61: 131-137.
- Klopell, F.C., M. Lemos, J.P.B. Sousa, E. Comunello, E.L. Maistro, J.K. Bastos and S.F. Andrade. 2007. Nerolidol, an antiulcer constituent from the essential oil of *Baccharis dracunculifolia* DC (Asteraceae). Z. Naturforsch, 62: 537-542.
- Ksouri, R., H. Falleh, W. Megdiche, N. Trabelsi, B. Mhamdi, K. Chaieb, A. Bakrouf, C. Magne and C. Abdelly. 2009. Antioxidant and antimicrobial activities of the edible medicinal halophyte *Tamarix gallica* L. and related polyphenolic constituents. *Food Chem. Toxicol.*, 47(8): 2083-2091.
- Lapczynski, A., S.P. Bhatia, C.S. Letizia and A.M. Api. 2008. Fragrance material review on nerolidol (isomer unspecified). *Food Chem. Toxicol.*, 46: 5247-5250.
- Larsen, K., H. Ibrahim, S.H. Khaw and L.G. Saw. 1999. *Gingers of Peninsular Malaysia and Singapore*. (1st Ed) Natural History Publications, Borneo, Malaysia.
- Mejdoub, R. and S. Katsiosis. 1998. Factors influencing the yield and the quality of the obtaining essential oil from the leaves of *Eucalyptus citriodora* Hook. Growing in Crete. *Sci. Pharm.*, 66(93): 93-105.
- Okut, N., M. Yagmur, N. Selcuk and B. Yildirim. 2017. Chemical composition of essential oil of *Mentha longifolia* L. subsp. Longifolia growing wild. *Pak. J. Bot.*, 49(2): 525-529.
- Onyenekwe, P.C. and S. Hashimoto. 1999. The composition of the essential oil of dried Nigerian ginger (*Zingiber* officinale Roscoe). Eur. Food Res. Technol., 209: 407-410.
- Rekha, B., C. Muthukumar, S.V. Bakiyalakshmi and G. Shakila. 2014. *In vitro* pharmacological activity of essential oil – Linalool from *Jasminum polyantham*. *Pharmacol. Toxicol.*, 1(1): 1-6.
- Saensouk, S. and K. Larsen. 2001. *Boesenbergia baimii*, a new species of Zingiberaceae from Thailand. *Nord. J. Bot.*, 21(6): 595-597.

- Salama, S.M., M.A. Abdulla, A.S. Alrashdi and A.H.A. Hadi. 2013. Mechanism of Hepatoprotective effect of *Boesenbergia rotunda* in Thioacetamide-Induced liver damage in rats. *Evid-based Compl. Alt. Med.*, 13 pages.
- Sarwar, M. 2012. Anti-cancer activity exhibited by Boesenbergia species plants: An experimental study. Res. J. Med. Plant, 6(1): 2.
- Sharma, C.K. and S.S. Kanwar. 2012. Synthesis of methyl cinnamate using immobilized lipase from *B. licheniformis* MTCC-10498. *Res. J. Recent Sci.*, 1(3): 68-71.
- Silva, M.P.N., G.L.S. Oliveira, R.B.F. Carvalho, D.P. Sousa, R.M. Freitas, P.L.S. Pinto and J. Mores. 2014. Antischistomotal activity of the terpene nerolidol. *Molecules*, 19: 3793-3803.
- Sivasothy, Y., W.K. Chong, A. Hamid, I.M. Eldeen, S.F. Sulaiman and K. Awang. 2011. Essential oils of *Zingiber* officinale var. rubrum Theilade and their antibacterial activities. Food Chem., 124: 514-517.
- Sudsai, T., C. Wattanapiromsakul and S. Tewtrakul. 2013. Inhibition of nitric oxide production by compounds from *Boesenbergia longiflora* using lipopolysaccharidestimulated RAW264.7 macrophage cells. *Songklanakarin* J. Sci. Technol., 35(3): 317-323.
- Suthisut, D., P.G. Fields and A. Chandrapatya. 2011. Fumigant toxicity of essential oils from three Thai plants (Zingiberaceae) and their major compounds against *Sitophilus zeamais, Tribolium castaneum* and two parasitoids. J. Stored Prod. Res., 47: 222-230.
- Techaprasan, J., C. Ngamriabsakul, S. Klinbunga, S. Chusacultanachai and T. Jenjittikul. 2006. Genetic variations and species identification of Thai *Boesenbergia* (Zingiberaceae) analyzed by chloroplast DNA polymorphism. J. Biochem. Mol. Biol., 39(4): 361-370.
- Toyat, A.J., N.A.P. Abdullah, R. Go, T.L. Abdullah, G. Saleh, M. Jiwan, F.R. Kundat and M.M. Magiman. 2015. Conservation strategies for Jerangau Merah (*Boesenbergia* stenophylla) using DNA profiling and micropropagation. *IJAFP*, 1: 89-101.
- Tripathi, M., P. Chawla, R. Upadhyay and S. Trivedi. 2013. Essential oils from family Zingiberaceae for antimicrobial activity- A review. *Int. J. Pharma Bio Sci.*, 4(4): 149-162.
- Vasconcelos-Silva, A.A., F.J. B. De Lima, T.S. De Brito, S. Lahlou and J.C. Magalhães. 2014. Vasorelaxation induced by methyl cinnamate, the major constituent of the essential oil of *Ocimum micranthum*, in rat isolated aorta. *Clin. Exp. Pharmacol. P.*, 41: 755-762.
- Vejayan, J., N.E. Selladuri, H. Ibrahim, A.S. Shuib and M.M. Yusoff. 2017. Biological activities of essential oils hydrodistillated from two closely related ginger species: *Alpinia malaccensis* var. *nobilis* and *Alpinia latilabris* leaves. J. Essent. Oil Bear. Pl., 20(4): 959-971.
- Yob, N.J., S.M. Joffry, M.M. Affandi, L.K. Teh, M.Z. Salleh and Z.A. Zakaria. 2011. *Zingiber zerumbet* (L.) Smith: A review of its ethnomedicinal, chemical, and pharmacological uses. *Evid-based Compl. Alt. Med.*, vol. 2011, Article ID 543216, 12 pages.
- Yuliana, N.D., S. Budijanto, R. Verpoorte and Y.H. Choi. 2013. NMR metabolomics for identification of adenosine A1 receptor binding compounds from *Boesenbergia rotunda* rhizomes extract. J. Ethnopharmacol., 150: 95-99.

(Received for publication 3 November 2017)