

PROFILE OF PLANT SPECIES IN THE TROPICAL DRY FOREST OF TOLIMA (COLOMBIA) EXHIBITING ANTHELMINTIC ACTIVITY IN SHEEP

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Abstract

Phytotherapy is an area of growing scientific interest. Studies on anthelmintic bioactivity of plant species provide new alternatives to face problems such as gastrointestinal parasitism, resistance to synthetic chemical products, environmental impact, and residual activity in products of animal origin. To know the chemical profile of plants in the tropical dry forest (TDF) of Tolima with anthelmintic activity in sheep, a systematic study based on the document "The Tropical Dry Forest in Colombia", which includes 2569 plant species among native, naturalized, and exotic plants from Tolima, was conducted. By using the SCOPUS database, the academic Google search engine, and Microsoft Office 2010 Excel, each species was designated with the same criterion to identify those with anthelmintic activity and qualitative phytochemical content; then, the groups with the highest analysis were identified, and the species with simultaneous analysis were selected from the 3 chemical groups with the highest frequency, and the weight gain AND sheep search criteria was applied to the resulting group. 45 native, 6 naturalized, and 20 exotic species with anthelmintic activity were found. The Fabaceae (13 species), Amaranthaceae (4 species), Annonaceae, Apocynaceae, Euphorbiaceae, and Malvaceae (3 species each) families stand out. The three chemical groups with the high LEVELS OF PHYTOCHEMICALS were tannins, flavonoids, and alkaloids, and at a medium level saponins, steroids, and terpenoids. Simultaneous content of tannins, flavonoids and alkaloids was found in 19 native, 3 naturalized, and 12 exotic species, 5 of which present scientific report of use in sheep feed.

Keywords: **Keywords:** bioactivity, internal parasites, secondary metabolites, plants.

Introduction

The use of regional vegetational resources for controlling or treating diseases in humans and animals has been increasing worldwide. This has been accompanied by research and the discovery of new molecules with biological activity against various diseases (carcinogenic, parasitic, bacterial, viral, fungal, metabolic disorders, etc.).

On the other hand, the context of farmed animal production presents environmental, economic, social, and commercial demands to develop proposals for obtaining safe products (meat, milk, and eggs) that are natural-environment friendly, and that generate economic benefits to producers and improve their life quality.

The starting point of the research on the use of vegetation resources for therapeutic purposes in animal health is based on documentary and ethnoveterinary studies to know both the scientific community findings on the composition and activity of plants under controlled conditions of observation and experimentation, as well as the aspects related to plant species, preparation, application, location, and origin of knowledge in the communities in the targeted region.

Once this initial part is approached, qualitative and quantitative phytochemical studies to identify and isolate fractions and biomolecules to, subsequently, develop the necessary biological tests to verify their activity can be initiated. The purpose of the present documentary study is to define the vegetal species of the tropical dry forest (TDF) of the department of Tolima with scientific anthelmintic activity in sheep.

Materials and Methods

The work was carried out in the TDF areas of the department of Tolima, and our bibliographic reference base, was Annex 1 of "The Tropical Dry Forest in Colombia", document, edited by Pizano and García which

includes 2569 plant species that include native, naturalized and exotic species.

Based on the species reported for the Tolima department, the SCOPUS database search criteria entered was the scientific name of the AND anthelmintic plant species restricted to abstracts only, with a data range from 2001 to 2017, and all kind of documents. Subsequently, to verify the results of the biological activity, the corresponding articles were reviewed.

For the group of plants with anthelmintic activity, the SCOPUS database and academic Google search criteria applied was AND phytochemical scientific name, which permitted to know the phytochemical qualitative analysis (reducing sugars, alkaloids, steroids, terpenoids, tannins, glycosides, carbohydrates, flavonoids, saponins, phenols, cardiac glycosides, and other metabolites).

By using Microsoft Office 2010 Excel, a filter was applied to select plants that, according to the report in the scientific literature, simultaneously contained the three secondary metabolites with the highest scoring peak. Finally, the list of plants resulting in the previous phases was searched on SCOPUS by plant's scientific name AND weight gain, restricted to abstracts only, with a data range of all years to this current year, and all kinds of documents criteria, and depending on the results, the word 'sheep' was also used, obtaining a list of plants reported to be used in food and anthelmintic activity.

Results and Discussion

Tropical Dry Forest Plants Associated with Anthelmintic Activity: The following are the native plants identified in the Tolima TDF with anthelmintic activity: *Trianthema portulacastrum* (Family: Aizoaceae); *Alternanthera sessilis*, *Amaranthus spinosus*, *Amaranthus viridis* and *Celosia argentea* (Family: Amaranthaceae); *Spondias mombin* (Family: Anacardiaceae); *Annona muricata*, *Annona reticulata* and *Annona squamosa* (Family:

Annonaceae); *Asclepias curassavica* (Family: Apocynaceae); *Pistia stratiotes* (Family: Araceae), *Bidens pilosa*, *Eclipta prostrata* (Family: Asteraceae); *Crescentia cujete* and *Tabebuia rosea* (Family: Bignoniaceae); *Bixa Orellana* (Family: Bixaceae); *Cordia alba* (Family: Boraginaceae); *Bursera graveolens* (Family: Burseraceae); *Evolvulus alsinoides* (Family: Convolvulaceae); *Euphorbia hirta*, *Euphorbia thymifolia* and *Hura crepitans* (Family: Euphorbiaceae); *Abrus precatorius*, *Acacia pennatula* and *Gliricidia sepium* (Family: Fabaceae); *Ocimum campechianum* (Family: Lamiaceae); *Spigelia anthelmia* (Family: Loganiaceae); *Sida acuta*, *Sida glomerata* and *Waltheria indica* (Family: Malvaceae); *Cissampelos pareira* (Family: Menispermaceae); *Ficus insípida* and *Ficus máxima* (Family: Moraceae); *Psidium guajava* (Family: Myrtaceae); *Oxalis corniculata* (Family: Oxalidaceae); *Argemone mexicana* (Family: Papaveraceae); *Phyllanthus niruri* (Family: Phyllanthaceae); *Scoparia dulcis* (Family: Plantaginaceae); *Portulaca oleracea* (Family: Portulacaceae); *Pityrogramma calomelanos* (Family: Pteridaceae), *Paullinia pinnata* and *Sapindus saponaria* (Family: Sapindaceae); *Manilkara zapota* (Family: Sapotaceae); *Cecropia peltata* (Family: Urticaceae) and *Lantana camara* (Family: Verbenaceae). In summary, 45 native plants with antihelminthic activity were found; the families with more than one plant were Amaranthaceae (4), Annonaceae (3), Euphorbiaceae (3), Fabaceae (3), Asteraceae (2), Bignoniaceae (2), Malvaceae (3), Moraceae (2) and Sapindaceae (2).

The following are the naturalized plants identified in the Tolima TDF with antihelmintic activity: *Calotropis procera* (Family: Apocynaceae); *Heliotropium indicum* (Family: Boraginaceae), *Momordica charantia* (Family: Cucurbitaceae); *Cyperus rotundus* (Family: Cyperaceae); *Senna occidentalis* (Family: Fabaceae) and *Cynodon dactylon* (Family: Poaceae). 6 naturalized plants, which belong to different families, with reports of antihelminthic activity were found.

Furthermore, the following are the exotic plants identified in the Tolima TDF with antihelmintic activity: *Anacardium occidentale*, *Mangifera indica* (Anacardiaceae); *Cascabela thevetia* (Family: Apocynaceae); *Cocos nucifera* (Family: Arecaceae); *Ananas comosus* (Family: Bromeliaceae), *Carica papaya* (Family: Caricaceae); *Bauhinia variegata*, *Caesalpinia pulcherrima*, *Cajanus cajan*, *Cassia fistula*, *Crotalaria retusa*, *Delonix regia*, *Leucaena leucocephala*, *Pithecellobium dulce* and *Tamarindus indica* (Family: Fabaceae); *Azadirachta indica* and *Melia azedarach* (Family: Meliaceae); *Mirabilis jalapa* (Family: Nyctaginaceae), *Cenchrus ciliaris* (Family: Poaceae) y *Antigonon leptopus* (Family: Polygalaceae). In summary, 20 plants were found and the families with the highest participation were Fabaceae (9), Anacardiaceae (2) and Meliaceae (2).

Main Chemical Groups in Vegetal Species: The systematic review about the content of secondary metabolites in native species shows that tannins, flavonoids, alkaloids, and phenols are the groups with the highest levels percentage in these vegetal species, while terpenoids, saponins and steroids are in a medium level (Edeoga *et al.*, 2005; Okoli *et al.*, 2009; Kumar *et al.*, 2010; Igwe *et al.*, 2010; Jayaveera *et al.*, 2010;

Valdés *et al.*, 2010; Jhade *et al.*, 2011; Bhalke & Chavan, 2011; Padhi *et al.*, 2011; Omogbai & Eze, 2011; Basma *et al.*, 2011; Olajuyigbe *et al.*, 2011; Nagarajan *et al.*, 2011; Savithramma *et al.*, 2011; Lor *et al.*, 2011; Antara, 2012; Accioly *et al.*, 2012; George *et al.*, 2012; Agrawal *et al.*, 2012; Kyei *et al.*, 2012; Akharaiyi *et al.*, 2012; Ahmed *et al.*, 2013; Souza *et al.*, 2013; Maobe *et al.*, 2013; John *et al.*, 2013; Joselin *et al.*, 2013; Deshmukh *et al.*, 2013; Gnanaraja *et al.*, 2013; Sinha, 2013; Sharmila *et al.*, 2013; Beltrán *et al.*, 2013; Naz & Bano, 2013; Kavitha *et al.*, 2014; Pizano & García, 2014; Asif *et al.*, 2014; Mondal *et al.*, 2014; Gavamukulya *et al.*, 2014; Jayanthi *et al.*, 2014; Yang, 2014; Bhuvaneswari *et al.*, 2014; Hussain *et al.*, 2014; Atif *et al.*, 2014; Veni & Pushpanathan, 2014; Shanmugam *et al.*, 2014; Rajesh *et al.*, 2014; Jamkhande & Wattamwar, 2015; Saleem & Devi, 2015; Elufioye & Olaifa, 2015; Khanal *et al.*, 2015; Gopi, 2015; Adeyemi, 2016; Rajalakshmi *et al.*, 2016; Xuan & Khanh, 2016; Granja, *et al.*, 2016; Cruz & Cui-Lim, 2016; Kumar & Simon, 2016; Sukalingam, *et al.*, 2017; Kota, *et al.*, 2017; Rajeswari & Vijayashalini, 2017; Tang *et al.*, 2017; Ahuchaogu *et al.*, 2017; Agu & Okolie, 2017; Ma, *et al.*, 2017; Sri *et al.*, 2017; Ribeiro *et al.*, 2017; Paul *et al.*, 2017).

With regard to naturalized species, the systematic review on the content of secondary metabolites found that tannins, flavonoids, alkaloids, and phenols had the highest compositional percentage followed, at a medium level by steroids, saponins and terpenoids (Fig. 1) (Saidu *et al.*, 2011; Joshi & Kaur, 2013; Adeniyi *et al.*, 2013; Mohammad *et al.*, 2014; Chetia *et al.*, 2014; Abdillah *et al.*, 2015; Odeja *et al.*, 2015; Al-Snafi, 2016; Nandagoapalan *et al.*, 2016; Hassan *et al.*, 2017; Akindele *et al.*, 2017; Katiyar *et al.*, 2017; Kasarkar *et al.*, 2017).

In the analysis of antihelmintic activity of foreign plants, the systematization of the qualitative composition of the species shows that tannins, phenols, alkaloids, saponins and flavonoids are the metabolites with the highest analysis, followed at a medium level by terpenoids and steroids (Ademola *et al.*, 2005; Mali *et al.*, 2008; Bouzada *et al.*, 2009; Sivasankari *et al.*, 2010; Kumar *et al.*, 2010; Bhadriya *et al.*, 2011; Antonisamy *et al.*, 2012; Kumar *et al.*, 2013; Susmitha *et al.*, 2013; Sultana *et al.*, 2013; Kalpana *et al.*, 2014; Doraiswamy *et al.*, 2014; Agrawal *et al.*, 2014; Olabinri *et al.*, 2014; Mustapha *et al.*, 2015; Koffi *et al.*, 2015; Lima *et al.*, 2015; Roopan, 2016; Monji *et al.*, 2016; Anju & Zachariah, 2016; Kousalya & Jayanthi, 2016; Modi *et al.*, 2016; Adebayo *et al.*, 2016; Chan *et al.*, 2017; Wilberforce & Olivia, 2017; Anadebe *et al.*, 2017; Sujatha *et al.*, 2017; Yadav *et al.*, 2017; Kumari, 2017; Yadima *et al.*, 2017; Cheenickal & Mendez, 2017; Hanani *et al.*, 2017; Arora *et al.*, 2017; Sravanti *et al.*, 2017).

By jointly systematizing the content of metabolites in the three groups of plants, the metabolites with the highest levels were tannins, flavonoids, and alkaloids, while phenols, saponins, steroids and terpenoids are located at a medium level (Fig. 2). A series of studies are available for tannins, alkaloids, and flavonoids antihelminthic activity (Anthnasiadou *et al.*, 2001; Da Silva *et al.*, 2008; Kozan *et al.*, 2013). The mode of action that supports the antihelminthic effect of tannins has been discussed, but remains unidentified (Engström *et al.*, 2016). It has been suggested that the direct effect of plants is based on their interactions with vital egg proteins and larvae for biological function and development (Molan, 2014).

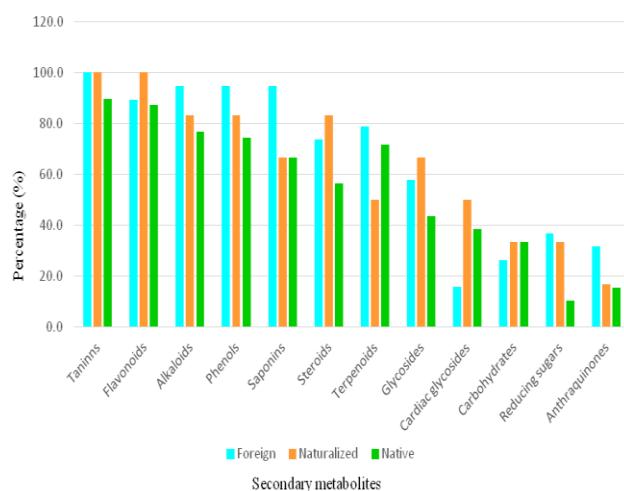


Fig. 1. Percentage content of metabolites in native, exotic, and naturalized plants with anthelmintic activity.

Table 1. Native plants with flavonoids, tannins, and alkaloids content.

Species	Family
<i>Trianthema portulacastrum</i>	Aizoaceae
<i>Amaranthus viridis</i>	Amaranthaceae
<i>Spondias mombin</i>	Anacardiaceae
<i>Annona muricata</i>	Annonaceae
<i>Annona reticulata</i>	
<i>Bidens pilosa</i>	Asteraceae
<i>Euphorbia thymifolia</i>	Euphorbiaceae
<i>Hura crepitans</i>	
<i>Gliricidia sepium</i>	Fabaceae
<i>Spigelia anthelmia</i>	Loganiaceae
<i>Sida acuta</i>	
<i>Waltheria indica</i>	
<i>Psidium guajava</i>	Myrtaceae
<i>Oxalis corniculata</i>	Oxalidaceae
<i>Argemone mexicana</i>	Papaveraceae
<i>Phyllanthus niruri</i>	Phyllanthaceae
<i>Portulaca oleracea</i>	Portulacaceae
<i>Paullinia pinnata</i>	Sapindaceae
<i>Cecropia peltata</i>	Urticaceae

In this regard, it has been reported that condensed tannins show anthelmintic activity. Inhibition of the larval viability of *Haemonchus contortus*, *Teladorsagia circumcincta* and *Trichostrongylus vitrinus* was observed when these species were subjected to the action of *Schinopsis spp.* whose extract contained 73% condensed tannins (Hoste *et al.*, 2006). The same extract reduced the burden of sheep eggs and worms with monospecific infection of *H. contortus* (Max *et al.*, 2007). Flavonoids belong to the largest group of secondary metabolites (> 6,000 identified) and widely distributed (Hoste *et al.*, 2006; Max *et al.*, 2007) found in almost all photosynthetic plants. A study (Kozan *et al.*, 2013) showed for the first time that flavonoid glycosides (luteolin-7-β-O-glucopyranoside and quercetin-3-O-β-glucopyranoside) of the acetic extract of *Vicia pannonica* Crantz. var. *Purpurascens* possess a significant *In vitro* anthelmintic activity against *Trichostrongylus sp.*. There is also evidence on the activity of luteolin and quercetin in combination with condensed tannins on the molt of L₃ larvae of *H. contortus* (Klongsiriwet *et al.*, 2015).

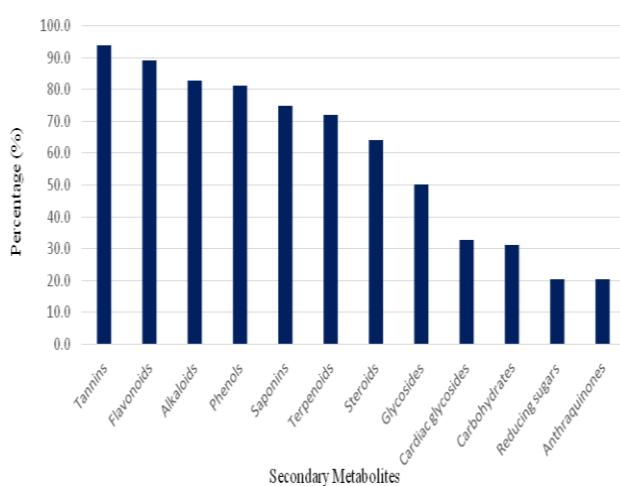


Fig. 2. Percentage consolidated content of metabolites in native, exotic, and naturalized plants with anthelmintic activity.

Table 2. Naturalized plants with flavonoids, tannins and alkaloids content.

Scientific name	Family
<i>Calotropis procera</i>	Apocynaceae
<i>Momordica charantia</i>	Cucurbitaceae
<i>Senna occidentalis</i>	Fabaceae

Table 3. Exotic plants with flavonoids, tannins, and alkaloids content.

Scientific name	Family
<i>Mangifera indica</i>	Anacardiaceae
<i>Cocos nucifera</i>	Arecaceae
<i>Ananas comosus</i>	Bromeliaceae
<i>Carica papaya</i>	Caricaceae
<i>Bauhinia variegata</i>	
<i>Cassia fistula</i>	
<i>Crotalaria retusa</i>	
<i>Leucaena leucocephala</i>	Fabaceae
<i>Pithecellobium dulce</i>	
<i>Tamarindus indica</i>	
<i>Melia azedarach</i>	Meliaceae
<i>Mirabilis jalapa</i>	Nyctaginaceae

Regarding alkaloids, it has been reported that some isolated alkaloids were highly toxic to HL60 cells in tissue culture, such that those with a high RM (relative mobility) 50/IC (inhibitory concentration) 50 (selectivity index: SI > 100) were proposed as anthelmintic alkaloid molecules such as alocryptopine, dehydrocordalina and papaverine. However, the inhibition mechanism of larval motility is unclear so far (Hrkova & Velebný, 2012). Other researchers have found antihelminthic activity with molecules from this group such as dicentrin, isoquinoline alkaloid that showed biological activity *In vitro* in larvae of *H. contortus* (Ayers *et al.*, 2007). The extract of *Spigelia anthelmia* showed strong anthelmintic activity against *H. contortus* and the chemical study of this extract revealed as one of the main constituents of spigantine alkaloid (Morais *et al.*, 2002).

Finally, as a result of applying the filter to native plants with anthelmintic activity, which simultaneously contain flavonoids, tannins and alkaloids, a total of 19

plants were obtained, which are listed in (Table 1). For the naturalized plants, only 3 plants were obtained (Table 2) and for the exotic plants the same filter yielded 12 plants (Table 3).

Plants with anthelmintic activity used as food source:

A total of 5 plants were obtained of which 2 native (*Gliricidia sepium* and *Psidium guajava*), 1 naturalized (*Calotropis procera*), and 2 exotic (*Leucaena leucocephala* and sweet *Pithecellobium*) plants have been reported as being used in animal feeding activities. For the specific case of sheep, *Leucaena leucocephala*, *Gliricidia sepium*, *Calotropis procera* and sweet *Pithecellobium* species have also been evaluated as a food source (De Azevêdo *et al.*, 2010; Archimède *et al.*, 2010; Díaz *et al.*, 2013; Andrade *et al.*, 2014).

Conclusions

A considerable number of Tolima TDF plants, among native, naturalized and introduced plants, have anthelmintic activity. The families with the highest number of plants shown in this report are Fabaceae, Amaranthaceae, Annonaceae and Euphorbiaceae.

The phytochemical profile of the plants with scientific anthelmintic activity shows that tannins, flavonoids, and alkaloids are secondary, predominant metabolites in these vegetal species.

Although antihelminthic activity of the three metabolite groups has been found, greater knowledge and understanding of the mechanisms of action involved is required.

The following are the plants with anthelmintic activity used in animal feed: *Gliricidia sepium*, *Psidium guajava*, *Calotropis procera*, *Leucaena leucocephala* and *Pithecellobium dulce*, constituting species with potential nutraceutical application in sheep.

References

- Abdillah, S., R.M. Tambunan, Y. Farida, N.M.D. Sandhiutami and R.M. Dewi. 2015. Phytochemical screening and antimalarial activity of some plants traditionally used in indonesia. *Asian Pacific J. Trop. Disease*, 5(6): 454-457.
- Accioly, M.P., C.M.L. Bevilaqua, F.C. Rondon, S.M. Morais, L.K. Machado, C.A. Almeida and R.P. Cardoso. 2012. Leishmanicidal activity *In vitro* of *Musa paradisiaca* L. and *Spondias mombin* L. fractions. *Vet. Parasitol.*, 187(1): 79-84.
- Adebayo, M.R., S.K. Babatunde, A.E. Ajiboye and L.M. Habeeb. 2016. Antimycotic and phytochemical screening of the fruit pulp extract of Tamarind (*Tamarindus indica*) on *Candida albicans*. *J. Microbiol. & Antimicrobial Agents*, 2(1): 16-21.
- Ademola, I.O., A.I. Akanbi and S.O. Idowu. 2005. Comparative nematocidal activity of chromatographic fractions of *leucaena leucocephala* seed against gastrointestinal sheep nematodes. *Pharm. Biol.*, 43(7): 599-604.
- Adeniyi, T.A., P.A. Adeonipekun and A.E. Omotayo. 2013. Investigating the phytochemicals and antimicrobial properties of three sedge (cyperaceae) species. *Int. J. Trop. Med.*, 8(4): 92-98.
- Adeyemi, D.K. 2016. Phytochemical screening and *In vitro* evaluation of free radical scavenging activity of *Pistia stratiotes* extracts. *Asian J. Biomed. & Pharm. Sci.*, 6(53): 08.
- Agrawal, K., A. Joshi, S. Ghildiyal, M.K. Gautam, M. Gangwar, R.K. Goel and V.K. Joshi. 2014. Qualitative phytochemical and physicochemical analysis of cassia fistula L. fruit. *Med. Plants*, 6(2): 138-142.
- Agrawal, M., Y. Agrawal, P. Itankar, A. Patil, J. Vyas and A. Kelkar. 2012. Phytochemical and HPTLC studies of various extracts of *Annona squamosa* (Annonaceae). *Int. J. Pharm. Tech. Res.*, 4(1): 364-368.
- Agu, K.C. and P.N. Okolie. 2017. Proximate composition, phytochemical analysis, and *In vitro* antioxidant potentials of extracts of *Annona muricata* (Soursop). *Food Sci. & Nutr.*, 5(5): 1029-1036.
- Ahmed, S.A., S. Hanif and T. Iftekhar. 2013. Phytochemical profiling with antioxidant and antimicrobial screening of *Amaranthus viridis* L. leaf and seed extracts. *Open J. Med. Microb.*, 3: 164-171.
- Ahuchaogu, A.A., A.I. Obike, C.S. Egedeuzu, P.O. Ukaogo and O.J. Chukwu. 2017. Phytochemical screening and antibacterial activity of *Spondias mombin* leaves. *Asian J. Chem. Sci.*, 3(1): 1-6.
- Akharaiyi, F.C., B. Boboye and F.C. Adetuyi. 2012. Antibacterial, phytochemical and antioxidant activities of the leaf extracts of *gliricidia sepium* and *spathodea campanulata*. *World App. Sci. J.*, 16(4): 523-530.
- Akindele, P.O., O.A. Fatunla, K.A. Ibrahim and C.O. Afolayan. 2017. Antibacterial and phytochemical screening of *calotropis procera* leaf extracts against vancomycin and methicillin resistant bacteria isolated from wound samples in hospital patients. *J. Compl. & Alternat. Med. Res.*, 2(1): 1-14.
- Al-Snafi, A.E. 2016. Chemical constituents and pharmacological effects of *Cynodon dactylon*-A review. *IOSR J. Pharm.*, 6(7): 17-31.
- Anadebe, V.C., N.A. Okafor, J.O. Ezeugo, I.J. Amanjide and B.C. Ogide. 2017. GC-MS Analysis of phytochemical compounds in cajanus cajan leaf. *J. Chem. & Pharm. Res.*, 9(5): 360-363.
- Andrade, I.R.A., M.J.D. Cândido, R.C.F. Pompeu, V.P. Guimarães, L.V. Silva and M.E.S. Evangelista. 2014. Productive and economic performance of feedlot sheep using different protein sources in concentrate food. Desempenho produtivo e econômico do confinamento de ovinos utilizando diferentes fontes proteicas na ração concentrada. *Revista Brasileira De Saude e Producao Animal*, 15(3): 717-730.
- Anju, V. and S.M. Zachariah. 2016. Phytochemical screening, isolation, antibacterial and anticancer activity studies of *Caesalpina pulcherrima* Linn leaves by HPTLC analysis. *Int. J. Pharm. & Biol. Sci.*, 8(2): 12-29.
- Antara, C. 2012. Evaluation of physiochemical and phytochemical parameters of amaranthus spinosus leaves. *Int. J. Pharm.*, 3(10): 210-211.
- Anthnasiadou, S., I. Kyriazakis, F. Jackson and R.L. Coop. 2001. Direct anthelmintic effects of condensed tannins towards different gastrointestinal nematodes of sheep: *In vitro* and *In vivo* studies. *Vet. Parasitol.*, 99: 205-219.
- Antonisamy, J.M., J.S. Aparna, S. Jeeva, S. Sukumaran and B. Anantham. 2012. Preliminary phytochemical studies on the methanolic flower extracts of some selected medicinal plants from india. *Asian Pacific J. Trop. Biomed.*, 2(1 SUPPL.): S79-S82.
- Archimède, H., E. González, P. Despois, T. Etienne and G. Alexandre. 2010. Substitution of corn and soybean with green banana fruits and *Gliricidia sepium* forage in sheep fed hay-based diets: Effects on intake, digestion and growth. *J. Ani. Physiol. & Ani. Nutr.*, 94(1): 118-128.
- Arora, S., G. Kumar and S. Meena. 2017. Screening and evaluation of bioactive components of *Cenchrus ciliaris* L. by GC-MS analysis. *Int. Res. J. Pharm.*, 8(6): 69-76.

- Asif, M., M. Atif, A.S.A. Malik, Z.C. Dan, I. Ahmad and A. Ahmad. 2014. Diuretic activity of *Trianthema portulacastrum* crude extract in albino rats. *Trop. J. Pharm. Res.*, 12(6): 967-972.
- Atif, M., M. Azharuddin, S.A. Rahman, M.I. Ahmed and S.B. Mahmood. 2014. Evaluation of anticataract potential of *Waltheria indica* in albino rats. *Asian J. Plant Sci. & Res.*, 4(6): 52-58.
- Ayers, S., D.L. Zink, K. Mohn, J.S. Powell, C.M. Brown, T. Murphy and S.B. Singh. 2007. Anthelmintic activity of aporphine alkaloids from *Cissampelos capensis*. *Planta Med.*, 73(3): 296-297.
- Basma, A.A., Z. Zakaria, L.Y. Latha and S. Sasidharan. 2011. Antioxidant activity and phytochemical screening of the methanol extracts of *Euphorbia hirta* L. *Asian Pacific J. Trop. Med.*, 4(5): 386-390.
- Beltrán, C.E., F. Díaz and H. Gómez. 2013. Tamizaje fitoquímico preliminar de especies de plantas promisorias de la costa atlántica colombiana. *Rev. Cub de Plant. Med.*, 18(4): 619-631.
- Bhadoriya, S.S., V. Uplanchiwar, V. Mishra, A. Ganeshpurkar, S. Raut and S.K. Jain. 2011. *In vitro* anthelmintic and antimicrobial potential of flavonoid rich fraction from tamarindus indica seed coat. *Pharmacologyonline*, 3: 412-420.
- Bhalke, R.D and M.J. Chavan. 2011. Analgesic and CNS depressant activities of extracts of *Annona reticulata* Linn Bark. *Phytopharmacol.*, 1(5): 160-165.
- Bhuvaneswari, S., S. Deepa, N. Sripriya, L. Prameela and P.N.K. Udaya. 2014. Antioxidant activity and phytochemistry of various flowers from Tamil Nadu, India. *Int. J. Res. Pharm.*, 5(1): 40-45.
- Bouzada, M.L.M., R.L. Fabri, M. Nogueira, T.U.P. Konno, G.G. Duarte and E. Scio. 2009. Antibacterial, cytotoxic and phytochemical screening of some traditional medicinal plants in brazil. *Pharm. Biol.*, 47(1): 44-52.
- Chan, E.W.C., S. Baba, H.T. Chan, M. Kainuma, T. Inoue and S.K. Wong. 2017. Ulam herbs: A review on the medicinal properties of *Anacardium occidentale* and *Barringtonia racemosa*. *J. Appl. Pharm. Sci.*, 7(02): 241-247.
- Cheenickal, M. and R.M. Mendez. 2017. Phytochemical screening and the antimicrobial activity of leaves of *Azadirachta indica*, A. Juss. *Int. J. Sci. & Engin. Res.*, 8(5): 721-724.
- Chetia, J., S. Upadhyaya and D.K. Bora. 2014. Screening of phytochemicals, antioxidant and antimicrobial activity of some tea garden weeds. *Int. J. Pharm. Sci. Review and Res.*, 26(1): 193-196.
- Cruz, M.G.M. and K.M.R. Cui-Lim. 2016. Phytochemical screening of the ethanol extract of (Jacq.) Steud *Gliricidia sepium* (Kakawate). *Asian J. Pharm. and Pharmacol.*, 2(6): 150-153.
- Da Silva, V.C., M.G. Carvalho and H.R. Borba. 2008. Anthelmintic activity of flavonoids isolated from roots of *Andira anthelmia* (Leguminosae). *Rev. Bras. Farmacog.*, 18(4): 573-576.
- De Azevêdo Silva, A.M., R.G. Da Costa, J.M. Pereira, I.A. Bakke, L.K. Da Silva, G.E. Lira and G.H. Da Nóbrega. 2010. Nutritional value of silk flower hay for lambs. *Revista Brasileira De Zootecnia* 39(12): 2739-2743.
- Deshmukh, S.N., B. Shrivastava, P. Sharma, H.K. Jain and N. Ganesh. 2013. Pharmacognostic standardization, physico and phytochemical evaluation of bixa orellana linn. seed. *Int. J. Pharm. Sci. Review & Res.*, 23(1): 302-305.
- Díaz, B., A. Elías and E.C. Valiño. 2013. Nutritional and economical efficiency of three biosilages from agroindustrial wastes in beef cattle. *Cuban J. Agri. Sci.*, 47(2): 143-150.
- Doraiswamy, H., N. Kathavarayan, R.C. Sharma and M.V. Krishna. 2014. Molecular docking analysis of secondary metabolites of *trigonella foenum graecum* and *carica papaya* with FTO: An insilico approach. *Int. J. Pharm. Sci. Review & Res.*, 27(1): 105-110.
- Edeoga, H.O., D.E. Okwu and B.O. Mbaebie. 2005. Phytochemical constituents of some Nigerian medicinal plants. *Afr. J. Biotechnol.*, 4(7): 685-688.
- Elufioye, O.T. and A.O. Olaifa. 2015. Pharmacognostic evaluation of *Spigelia anthelmia* Linn (Loganiaceae). *Europ. J. Med. Plants* 8(2): 87-96.
- Engström, M.T., M. Karonen, J.R. Ahern, N. Baert, B. Payré, H. Hoste and J.P. Salminen. 2016. Chemical structures of plant hydrolyzable tannins reveal their *In vitro* activity against egg hatching and motility of *Haemonchus contortus* nematodes. *J. Agri. & Food Chem.*, 3; 64(4): 840-51.
- Gavamukulya, Y., F. Abou-Elella, F. Wamunyokoli and H. AEI-Shemy. 2014. Phytochemical screening, anti-oxidant activity and *In vitro* anticancer potential of ethanolic and water leaves extracts of *annonia muricata* (graviola). *Asian Pacific J. Trop. Med.*, 7(S1): S355-S363.
- George, V.C., D.R. Kumar, V. Rajkumar, P.K. Suresh and R.A. Kumar. 2012. Quantitative analysis of the relative antineoplastic potential of the n-butanolic leaf extract of *Annona muricata* Linn in normal and immortalized human cell lines. *Asi. Pacif. J. Cancer Preven.*, 13(2): 699-704.
- Gnanaraja, R., V. Prakash, S. Peter and M. Mahendraverman. 2013. Qualitative and quantitative phytochemicals analysis of selected fabaceae medicinal plants from Allahabad region. *The Pharm. Innov. J.*, 3(7): 53-56.
- Gopi, S. 2015. Phytochemical screening, *In vitro* antioxidant and cytotoxic studies on *Waltheria indica* Linn. *Int. J. Curr. Res.*, 7(1): 11834-11843.
- Granja, F.G., A.P. de Oliveira, C.M.S. Souza de Castro, R.G. de Oliveira, I.M. Melo, X. Pereira, D. Marrero and J.R. Guedes da Silva. 2016. Phytochemical screening and antioxidant activity of methanolic fraction from the leaves of *Crescentia cujete* L. (Bignoniaceae). *J. Chem. Pharm. Res.*, 8(2): 231-236.
- Hanani, E., R. Prastiwi and L. Karlina. 2017. Indonesian *Mirabilis jalapa* Linn: A pharmacognostical and preliminary phytochemical investigations. *Pharm. J.*, 9(5): 683-688.
- Hassan, M.H.A., M.A. Ismail, A.M. Moharran and A.A. Shoreit. 2017. Phytochemical and antimicrobial of latex serum of *calotropis procera* and its silver nanoparticles against some reference pathogenic strains. *J. Ecol. Health & Environ.*, 5(3): 65-75.
- Hoste, H., F. Jackson, S. Athanasiadou, S.M. Thamsborg and S.O. Hoskin. 2006. The effects of tannin-rich plants on parasitic nematodes in ruminants. *Trends in Parasitol.*, 22: 253-261.
- Hrkova, G. and S. Velebny. 2012. Pharmacological potential of selected natural compounds in the control of parasitic diseases. *Springer Science & Business Media*.
- Hussain, M., U. Farooq, M. Rashid, H. Bakhsh, A. Majeed, I.A. Khan and A. Aziz. 2014. Antimicrobial activity of fresh latex juice and extract of *Euphorbia hirta* and *Euphorbia thymifolia*: An *In vitro* comparative study. *Int. J. Pharm. Sci.*, 4(3): 546-553.
- Igwe, C.U., G.O.C. Onyeze, V.A. Onwuliri, C.G. Osuagwu and A.O. Ojiako. 2010. Evaluation of the chemical compositions of the leaf of *Spondias mombin* Linn from Nigeria. *Aust. J. Basic & App. Sci.*, 4(5): 706-710.
- Jamkhande, P.G. and A.S. Wattamwar. 2015. *Annona reticulata* Linn. (bullock's heart): Plant profile, phytochemistry and pharmacological properties. *J. Trad. & Complement. Med.*, 5(3): 144-152.

- Jayanthi, A., P. Chakraborty, A. Mathew and K. Khare. 2014. Cytotoxicity and antimicrobial effects of *Pistia stratiotes* leaves. *Int. J. Drug Develop. & Res.*, 6(4): 208-217.
- Jayaveera, K.N., R.K. Yoganandham, Y. Govindarajula and R. Kumanan. 2010. Phytochemical screenings, antibacterial activity and physico chemical constants of ethanolic extract of *Euphorbia thymifolia* Linn. *Int. J. Pharm. & Pharm. Sci.*, 2(3): 81-82.
- Jhade, D., D. Ahirwar, R. Jain, N. Sharma and S. Gupta. 2011. Pharmacognostic standardization, physico- and phytochemical evaluation of *Amaranthus spinosus* Linn. *Root. J. Young Phar.*, 3(3): 221-225.
- John, N.R., V.C. Gala, A.M. Bhagwat and C. Sawant. 2013. HPTLC analysis of *Eclipta prostrata* and *Psidium guajava* extracts and their effect on cell-surface hydrophobicity of a consortium of dental plaque isolates. *Int. J. Pharm. & Pharm. Sci.*, 5(3): 935-940.
- Joselin, J., T.S. Shynin, A. Rajam and S. Jeeva. 2013. Phytochemical evaluation of Bignoniacae flowers. *J. Chem. & Pharm. Res.*, 5(4): 106-111.
- Joshi, M and S. Kaur. 2013. *In vitro* evaluation of antimicrobial activity and phytochemical analysis of calotropis procera, eichhornia crassipes and datura innoxia leaves. *Asian J. Pharm. & Clin. Res.*, 6(SUPPL.5): 25-28.
- Kalpana, M.B., P.G. Sriram and S. Subramanian. 2014. Studies on the antidiabetic activity of ananas comosus leaves in STZ induced diabetic rats. *Der Pharm. Lett.*, 6(2): 22-30.
- Kasarkar, A.R., M.S. Thakur, S.S. Chougale, S.S. Wadkar, S.B. Jadhav, S.A. Patil and S.B. Kubal. 2017. Preliminary phytochemical investigation of the leaves of *Cynodon dactylon* (L.) Pers., *Cyperus rotundus* L. and *Typha angustifolia* L. *J. Med. Plants*, 5(4): 203-205.
- Katiyar, D., V. Singh and Ali. Mohd. 2017. Phytochemical and pharmacological profile of *Momordica charantia*: A review. *Biochem. and Therap. Uses of Med. Plants*, 1-33.
- Kavitha, D., R. Parvatham and P.R. Padma. 2014. Assessment of trianthema portulacastrum for its antimicrobial potential and investigation of their phytochemicals using HPTLC, GC-MS, and IR. *Int. J. Pharm. and Pharm. Sci.*, 6(1): 675-686.
- Khanal, D.P., B. Raut and K.S. Dangol. 2015. Phytochemical screening, pharmacognostic evaluation and biological activity of *Amaranthus spinosus* L. *J. Manmohan Mem. Inst. Health Sci.*, 1(4): 29-34.
- Klongsiriwet, C., J. Quijada, A.R. Williams, I. Mueller-Harvey, E.M. Williamson and H. Hoste. 2015. Synergistic inhibition of *Haemonchus contortus* exsheathment by flavonoid monomers and condensed tannins. *Int. J. for Parasit. Drugs & Drug Resist.*, 5(3): 127-134.
- Koffi, A.J., K.B. Bla, H.F. Yapi, A.P. Bidie and A.J. Djaman. 2015. Phytochemical screening of some medicinal plants in côte D'ivoire and evaluation of their extraction efficiency. *Int. J. Pharm. & Phytochem. Res.*, 7(3): 563-569.
- Kota, S., V.R. Govada, R.K. Anantha and M.K. Verma. 2017. An investigation into phytochemical constituents, antioxidant, antibacterial and anti-cataract activity of *Alternanthera sessilis*, a predominant wild leafy vegetable of South India. *Biocatal. & Agri Biotechnol.*, 10: 197-203.
- Kousalya, P and V. Jayanthi. 2016. Evaluation of phytochemicals and quantification of phenol, flavonoids and tannins of pods of *Leucaena leucocephala* (Lam.) De Wit. *Amer. Eura. J. Agri. & Environ. Sci.*, 16 (9): 1561-1564.
- Kozan, E., S.A. Anul and I.I. Tatli. 2013. *In vitro* anthelmintic effect of *Vicia pannonica* var. *purpurascens* on trichostrongylosis in sheep. *Exp. Parasit.*, 134(3): 299-303.
- Kumar, A., K. Lakshman, K.N. Jayaveera, S.D. Shekar and C. Vivek. 2010. Antinociceptive and antipyretic activities of *Amaranthus viridis* Linn in different experimental models. *Arch. Biol. Sci.*, 62(2): 397-402.
- Kumar, M., K. Nehra and J.S. Duhan. 2013. Phytochemical analysis and antimicrobial efficacy of leaf extracts of *pithecellobium dulce*. *Asian J. Pharm. & Clin. Res.*, 6(1): 70-76.
- Kumar, N.S. and N. Simon. 2016. *In vitro* antibacterial activity and phytochemical analysis of *Glicicidia sepium* (L.) leaf extracts. *J. Pharm. & Phytochem.*, 5(2): 131.
- Kumar, V.K., N.R. Sankar, S. Ramya, R.V. Sahaja, K. Saritha, K.G. Reddy and N.V. Naidu. 2010. Phytochemical screening and antimicrobial activity of the leaf extract of *mirabilis jalapa* against pathogenic microorganisms. *Int. J. Phytomed.*, 2(4): 402-407.
- Kumari, S. 2017. Evaluation of phytochemical analysis and antioxidant and antifungal activity of *Pithecellobium dulce* leaves extract. *Asian J. Pharm. & Clin. Res.*, 10(1): 370-375.
- Kyei, S., G.A. Koffuor and J.N. Boampong. 2012. Antiarthritic effect of aqueous and ethanolic leaf extracts of *Pistia stratiotes* in adjuvant-induced arthritis in Sprague-Dawley rats. *J. Exp. Pharm.*, 4: 41-51.
- Lima, E.B.C., C.N.S. Sousa, L.N. Meneses, N.C. Ximenes, J.M.A. Santos, G.S. Vasconcelos and S.M.M. Vasconcelos. 2015. *Cocos nucifera* (L.) (Arecaceae): A phytochemical and pharmacological review. *Braz. J. Med. & Biol. Res.*, 48(11): 953-964.
- Lor, L.D., M.O. Uguru, P.N. Olotu, T.L. Ohemu and A. Ukpere. 2011. Evaluation of analgesic and anti-inflammatory activities and phytochemical screening of the leaves extract of *Paullinia pinnata* (Sapindaceae). *J. Chem. & Pharm. Res.*, 3(4): 351-356.
- Mali, R.G., S.G. Mahajan and A.A. Mehta. 2008. Evaluations of *bauhinia variegata* linn stem bark for anthelmintic and antimicrobial properties. *J. Nat. Remed.*, 8(1): 39-43.
- Maobe, M.A., E. Gatebe, L. Gitu and H. Rotich. 2013. Preliminary phytochemical screening of eight selected medicinal herbs used for the treatment of diabetes, malaria and pneumonia in Kisii region, southwest Kenya. *Eur. J. App. Sci.*, 5(10): 01-06.
- Max, R.A., J. Dawson, D. Wakelin, P. Butterly, A. Kimambo, A., Kassuku and L. Mtenga. 2002. L. Effect of condensed tannin extract on gastrointestinal nematodes of small ruminants. *Sokoine Univer. Agri.*, 43-56.
- Modi, A., V. Mishra, A. Bhatt, A. Jain, M.H. Mansoori, E. Gurnany and V. Kumar. 2016. *Delonix regia*: Historic perspectives and modern phytochemical and pharmacological researches. *Chin. J. Nat. Med.*, 14(1): 31-39.
- Mohammad, S.A., S.A. Nabi, S. Marella, K.T. Thandaiah, M.V. Jyothi and C.A. Rao. 2014. Phytochemical screening and antihyperglycemic activity of *heliotropium indicum* whole plant in streptozotocin induced diabetic rats. *J. Appl. Pharm. Sci.*, 4(12): 065-071.
- Molan, A.L. 2014. Effect of purified condensed tannins from pine bark on larval motility, egg hatching and larval development of *Teladorsagia circumcincta* and *Trichostrongylus colubriformis* (Nematoda: Trichostrongylidae). *Folia Parasitologica*, 61(4): 371.
- Mondal, H., H. Hossain, K. Awang, S. Saha, S. Mamun-Ur-Rashid, M.K. Islam and J.A. Shilpi. 2014. Anthelmintic activity of ellagic acid, a major constituent of *Alternanthera sessilis* against *Haemonchus contortus*. *Pak. Vet. J.*, 35(1): 58-62.
- Monji, F., P.G. Adaikan, L.C. Lau, B.B. Said, Y. Gong, H.M. Tan and M. Choolani. 2016. Investigation of uterotonic properties of *Ananas comosus* extracts. *J. Ethnopharm.*, 193: 21-29.

- Morais, S.M.D., C.M.L. Beviláqua, J.A. Souza and L.M.D. Assis. 2002. Chemical investigation of *Spigelia anthelmia* Linn. used in Brazilian folk medicine as antihelminthic. *Revista Brasil. de Farmacogn.*, 12: 81-82.
- Mustapha, A.A., G. Owuna, J.O. Ogaji, U. Is-Haq Is-Haq and M.M. Idris. 2015. Phytochemical screening and inhibitory activities of anacardium occidentale leave extracts against some clinically important bacterial isolates. *Int. J. Pharm. & Phytochem. Res.*, 7(2): 365-369.
- Nagarajan, K., N. Chauhan, A. Mittal, V. Singh, R.B. Bodla and R.K. Tiwari. 2011. Phytochemical extraction, optimization and physico-chemical characterization of two bioactive isolates from the leaves and stem of *Cissampelos pareira*. *Der Pharma Chemical*, 3: 327-337.
- Nandagoapalan, V., A. Doss and C. Marimuthu. 2016. Phytochemical analysis of some traditional medicinal plants. *Biosci. Discover.*, 7(1): 17-20.
- Naz, R. and A. Bano. 2013. Phytochemical screening, antioxidants and antimicrobial potential of *Lantana camara* in different solvents. *Asian Pacif. J. Trop. Dis.*, 3(6): 480-486.
- Odeja, O., G. Obi, C.E. Ogwuche, E.E. Elemike and Y. Oderinlo. 2015. Phytochemical screening, antioxidant and antimicrobial activities of *Senna occidentalis* (L.) leaves extract. *Clini. Phytosci.*, 1-6.
- Okoli, R.I., A.A. Turay, J.K. Mensah and A.O. Aigbe. 2009. Phytochemical and antimicrobial properties of four herbs from Edo State, Nigeria. *Report & Opin.*, 1(5): 67-73.
- Olabinri, B.M., E.A. Adepoju, A.A. Zainab and A.A. Ahmed. 2014. Phytochemical profiling of phytoconstituents of grape, jatropha curcas and neem (*Azadirachta indica*) extracts. *J. Pharm. & Phytotherap.*, 6(2): 17-23.
- Olajuyigbe, O.O., A.E. Babalola and A.J. Afolayan. 2011. Antibacterial and phytochemical screening of crude ethanolic extracts of *Waltheria indica* Linn. *Afr. J. Microb. Res.*, 5(22): 3760-3764.
- Omogbai, B.A. and F.A. Eze. 2011. Phytochemical screening and susceptibility of bacteria pathogens to extracts of *Evolvulus alsinoides*. *Sci. Wor. J.*, 6(1): 5-8.
- Padhi, L.P., S.K. Panda, S.N. Satapathy and S.K. Dutta. 2011. *In vitro* evaluation of antibacterial potential of *Annona squamosa* L. and *Annona reticulata* L. from Similipal Biosphere Reserve, Orissa, India. *J. Agri. Tech.*, 7(1): 133-42.
- Paul, M., K. Vasudevan and K.R. Krishnaja. 2017. Scoparia dulcis: A review on its phytochemical and pharmacological profile. *Innoriginal: Int. J. Sci.*, 4(4): 17-21.
- Pizano, C. and H. García. 2014. El Bosque Seco Tropical en Colombia. Instituto de Investigación de Recursos Biológicos Alexander von Humboldt (IAvH). Bogotá DC, Colombia.
- Rajalakshmi, P., M. Pugalenth and V. Vadivel. 2016. *In vitro* and inhibitory activity of pathogens on leaves of *Argemone mexicana* L. and *Premna tomentosa* L. *Int. J. Herb. Med.*, 4(5): 84-90.
- Rajesh, K., S. Vasantha, N. Rajesh and Pannneerselvam. 2014. Qualitative and quantitative phytochemical analysis in four pteridophytes. *Int. J. Pharm. Sci. Rev Res.*, 27(2): 408-412.
- Rajeswari, S. and P. Vijayashalini. 2017. Comparision of qualitative phytochemical analysis of *Amaranthus polygonoides* L. and *Amaranthus viridis* L. *Int. J. Appl. Res.*, 3(3): 280-283.
- Ribeiro, W.L., W.P. Andre, G.S. Cavalcante, J.V. de Araújo-Filho, J. M. Santos, I.T. Macedo and C.M. Bevílaqua. 2017. Effects of *Spigelia anthelmia* decoction on sheep gastrointestinal nematodes. *Small Rum. Res.*, 153: 146-152.
- Roopan, S.M. 2016. An overview of phytoconstituents, biotechnological applications, and nutritive aspects of coconut (*Cocos nucifera*). *Appl. Biochem. & Biotechnol.*, 179(8): 1309-1324.
- Saidu, A.N., E.O. Aina, A. Mann and U.I. Leje. 2011. The effect of aqueous extract of senna occidentalis leaves on rats infected with salmonella typhi. *Aust. J. Basic & App. Sci.*, 5(12): 1863-1867.
- Saleem, A. and N. Devi. 2015. Antioxidant activity of methanolic extract of *Eclipta Prostrata* (L.). *Int. J. Phytopharm.*, 5(2): 21-24.
- Savithramma, N., M.L. Rao and D. Suhrulatha. 2011. Screening of medicinal plants for secondary metabolites. *Middle-East J. Sci. Res.*, 8(3): 579-584.
- Shanmugam, B., K.R. Shanmugam, S. Ravi, G.V. Subbaiah, K. Mallikarjuna and K.S. Reddy. 2014. Antibacterial activity and phytochemical screening of *Phyllanthus niruri* in ethanolic, methanolic and aqueous extracts. *Int. J. Pharm. Sci. Rev. Res.*, 27(2): 85-89.
- Sharmila, S., R.L. Jeyanthi, M.P. Das, M. Saduzzaman and S. Bala. 2013. A comparative study on phytochemical analysis of murraya koenigii and manilkara zapota. *Res. J. Pharm. Biol. & Chem. Sci.*, 4(2): 1104-1109.
- Sinha, S.N. 2013. Phytochemical profiles and antioxidant activities of the leaf extracts of *Gliricidia sepium*. *Int. J. Innov. Biosci.*, 3(3): 87-91.
- Sivasankari, K., S. Janaky and T. Sekar. 2010. Evaluation of phytochemicals in select medicinal plants of the *Caesalpinia* species. *Ind. J. Sci. & Technol.*, 3(12): 1118-1121.
- Souza, T.M., M.F. Moraes-Braga, J.G. Costa, A.A. Saraiva, M.A. Lima and H.D. Coutinho. 2013. Herbs in association with drugs: Enhancement of the aminoglycoside-antibiotic activity by *Pityrogramma calomelanos* (L.) Link. *J. You. Pharm.*, 5(4): 188-190.
- Sravanthi, M., M. Padmaja, D. Muni Kumar and K.P.J. Hemalatha. 2017. *In vitro* antimicrobial properties and phytochemical screening of crude extracts of *Antigonon leptopus* Hook. & Arn. Leaf. *Int. J. Innov. Pharm. Sci. & Res.*, 5(2): 29-41.
- Sri Rahmaningsih, Prajitno, Aulanniam and Maftuch. (2017). Bioactive compounds from majapahit fruit (*Crescentia cujete*) as a potential natural antibacterial. *Int. J. Chem. Technol. Res.*, 10(3): 90-99.
- Sujatha, J., A. Asokan and Rajeshkumar. 2017. Antioxidant effect and phytochemical analysis of chloroform extract of *Cassis fistula* using FT-IR, HPLC and GC-MS analysis. *Int. J. Pharm. Sci. Rev. & Res.*, 46(1): 129-133.
- Sukalingam, K., K. Ganeshan and B. Xu. 2017. *Trianthema portulacastrum* L. (giant pigweed): phytochemistry and pharmacological properties. *Phytochem. Reviews*, 16(3): 461-478.
- Sultana, S., N. Akhtar and H.M. Asif. 2013. Phytochemical screening and antipyretic effects of hydro-methanol extract of melia azedarach leaves in rabbits. *Bangl. J. Pharm.*, 8(2): 214-217.
- Susmitha, S., K.K. Vidyamol, P. Ranganayaki and R. Vijayaragavan. 2013. Phytochemical extraction and antimicrobial properties of *Azadirachta indica* (neem). *Glob. J. Pharm.*, 7(3): 316-320.
- Tang, Y., H.L. Xin and M.L. Guo. 2016. Review on research of the phytochemistry and pharmacological activities of *Celosia argentea*. *Revista Brasileira de Farmacognosia*, 26(6): 787-796.
- Valdés, A.F., M.J. Mendiola, R.D. Acuña, L.Y. Caballero, L.R. Scull and G.Y. Gutiérrez. 2010. Antimalarial activity and cytotoxicity of hydroalcoholic extracts from six plant species used in Cuban traditional medicine. *Rev Cub de Medi. Trop.*, 63(1): 52-57.

- Veni, T. and T. Pushpanathan. 2014. Investigations of antimicrobial and phytochemical analysis of argemone mexicana medicinal plant extracts against bacteria with gastrointestinal relevance. *Asian J. Pharm. & Clin. Res.*, 7(2): 93-97.
- Wilberforce, J.O. and E.I.N. Olivia. 2017. Phytochemical screening and antimicrobial activity of leaves extracts of *Mangifera indica* and *Carica papaya*. *Int. J. Curr. Microbiol. App. Sci.*, 6(9): 3253-3259.
- Xuan, T.D. and T.D. Khanh. 2016. Chemistry and pharmacology of *Bidens pilosa*: an overview. *J. Pharm. Invest.*, 46(2): 91-132.
- Yadav, R., R.K. Khare and A. Singhal. 2017. Qualitative phytochemical screening of some selected medicinal plants of Shivpuri district (MP). *Int. J. Life. Sci. Sci. Res.*, 3(1): 844-847.
- Yadima, S.G., M. Muhammad and B. Usman. 2017. Phytochemical screening and antibacterial activities of aqueous extracts of *Diospyros mespiliformis*, *Mitragyna inermis*, *Piliostigma reticulatum* and *Tamarindus indica* in Northern Nigeria. *Arid Zone J. Engin, Technol. & Environ.*, 13(3): 411-419.
- Yang, W.C. 2014. Botanical, pharmacological, phytochemical, and toxicological aspects of the antidiabetic plant *Bidens pilosa* L. *Evidence-Based Complem. and Altern. Med.*, 2014: 1-14.

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