

## ASSESSMENT OF SALT TOLERANCE IN *EUCALYPTUS*, RAIN TREE AND THAI NEEM UNDER LABORATORY AND THE FIELD CONDITIONS

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### Abstract

The aim of this investigation was to discover an effective index for salt-tolerant selection in plant tissue culture system and to assay the physiological responses in a salinity field trial. Net-photosynthetic rate (NPR) in salt-tolerant clones of *Eucalyptus* (*Eucalyptus camaldulensis* Dehnh.), Rain tree (*Samanea saman* Merr.) and Thai neem (*Azadirachta siamensis* Val.) was gradually reduced when exposed to 0.17, 0.34, 0.68 or 1.02 M NaCl salts contained in the culture media, while that in salt-sensitive clones was sharply decreased. The reduction percentage of NPR in salt-tolerant species grown under salt stress was lower than that in salt-sensitive species by a factor of 2-3 folds. The NPR reduction in plant species grown under salt-stress was evidently investigated as effective index for salt-tolerance. In addition, physiological characteristics, chlorophyll content and maximum quantum yield of PSII ( $F_v/F_m$ ), in salt-tolerant clones were significantly adapted to salinity field trial at Mahasarakham province, Northeastern region of Thailand, leading to high survival percentage and grew well when compared to the salt-sensitive clones. The salt-tolerant clones of forest tree species can be further used for salinity phytoremediation and ecological succession.

### Introduction

Saline affected areas are dividing into inland and coastal salinity areas, which are a serious problem in many regions of the world *i.e* Africa, Asia, Europe, Latin America, Near East, Australia and North America. The salinity areas are widely spread over 397 million hectares worldwide, which are three times larger than agricultural area (Anon., 2000). The inland salinity area is one of the major problems which is continuously expanding, depending on natural underground salt, unsustainable agricultural cultivation, low quality irrigation, industrial waste and human-induced salinization (Pitman & Läuchli, 2002). Saline soil is an area comprising of various mineral salts in both cations,  $\text{Na}^+$ ,  $\text{Ca}^{2+}$ ,  $\text{Mg}^{2+}$ , and  $\text{K}^+$ , and anions,  $\text{Cl}^-$ ,  $\text{SO}_4^{2-}$ ,  $\text{HCO}_3^-$ ,  $\text{CO}_3^{2-}$ , and  $\text{NO}_3^-$  (Tanji, 2002). The ions directly affect plant growth and development causing either osmotic effect or ionic effect (Neumann, 1997; Mansour & Salama, 2004; Parida & Das, 2005; Chinnusamy *et al.*, 2005; Läuchli & Grattan, 2007). A primary response of plants exposed to salt stress is a decrease in plant water potential, resulting in a detrimental of water use efficiency (Glenn & Brown, 1998). The defense response of higher plants to salt stress is a complex system, which depends on a particular stage of morphological and developmental processes, salt tolerant ability (halophyte or glycophyte) and environmental effects (Ashraf & Harris, 2004). The responses of each genotype are displayed as the cascades of their genetic background and phenotypic expressions. Salt-tolerant or halophytic species seem to lack the unique metabolic machinery, which is

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sensitive to or activated by high toxic-ions, especially  $\text{Na}^+$  and  $\text{Cl}^-$ . Defense responses of halophilic species comprise of many mechanisms such as osmoregulation, ion-homeostasis, antioxidant and hormonal systems (Hasegawa *et al.*, 2000; Ashraf & Foolad, 2007). Therefore, there are many reports that depict that salt-tolerant species can be categorized using physiological criteria such as chlorophyll content and chlorophyll fluorescence (Percival & Fraser, 2001). In that case, net-photosynthetic rate (NPR) is one of the candidate physiological parameters, which is simple, rapid, and sensitive method to establish the salt-tolerance index (Ashraf, 2004).

A major cause of salinity problem is a human activity i.e., deforestation, waste releasing, poor water irrigation, enriched fertilizer etc., (Pannell & Ewing, 2006). Reforestation is one of the most practical and effective strategies to solve the saline soil problems as phytoremediation. Trees cause remediation of saline soil in terms of lowering saline water table, using underground water and decreasing evaporation rate (Barrett-Lennard, 2002). However, the lack of salt-tolerant species is an important barrier to solve this problem. Screening for salt tolerance has been investigated in woody plant species such as Thai neem (Cha-um *et al.*, 2004), olive (Melgar *et al.*, 2008; Marin *et al.*, 1995), *Eucalyptus* (Marcar *et al.*, 2002), acacia (Nguyen *et al.*, 2004), pine (Khasa *et al.*, 2002), toothbrush tree (Ramoliya *et al.*, 2004) and mulberry (Tewary, 2000). Photoautotrophic growth of *In vitro* plantlets is one of fruitful technology to grow the plant with enhanced photosynthesis, leading to stimulation of the physiological, anatomical and morphological characteristics (Kozai *et al.*, 1997). The plantlets cultured under a photoautotrophic system should express phenotypic responses to salt stress, which are more reflective mimic to those grown *ex-vitro*. The photoautotrophic system has been successfully applied to study the salt stress responses in *Albizia lebbek* (Kirdmanee *et al.*, 1997) and screening for salt tolerance (Kirdmanee & Mosaleeyanon, 2000; Cha-um *et al.*, 2004). Nevertheless, the *In vitro* environments are quite different from *ex-vitro* conditions, causing on erratic evaluation of salt-tolerant selections. Field trial of salt-tolerant lines from *In vitro* selections is necessarily required to confirm the adaptation and degree of salt-tolerance. In this study, *Eucalyptus*, Rain tree and Thai neem were chosen as the model plant species, which are widely distributed in all regions of Thailand and dominated in conditions of nutrient deficiency, drought or salinity (Koul *et al.*, 1990; Akilan *et al.*, 1997; Staples & Elevitch, 2005). The objective of this investigation was to discover an effective index for *In vitro* salt-tolerant selection and to assay the physiological responses in a salinity field trial.

## Materials and Methods

**I. Investigation of salinity tolerant index:** Salt-tolerant and salt-sensitive clones of *Eucalyptus*, Rain tree, and Thai neem from photoautotrophic *In vitro* selection were photoautotrophically cultured on liquid-MS media, following the procedure of Cha-um *et al.*, (2003). After 42 days culturing, the culture media were adjusted to 0, 0.17, 0.34, 0.68 and 1.02 M NaCl. Carbon dioxide ( $\text{CO}_2$ ) inside and outside culture vessel containing plantlets was measured using a Gas Chromatograph (GC; Model GC-17A, Shimadzu Co. Ltd., Japan) and net photosynthetic rate (NPR) was calculated according to Fujiwara *et al.*, (1987). The ratio of NPR values of plantlets cultured on MS medium supplemented with 1.02 M NaCl (salt stress) to 0 M NaCl (control) were investigated as salt tolerance index (STI). The STI in each clone was calculated according to the following equation.

$$\text{STI} = \frac{\text{NPR at } 1.02 \text{ M NaCl}}{\text{NPR at } 0 \text{ M NaCl}}$$

The experiment was arranged in a Completely Randomized Design (CRD) with 10 replications and 4 plantlets per replication. Analysis of variance (ANOVA) was analyzed using the SPSS software (SPSS for Windows, SPSS Inc., USA) as well as mean values in each treatment were compared by Duncan's New Multiple Range Test (DMRT).

**II. Physiological responses to salinity in field trials:** The selected salt-tolerant and salt-sensitive clones of *Eucalyptus*, Rain tree and Thai neem from laboratory were photoautotrophically acclimatized, according to Cha-um *et al.*, (2003). The plantlets were transplanted to open plastic bags (size 10W×10L×20H cm), containing a mixture of two parts of soil and one part of vermiculite. The transplanted plantlets were incubated in a glasshouse at 30±2°C ambient temperature, 75±5% relative humidity (RH) and 500±100  $\mu\text{mol m}^{-2} \text{ s}^{-1}$  photosynthetic proton flux density (PPFD) by natural light intensity at plant level with 10 h d<sup>-1</sup> photoperiod. Six-month-old transplanting plantlets were acclimatized by irrigation with 50 mM NaCl under 50% shading natural light intensity for 7 days and then directly planted in either a container (0.5 m in diameter and 1.0 m in height) or salinity field at Mahasarakham, Northeastern region of Thailand. Total chlorophyll content, maximum quantum yield of PSII ( $F_v/F_m$ ), plant height and survival percentage in all clones were measured in rainy, winter and summer seasons. Chlorophyll content in the third leaf of plant from shoot tip was measured using a Chlorophyll Meter SPAD-502 (Minolta, Minolta Co, Ltd, UK) as described by Hussain *et al.*, (2000). The  $F_v/F_m$  of adaxial side of the leaf was monitored using a Plant Efficiency Analyser (PEA version 2.05; Hansatech Instruments Ltd., UK) in the pulse amplitude modulation mode, as previously described by Loggini *et al.*, (1999).

The experiment was arranged in a Completely Randomized Design (CRD) with 10 replications and 4 plants per replication. Analysis of variance (ANOVA) was computed using the SPSS software (SPSS for Windows, SPSS Inc., USA) as well as mean values in each treatment were compared by the *t*-test. The correlation between chlorophyll content and  $F_v/F_m$  was evaluated by Pearson's correlation coefficients.

## Results and Discussion

**I. Investigation of salinity tolerant index:** Net photosynthetic rate (NPR) of Rain tree, *Eucalyptus* and Thai neem grown under photoautotrophic *In vitro* displayed as a similar pattern that reduced with increasing the NaCl concentration in the culture media. The NPR reduction percentage of salt-tolerant clones gradually increased, while the salt-sensitive clone was sharply augmented, relating to NaCl concentrations (Fig. 1). At 1.02 M NaCl, the NPR of the salt-tolerant clones of *Eucalyptus*, Rain tree and Thai neem remained higher than those of salt-sensitive clones by 1.17, 1.54 and 1.45 folds, respectively (Table 1). The NPR of salt-tolerant and salt-sensitive plantlets was dramatically inhibited by high salt concentrations (0.68-1.02 M NaCl). In addition, the NPR reduction of salt-sensitive plantlets grown under salt stress quickly declined when compared to salt-tolerant plantlets. The ratio of NPR of salt-stress (1.02 M NaCl) and control plantlets was directly applied to sensitive indicator as salt-tolerant index (STI) for rapid salt-tolerant screening (Table 1).

**Table 1.** Net-photosynthetic rate of salt-tolerant (ST) and salt-sensitive (SS) clones of *Eucalyptus* (EU), Rain tree (RT) and Thai neem (TN) grown under varying various concentrations of sodium chloride (0-1.02 M NaCl) under *In vitro* photoautotrophic system for a day. Salt-tolerance index (STI) is represented by the ratio of NPR at 1.02 M NaCl and NPR at 0 M NaCl.

NaCl (M)	Net-photosynthetic rate ( $\mu\text{mol m}^{-2} \text{s}^{-1}$ )					
	ST-EU	SS-EU	ST-RT	SS-RT	ST-TN	SS-TN
0	13.99a	13.91a	16.97a	16.62a	11.39a	11.06a
0.17	13.57ab	13.11ab	16.77a	16.47a	11.25a	10.14b
0.34	13.47ab	12.97b	16.43a	15.28a	10.76b	9.76b
0.68	12.87b	11.55b	15.29b	13.03b	10.43b	8.46c
1.02	11.69c	9.96c	14.26c	9.28c	9.81c	6.77d
STI	0.84	0.72	0.84	0.56	0.86	0.61

The different letters in each column are significantly different at  $P \leq 0.01$  by New Duncan's Multiple Range test (DMRT)

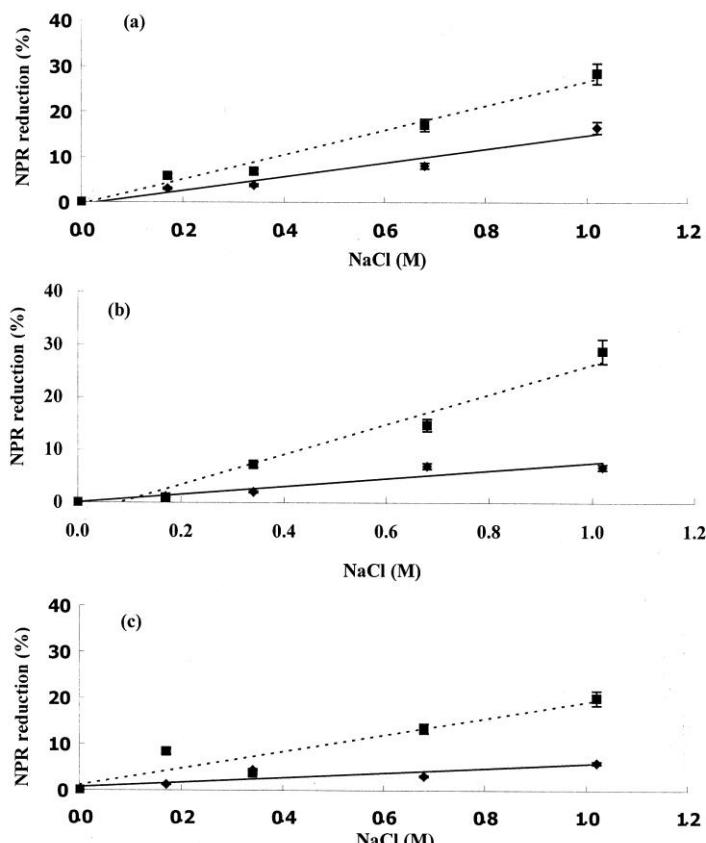


Fig. 1. Net photosynthetic rate (NPR) reduction percentage of salt-tolerant (dark-line) and salt-sensitive (dot-line) clones of *Eucalyptus* (a), Rain tree (b) and Thai neem (c) photoautotrophically grown under 0, 0.17, 0.32, 0.68 or 1.02 M NaCl in the culture medium for a day treatment.

There are many documents on NPR reduction of tree species, growing in the salinity condition such as *Populus* species (Sixto *et al.*, 2005), tamarind (Gebauer *et al.*, 2004), Thai neem (Cha-um *et al.*, 2004), *Citrus* (Torrecillas *et al.*, 2003) and olive tree (Chartzoulakis *et al.*, 2002). Salt stress is directly inhibited either on light reaction, pigment degradation, water oxidation and electron transportation, or dark reaction, Rubisco enzyme, stomatal conductance, CO<sub>2</sub>-assimilation and transpiration rate (Chartzoulakis *et al.*, 2002; Meloni *et al.*, 2003; Tezara *et al.*, 2003). Moreover, the NPR reduction in plant cultured under photoautotrophic salt-stress condition had been successfully applied to use as effective index, simple method, and rapid evaluation for salt-tolerant selection (Kirdmanee & Mosaleeyanon, 2000; Cha-um *et al.*, 2004). The STI of forest species, *Eucalyptus*, Rain tree and Thai neem, should be played as an effective index for salt-tolerant selection in breeding program of forest tree species.

**II. Physiological responses to salinity in the field trials:** Selected clones of salt-tolerant and salt-sensitive *Eucalyptus*, Rain tree and Thai neem from laboratory classification were intensively pretreated with salt-solution in the field trial environments of the salinity field. These plants were greatly adapted to those conditions in both salinity field and container of the rainy season in May-October 2004 because of high precipitation rate at 100-250 mm per month with high relative humidity (RH). Plant height of salt-tolerant clones in *Eucalyptus*, Rain tree and Thai neem was significantly higher than those salt-sensitive clones in both salinity field and container. In addition, the plant height of forest species grown in the container exhibited more than those grown on the salinity field trial in all seasons (Table 2). In the winter season of salinity field, plant height of salt-tolerant clones in *Eucalyptus*, Rain tree and Thai neem was better than those salt sensitive clones by the factors of 1.14, 1.54 and 1.65 folds, respectively. In rainy season, survival percentage of salt-sensitive clones of *Eucalyptus* and Rain tree was 100%, while Thai neem was gradually reduced to 80-90%. On the other hand, the survival percentage of salt-sensitive clones of forest species grown on both salinity field and container continuously decreased higher than those salt-tolerant clones, especially in the winter and summer seasons. As well as, the salt sensitive clone of Thai neem absolutely died in the salinity field of summer season (Fig. 2). In all seasons, chlorophyll pigment concentrations in the leaf tissues of salt-sensitive clones were significantly dropped higher than those salt-tolerant species when grown in both salinity field and container. The chlorophyll concentration in salt-tolerant clones of *Eucalyptus*, Rain tree and Thai neem was higher than those salt-sensitive clones by the factors of 1.10, 1.24 and 1.28 folds, respectively in the salinity field of rainy season. The chlorophyll contents in both salt-tolerant and salt-sensitive species were gradually reduced in the winter season and then increased in the summer season, except salt-sensitive clone of Thai neem died already in the salinity field (Fig. 3). In the similar pattern, maximum quantum yield ( $F_v/F_m$ ) of salt-sensitive forest species was significantly reduced lower than those salt-tolerant species, especially in the winter season (Fig. 4). Moreover, the chlorophyll concentrations in the leaf tissues of *Eucalyptus*, Rain tree and Thai neem were positively related to  $F_v/F_m$  ( $r = 0.83$ ,  $r = 0.85$  and  $r = 0.88$ , respectively) (Fig. 5). It means that the adaptation ability of salt-tolerant species in salinity field was better than those salt-sensitive species, identifying by chlorophyll content and  $F_v/F_m$ , leading to high survival percentage and growth performance in salinity field.

**Table 2. Plant height (cm) of salt-tolerant (ST) and salt-sensitive (SS) clones of *Eucalyptus* (EU), Rain tree (RT) and Thai neem (TN) grown in a salinity field in the rainy, winter and summer seasons.**

Season	Plant height (cm)					
	ST-EU	SS-EU	ST-RT	SS-RT	ST-TN	SS-TN
Rainy						
Container	83.0a	62.9b	52.1a	43.9b	18.3a	13.6b
Salinity field	55.0a	43.1b	47.0a	40.4b	18.4a	13.5b
Winter						
Container	158.4a	125.0b	83.0a	62.9b	45.2a	31.9b
Salinity field	145.4a	120.5b	66.2a	43.1b	45.0a	27.3b
Summer						
Container	168.4a	132.1b	95.4a	80.7b	52.5a	36.1b
Salinity field	165.6a	127.3b	77.0a	58.8b	52.9	*

The different letters in each column are significantly different at  $P \leq 0.01$  by *t*-test

\* The plants absolutely died.

The conduction of field trial in a salinity area is one of the most important practices to evaluate the salt-tolerant ability of many plant species such as poplar (Sixto *et al.*, 2005), desert plants (Arndt *et al.*, 2004), *Eucalyptus* (Mahmood *et al.*, 2003; Morris *et al.*, 2006), tree species (Tomar *et al.*, 2003), deciduous tree species (Paludan-Muller *et al.*, 2002), and pannonian endemism (Mile *et al.*, 2002). The salt-tolerant species are significantly adapted to salinity by several defense mechanisms, limited-ions absorption and translocation (Chartzoulakis *et al.*, 2002; Paludan-Muller *et al.*, 2002; Fernandez-Ballester *et al.*, 2003; Estan, *et al.*, 2005), osmoregulation *i.e.* proline (de Lacerda *et al.*, 2005; Misra & Gupta, 2005), soluble carbohydrate (de Lacerda *et al.*, 2005), and glycine betaine (Khan *et al.*, 1998), and antioxidation (Vaidyanathan *et al.*, 2003) with regular biochemical and physiological characteristics *i.e.*, chlorophyll stabilization (Misra & Gupta, 2005), water use efficiency (Tezara *et al.*, 2003), chlorophyll fluorescence (Percival & Fraser, 2001), and net-photosynthetic rate (Tezara *et al.*, 2003), leading to growth stimulation as well as high survival percentage (Chartzoulakis *et al.*, 2002; Torrecillas *et al.*, 2003; Nasim *et al.*, 2008). The ability of salt-tolerance depends on various factors such as salt-concentration (Chartzoulakis *et al.*, 2002; Gebauer *et al.*, 2004), plant species (Percival & Fraser, 2001; Sixto *et al.*, 2005; Nasim *et al.*, 2008), and plant varieties (Chartzoulakis *et al.*, 2002; Mahmood *et al.*, 2003; Torrecillas *et al.*, 2003; Sixto *et al.*, 2005). In our results of the present study show that the salt-tolerant species of forest tree species significantly adapted to realistic saline soil and environments in terms of physiological adaptations, resulted in high survival percentage when compared to the sensitive species. In addition, the survival percentage of forest species was positively related to winter and summer seasons because of the low relative humidity with high light intensity and high temperature in the field trial. It means that the environmental conditions of a salinity field trial, especially summer season should be essentially considered while classifying plants for salt tolerance.

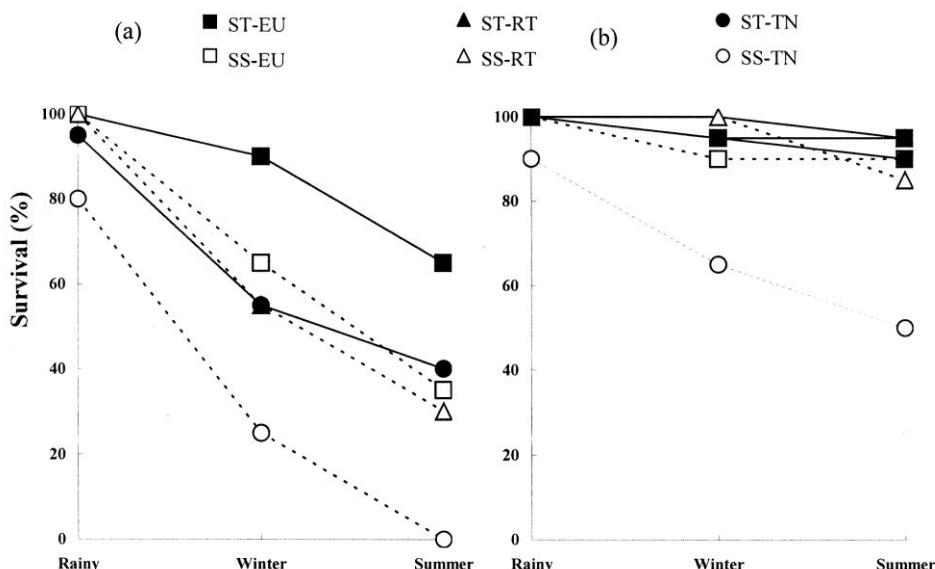


Fig. 2. Survival percentage of salt-tolerant (ST) and salt-sensitive (SS) clones of *Eucalyptus* (EU), Rain tree (RT) and Thai neem (TN) grown in a salinity field (a) and in a container (b) in the rainy, winter and summer seasons.

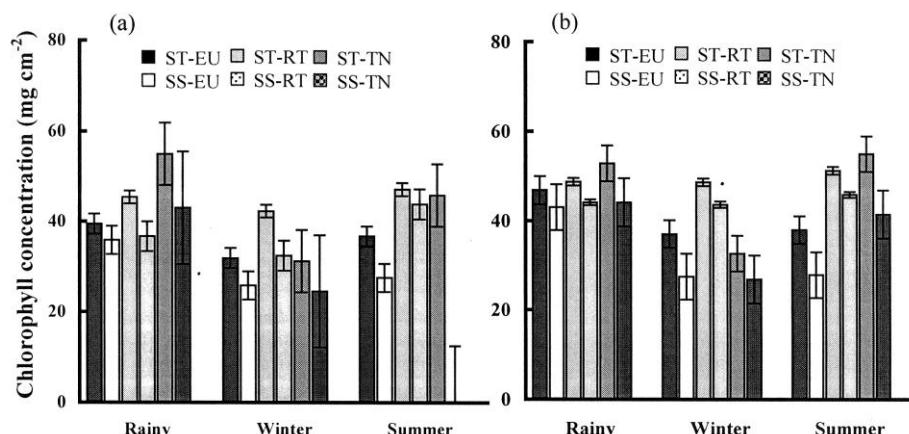


Fig. 3. Chlorophyll concentration in the leaf tissues of salt-tolerant (ST) and salt-sensitive (SS) clones of *Eucalyptus* (EU), Rain tree (RT) and Thai neem (TN) grown in a salinity field (a) and in a container (b) in the rainy, winter and summer seasons.

## Conclusion

Salt-tolerant index (STI) of *In vitro* photoautotrophic plantlets was a more rapid indicator for selection for salt-tolerance. The salt-tolerant species of forest trees, *Eucalyptus*, Rain tree, and Neem tree, identified by STI were greatly acclimatized to salinity field in terms of chlorophyll stabilization and water oxidation, resulting in high survival percentage with growth stimulation. The salt-tolerant forest species should be further reforested in salinity areas as an efficient phytoremediation.

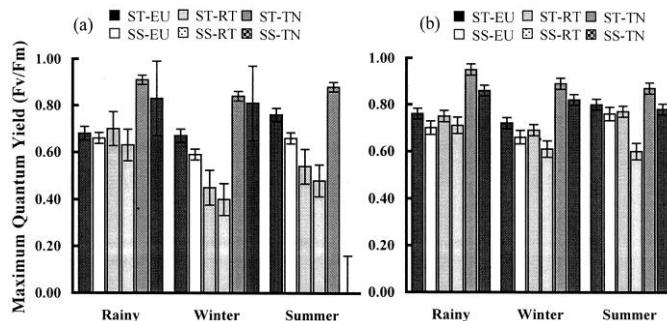


Fig. 4. Maximum quantum yield ( $F_v/F_m$ ) in the leaf tissues of salt-tolerant (ST) and salt-sensitive (SS) clones of *Eucalyptus* (EU), Rain tree (RT) and Thai neem (TN) grown in a salinity field (a) and a container (b) in the rainy, winter and summer seasons.

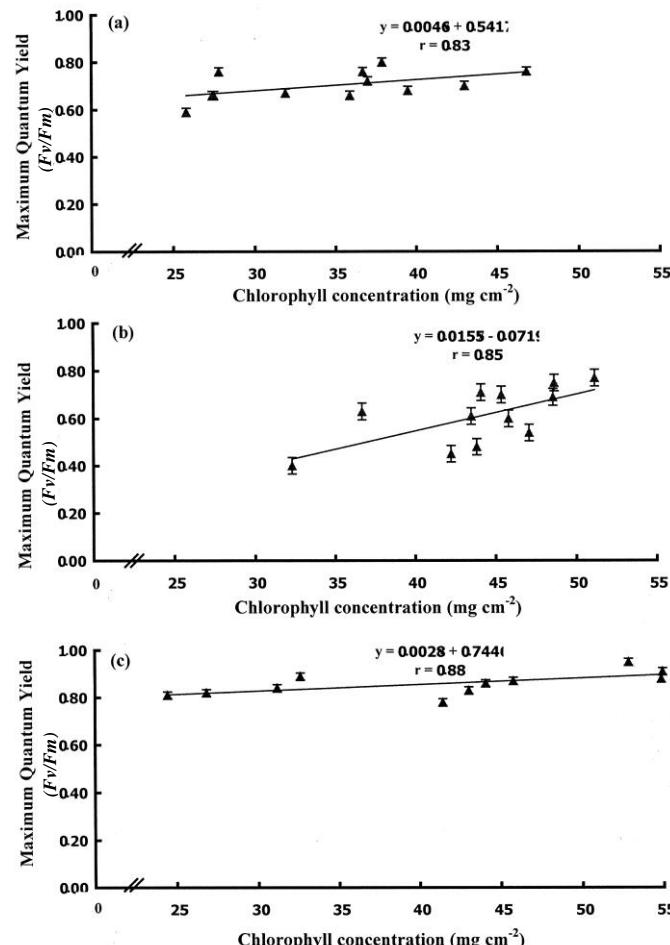


Fig. 5. Relationship between chlorophyll concentration and maximum quantum yield ( $F_v/F_m$ ) of *Eucalyptus* (a), Rain tree (b) and Thai neem (c) grown in a salinity field.

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## References

Akilan, K., R.C.C. Farrell, D.T. Bell and J.K. Marshall. 1997. Responses of clonal river red gum (*Eucalyptus camaldulensis*) to waterlogging by fresh and salt water. *Aust. J. Exp. Agric.*, 37: 243-248.

Anonymous. 2000. Extent and causes of salt-affected soils in participating countries. Land and Plant Nutrition Management Service. Global network on integrated soil management for sustainable use of salt-affected soils [online]. Available from <http://www.fao.org/ag/agl/agll/spush/topic2.html>.

Arndt, S.K., C. Arampatsis, A. Foetzki, X. Li, F. Zeng and X. Zhang. 2004. Contrasting patterns of leaf solute accumulation and salt adaptation in four phreatophytic desert plants in a hyperarid desert with saline groundwater. *J. Arid. Environ.*, 59: 259-270.

Ashraf, M. 2004. Some important physiological selection criteria for salt tolerance in plants. *Flora*, 199: 361-376.

Ashraf, M. and P.J.C. Harris. 2004. Potential biochemical indicators of salinity tolerance in plants. *Plant Sci.*, 166: 3-16.

Ashraf, M. and M.R. Foolad. 2007. Improving plant abiotic-stress resistance by exogenous application of osmoprotectants glycinebetaine and proline. *Environ. Exp. Bot.*, 59: 206-216.

Barrett-Lennard, E.G. 2002. Restoration of saline land through revegetation. *Agric. Water Manage.*, 53: 213-226.

Chartzoulakis, K., M. Loupassaki, M. Bertaki and I. Androulakis. 2002. Effects of NaCl salinity on growth, ion content and CO<sub>2</sub> assimilation rate of six olive cultivars. *Sci. Hort.*, 96: 235-247.

Cha-um, S., K. Mosaleeyanon, K. Supaibulwata and C. Kirdmanee. 2003. A more efficient transplanting system for Thai Neem (*Azadirachta siamensis* Val.) by reducing relative humidity. *Sci. Asia*, 29: 189-196.

Cha-um, S., K. Supaibulwata and C. Kirdmanee. 2004. Physiological responses of Thai neem (*Azadirachta siamensis* Val.) to salt stress for salt-tolerance screening program. *Sci. Asia*, 30: 17-23.

Chinnusamy, V., A. Jagendorf and J.K. Zhu. 2005. Understanding and improving salt tolerance in plants. *Crop Sci.*, 45: 437-448.

De Lacerda, C.F., J. Cambraia, M.A. Oliva and H.A. Ruiz. 2005. Changes in growth and in solute concentrations in sorghum leaves and roots during salt stress recovery. *Environ. Exp. Bot.*, 54: 69-76.

Estan, M.T., M.M. Martinez-Rodriguez, F. Perez-Alfocea, T.J. Flowers and M.C. Bolarin. 2005. Grafting raises the salt tolerance of tomato through limiting the transport of sodium and chloride to the shoot. *J. Exp. Bot.*, 56: 703-712.

Fernandez-Ballester, G., F. Garcia-Sanchez, A. Cerda and V. Martinez. 2003. Tolerance of citrus rootstock seedlings to saline stress based on their ability to regulate ion uptake and transport. *Tree Physiol.*, 23: 265-271.

Fujiwara, K., T. Kozai and L. Watanabe. 1987. Fundamental studies on environment in plant tissue culture vessel. (3) Measurement of carbon dioxide gas concentration in closed vessels containing tissue cultured plantlets and estimates of net photosynthetic rates of the plantlets. *J. Agric. Meteorolol.*, 43: 21-30.

Gebauer, J., K. El-Siddig, A.A. Salih and G. Ebert. 2004. *Tamarindus indica* L., seedlings are moderately salt-tolerant when exposed to NaCl-induced salinity. *Sci. Hort.*, 103: 1-8.

Glenn, E.P. and J.J. Brown. 1998. Effects of soil salt levels on the growth and water use efficiency of *Atriplex canescens* (Chenopodiaceae) varieties in drying soil. *Amer. J. Bot.*, 85: 10-16.

Hasegawa, P.M., R.A. Bressan, J.K. Zhu and H.J. Bohnert. 2000. Plant cellular and molecular responses to high salinity. *Annu. Rev. Plant Physiol. Plant Mol. Biol.*, 51: 463-499.

Hussain, F., K.F. Bronson, Y. Singh, B. Singh and S. Peng. 2000. Use of chlorophyll meter sufficiency indices for nitrogen management of irrigated rice in Asia. *Agron. J.*, 92: 875-879.

Khan, M.A., I.A. Ungar, A.M. Showalter and H.D. Dewald. 1998. NaCl-induced accumulation of glycinebetaine in four subtropical halophytes from Pakistan. *Physiol. Plant.*, 102: 487-492.

Khasa, P.D., B. Hambling, G. Kernaghan, M. Fung and E. Ngimbi. 2002. Genetic variability in salt tolerance of selected boreal woody seedlings. *Forest Ecol. Manage.*, 165: 257-269.

Kirdmanee, C., S. Cha-um and R. Wanussakul. 1997. Morphological and physiological comparisons of plantlets *In vitro*: response to salinity. *Acta Hort.*, 457: 181-186.

Kirdmanee, C. and K. Mosaleeyanon. 2000. Environmental engineering for transplant production. In: *Transplant Production in the 21<sup>st</sup> Century*, pp. 78-81. (Eds.): C. Kubota & C. Chun. Kluwer Academic Publishers, Netherlands.

Koul, O., M.B. Isman and C.M. Ketkar. 1990. Properties and uses of neem *Azadirachta indica*. *Can. J. Bot.*, 68: 1-11.

Kozai, T., C. Kubota and B.R. Jeong. 1997. Environmental control for the large-scale production of plants through *In vitro* techniques. *Plant Cell Tiss. Org. Cult.*, 51: 49-56.

Läuchli, A. and S.R. Grattan. 2007. Plant growth and development under salinity stress. In: *Advances in Molecular Breeding Toward Drought and Salt Tolerant Crops*, pp. 285-315. (Eds.): M.A. Jenks, P.M. Hasegawa and S.M. Jain. Springer, Dordrecht, Netherlands.

Loggini, B., A. Scartazza, E. Brugnoli and F. Navari-Izzo. 1999. Antioxidant defense system, pigment composition and photosynthetic efficiency in two wheat cultivars subjected to drought. *Plant Physiol.*, 119: 1091-1099.

Mahmood, K., N.E. Marcar, M.H. Naqvi, R.J. Arnold, D.F. Crawford, S. Iqbal and K.M. Aken. 2003. Genetic variation in *Eucalyptus camaldulensis* Dehnh., for growth and stem straightness in a provenance-family trial on salt land in Pakistan. *Forest Ecol. Manage.*, 176: 405-416.

Mansour, M.M.F. and K.H.A. Salama. 2004. Cellular basis of salinity tolerance in plants. *Environ. Exp. Bot.*, 52: 113-122.

Marcar, N.E., D.F. Crawford, A. Saunders, A.C. Matheson and R.A. Arnold. 2002. Genetic variation among and within provenances and families of *Eucalyptus grandis* W. Hill and *E. globulus* Labill. subsp. *globulus* seedlings in response to salinity and waterlogging. *Forest Ecol. Manage.*, 162: 231-249.

Marin, L., M. Benlloch and R. Fernández-Escobar. 1995. Screening of olive cultivars for salt tolerance. *Sci. Hort.*, 64: 113-116.

Melger, J.C., J.P. Syvertsen and F. García-Sánchez. 2008. Can elevated CO<sub>2</sub> improve salt tolerance in olive tree? *J. Plant Physiol.*, 165: 631-640.

Mile, O., I. Meszaros, S. Veres and G. Lakatos. 2002. Ecophysiological study on the salt tolerance of a pannonian endemism [*Lepidium crassifolium* (W. et K.)] in inland saline area. *Acta Biol. Szegediensis*, 46: 249-250.

Misra, N. and A.K. Gupta. 2005. Effects of salt stress on proline metabolism in two high yielding genotypes of green gram. *Plant Sci.*, 196: 331-339.

Morris, J., J. Collopy and K. Mahmood. 2006. Canopy conductance and water use in *Eucalyptus* plantations. *Pak. J. Bot.*, 38: 1485-1490.

Neumann, P. 1997. Salinity resistance and plant growth revisited. *Plant Cell Environ.*, 20: 1193-1198.

Nasim, M., R.H. Qureshi, T. Aziz, M. Saqib, S. Nawaz, S.T. Sahi and S. Pervaiz. 2008. Growth and ionic composition of salt-stressed *Eucalyptus camaldulensis* and *Eucalyptus tereticornis*. *Pak. J. Bot.*, 40: 799-805.

Nguyen, N.T., R.E.A. Moghaieb, H. Seneoka and K. Fujita. 2004. RAPD markers associated with salt tolerance in *Acacia auriculiformis* and *Acacia mangium*. *Plant Sci.*, 167: 797-805.

Paludan-Muller, G., H. Saxe, L.B. Pedersen and T.B. Randrup. 2002. Differences in salt sensitivity of four deciduous tree species to soil or airborne salt. *Physiol. Plant.*, 114: 223-230.

Pannel, D.J. and M.A. Ewing. 2006. Managing secondary dryland salinity: Option and challenges. *Agric. Water Manage.*, 80: 41-56.

Parida, A.K. and A.B. Das. 2005. Salt tolerance and salinity effects on plants: a review. *Ecotoxicol. Environ. Safe.*, 60: 324-349.

Percival, G.C. and G.A. Fraser. 2001. Measurement of the salinity and freezing tolerance of *Crataegus* genotypes using chlorophyll fluorescence. *J. Arboricult.*, 27: 233-245.

Pitman, M.G. and A. Läuchli. 2002. Global impact of salinity and agricultural ecosystems. In: *Salinity: Environment-Plant-Molecules*, pp. 3-20. (Eds.): A. Lauchli & U. Luttge. Kluwer Academic Publishers, Dordrecht, The Netherlands.

Ramoliya, P.J., H.M. Patel and A.N. Pandey. 2004. Effect of salinization of soil on growth and macro- and micro-nutrient accumulation in seedlings of *Salvadora persica* (Salvadoraceae). *Forest Ecol. Manage.*, 202: 181-193.

Reuveni, J., J. Gale and M. Zeroni. 1997. Differentiating day from night effects of high ambient [CO<sub>2</sub>] on the gas exchange and growth of *Xanthium strumarium* L., exposed to salinity stress. *Ann. Bot.*, 79: 191-196.

Sixto, H., J.M. Grau, N. Alba and R. Alia. 2005. Response to sodium chloride in different species and clones of genus *Populus* L. *Forest.*, 78: 93-104.

Staples, G.W. and C.R. Elevitch. 2005. *Samanea saman* (rain tree). In: *Species profiles for Pacific Island Agroforestry [online]*. (Ed.): C.R. Elevitch. Available from <http://www.traditionaltree.org>. Permanent Agriculture Resources (PAR), Holualoa, Hawaii.

Tanji, K.K. 2002. Salinity in the soil environment. In: *Salinity: Environment-Plant-Molecules*, pp. 21-51. (Eds.): A. Läuchli & U. Luttge. Kluwer Academic Publishers, Dordrecht, The Netherlands.

Tezara, W., D. Martinez, E. Rengifo and A. Herrera. 2003. Photosynthetic responses of the tropical spiny shrub *Lycium nodosum* (Solanaceae) to drought, soil salinity and saline spray. *Ann. Bot.*, 92: 757-765.

Tewary, P.K., A. Sharma, M.K. Raghunath and A. Sarkar. 2000. *In vitro* response of promising mulberry (*Morus* sp.) genotypes for tolerance to salt and osmotic stresses. *Plant Growth Reg.*, 30: 17-21.

Tomar, O.S., P.S. Minhas, V.K. Sharma, Y.P. Singh and R.K. Gupta. 2003. Performance of 31 tree species and soil conditions in a plantation established with saline irrigation. *Forest Ecol. Manage.*, 177: 333-346.

Torrecillas, A., P. Rodriguez and M.J. Sanchez-Blanco. 2003. Comparison of growth, leaf water relations and gas exchange of *Citrus albidus* and *C. monspeliensis* plants irrigated with water of different NaCl salinity levels. *Sci Hort.*, 97: 353-368.

Vaidyanathan, H., P. Sivakumar, R. Chakrabarty and G. Thomas. 2003. Scavenging of reactive oxygen species in NaCl-stressed rice (*Oryza sativa* L.)-differential response in salt-tolerant and sensitive varieties. *Plant Sci.*, 165: 1411-1418.

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