LOCATION OF MACROPHOMINA PHASEOLINA IN SEEDS OF PUMPKIN AND DEVELOPMENT OF CHARCOAL ROT DISEASE

NASREEN SULTANA, A.K. KHANZADA AND A. GHAFFAR

Crop Diseases Research Institute, Pakistan Agricultural Research Council, Karachi University Campus, Karachi-75270, Pakistan.

Abstract

Using component plating technique Macrophomina phaseolina was isolated from seed coat, tegricuit, cotyledons and embryo of pumpkin seeds. Infected seeds give rise to infected seedlings that transmit the pathogen into the fruits.

Introduction

Macrophomina phaseolina (Tassi) Goid., a soil inhabiting fungus is known to produce seed rot, seedling blight, wilt and root rot on over 500 species of plants in different parts of the world (Sinclair, 1982). At least 67 hosts of M. phaseolina have been reported from Pakistan (Mirza & Qureshi, 1988; Shahazad & Ghaffar, 1986; Shahazad et al., 1988). M. phaseolina has been reported from seeds of bottle-gourd and musk melon (Maholay, 1988, 1989). The fungus has been detected in association with seed coat and cotylenons of melon and was found to penetrate the fruit via the peduncle to infect the seed (Reuveni et al., 1983). Experiments were carried out to study the location of M. phaseolina in the seeds of pumpkin (Cucurbita pepo) and its effect on the germination of seed and development of the disease.

Materials and Methods

Seed samples of pumpkin were collected from Hyderabad, Sindh, Pakistan. Based on macroscopic observation, the seeds were separated into three catogories viz., i) seeds with black dotted spots; ii) healthy looking seeds with few dotted spots, iii) healthy looking seeds with no dotted spots. For determining the location of fungi in seed, the seeds were washed and soaked in distilled water for 2 hours and then aseptically dissected to separate different component viz., seed coat, tegment, cotyledons and embryo. Dissected seed parts were surface disinfected with 0.5% NaOCl solution and placed on blotters in Petri dishes (Du-Hyunglee et al., 1984).

For transmission studies, ten seeds were placed on 3 layeres of well soaked blotter in 9 cm diam Petri dishes. After five days, the lids were removed and dishes put in polyethylene bags to study seedling symptoms for 15 days. Single seed was placed in 200x20 mm test tube containing 15 ml of 1% plain water agar. The cotton plug was removed as the seedling reached to the top. A set of 15 Petri dishes and 150 test tubes for each category of seeds was incubated at 25°C under 12 hours of alternating

^{*}Department of Botany, University of Karachi, Karachi-75270, Pakistan.

Table 1. Percent recovery of Macrophomina phaseolina from seeds of pump kin.

Parts of seed	Ca	S	
	Α	В	C
Seed Coat	100a ²	62a	22a
Tegment	87a	67a	7b
Cotyledon	12b	7b	0Ь
Embryo	7b	9 a	0b

A: Seeds with black dotted spots, B: Healthy looking seeds with few black dotted spot, C: Healthy looking seeds. ²Number followed by the same latter within a column are not significantly different (P = 0.05) according to Duncan's multiple range test.

cycle of ADL and darkness (Khare et al., 1977). In another experiment, five seeds were sown in earthen pots containing 3 kg of autoclaved soil. A set of 20 pots was used for each category. Seeds were also sown in 3.5x5 m plot in two rows, 3.5 m long and 3 m apart. The pots and field experiments were carried out during winter (20-28°C) and then in summer (28-40°C) and emergence of seeds and M. phaseolina infection on seedling were recorded. The data were analysed using Duncan's Multiple Range test.

Table 2. Seed to seedling transmission of *Macrophomina phaseolina* under laboratory conditions in pumpkin seeds.

Categories	Preemer- gence infection	Petriplat	es		Test Tub	e
		Post Eme Infection At hypo- cotyl & radicle axis		Preemer- gence Infection	Post Eme Infection At hypo- cotyl & radicle axis	
Seeds with black dotted spots	9.3a	11.0a	3.0a	15.0a	8.0a	1.7a
Healthy looking seeds with few black dotted spots	4.7a	7.0a	4.7a	11.7a	6.0a	2.0a
Healthy looking seeds	0Ь	3.0b	0b	0.7b	2.0b	0b

Means followed by the same letter within a column are not significantly different (P=0.05) according to Duncan's multiple range test.

Results and Discussion

Where seeds with dotted spots were used, M. phaseolina infection in seed coat and in tegment was significantly higher than in cotyledons and embryo. M. phaseolina infection in healthy looking seeds was low in seed coat and tegment and was not detected on cotyledons and embryo (Table 1).

There was a direct correlation between cotyledons and embryo infection and loss in germination (Table 2,3). Most of the ungerminated seeds were covered with gray to black loose mycelium with minute black sclerotia. The seedling in which the seed coat was either attached to the peg or cotyledons showed browning of hypocotyl and root axis resulting in death of the seedling. In seedling where seed coat remained attached to the cotyledons during germination, there was yellowing and browning of cotyledonary leaves resulting in death of the aerial parts. Seedling mortality was significantly greater (p < 0.05) during summer as compared to winter sowing in both the pot and field experiments. Such similar results have been reported by Reuveni et al., (1983) who found that infected seed gave rise to infected plant with more severe symptoms under warmer conditions.

Adult plant grown from seed having dotted spot showed upto 14% M. phaseolina infection during summer with no infection in winter sowing. Reuveni et al., (1982) also reported that infection was more prevalent in summer than in winter grown Cucumis melo. Similarly, high temperature has been found to favour M. phaseolina infection on soybean (Agarwal et al., 1973; Meyer et al., 1974), pine (Hodges, 1962), corn and sorghum (Livingston, 1943) and bean and cowpea (Thompkins & Gardner, 1935). Disease severity in castor bean was highest where warm and moist spring were followed by hot and dry weather (Cook, 1955). In the present study infected plants showed stunted growth, vine decline, root and basal stem rot, reduction in number and size of fruit, fruit shedding just after formation and rotting of the developed fruit. Minute black sclerotia were also seen on the surface of seeds and inner mesocarpic tissues of

Table 3. Effect of sowing seed infected with *Macrophomina phaseolina* on plant emergence and seedling mortality.

		POTS		FIELD	
Categories		Emer- gence	Seedling Mortality	Emer- gence	Seedling Mortality
Seeds with black	Summer	17.0b	10.8a	17.6b	9.3a
dotted spots	Winter	19.3b	3.0b	19.0ab	4.0b
Healthy looking seeds with	Summer	20.0ab	7.8a	21.0ab	5.8ab
with few black dotted spots	Winter	20.5ab	2.0bc	21.8ab	1.5c
Healthy looking seeds	Summer	24.0a	4.3b	23.5a	5.0b
	Winter	25.0a	0.5c	25.0a	1.0c

Means followed by the same letter within a column are not significantly different (P=0.05) according to Duncan's multiple range test.

infected fruit. M. phaseolina was isolated from the disease plant part when placed on PDA medium. The characteristic charcoal rot symptoms appeared in the basal portion of few plants. Reuveni et al., (1983), reported that the fungus can penetrate melon fruit via peduncle and infect the seeds as well.

The results of the present study showed that *M. phaseolina* infection is extra as well as intra-embryonal which affects the germination of the seeds and produces disease symptoms on seedling and the adult plant. There is therefore need to use healthy seeds to prevent spread of inoculum in non-infested field.

References

- Agarval, D.K.S. Gangopodhyay and A.K. Sarbhoy. 1973. Effect of temperature on the charcoal rot disease of soybean. *Indian Phytopathol.*, 26: 587-589.
- Cook, A.A. 1955. Charcoal rot of castor bean in the United States. Plant Dis. Rep., 39: 233-235.
- Du-Hyunglee, S.B. Mathur and P. Neergaard. 1984. Detection and location of seed-borne inoculum of *Didymella bryoniae* and its transmission in seedlings of cucumber and pumpkin. *Phytopathology Z.*, 109: 301-308.
- Hodges, C.S. 1962. Black root rot of pine seedlings. Phytopathology, 52: 210-219.
- Khare, M.N., S.B. Mathur and P. Neergaard. 1977. A seedling symptom test for detection of Septoria nodonum in wheat seed. Seed Science & Technol., 5: 613-617.
- Livingston, J.E. 1945. Charcoal rot of corn and sorghum. Nebraska Agric. Exp. Stn. Res. Bull., 136: 32p.
- Maholay, M.N. 1988. Seed-borne diseases of cucurbits. Muskmelon (Cucumis melo L.). Seed & Farm, 14:11-12.
- Maholay, M.N. 1989. Seed-borne diseases of cucurbits III. Bottlegourd (Lagenaria siceraria (Mol.) Standl. Seed & Farms, 15: 30-31.
- Meyer, W.A., J.B. Sinclair and M.N. Khare. 1974. Factor affecting charcoal rot of soybean seedlings. *Phytopathology*, 64: 845-849.
- Mirza, J.H. and M.S.A. Qureshi. 1978. Fungi of Pakistan. Dept. of Plant Pathology, Univ. of Agric., Faisalabad, pp. 311.
- Reuveni, R., J. Krikun, A. Nachmias and E. Shlevin. 1982. The role of *Macrophomina phaseolina* in a collapse of melon plants in Israel. *Phytoparasitica*, 10: 51-56.
- Reuveni, R., A. Nachmias and J. Krikun. 1983. The role of seedborne inoculum on the development of *Macrophomina phaseolina* on melon. *Plant Diseases*, 67: 280-281.
- Shahzad, S. and A. Ghaffar. 1986. Macrophomina phaseolina (Tassi) Goid, on some new hosts in Pakistan. FAO Plant Protect. Bull., 34: 163.
- Shahzad, S., A. Sattar and A. Ghaffar. 1988. Additions to the host of *Macrophomina phaseolina*. Pak. J. Box, 20: 151-152.
- Sinclair, J.B. 1982. Compendium of soybean diseases. 2nd ed. American Phytopthological Society. pp. 104.
- Thompkins, C.M. and M.W. Gardner. 1935. Relation of temperature to infection of bean and cowpea seedlings by Rhizoctonia bataticola. Hilgardia, 9:219-230.

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