

CHEMICAL AND BIOLOGICAL CONTROL OF *FUSARIUM* ROOT ROT OF OKRA

ZAHOOR AHMAD¹, SAIFULLAH^{1*}, FAZLI RAZIQ¹, HAKIM KHAN¹ AND MUHAMMAD IDREES²

¹Department of Plant Pathology, Khyber Pakhtunkhwa Agricultural University, Peshawar-Pakistan

²Department of Agricultural Extension Education and Communication, Khyber Pakhtunkhwa Agricultural University, Peshawar-Pakistan

Abstract

The effect of *Trichoderma harzianum* and *T. viride* and three fungicides i.e. Benlate, Ridomil and Dithane M-45, was investigated on the management of Fusarium root rot in okra under screen house conditions. The disease incidence and percent mortality were significantly reduced ($p \leq 0.05$) by all the fungicides and antagonists when compared with untreated check plants. *T. harzianum* and Ridomil increased the yield by 83.6 and 80.2 %, respectively. Under *In vitro* study, Dithane M-45 proved to be more effective than Ridomil and Benlate used alone or integrated with any of the antagonists. Maximum colony diameter of the pathogen (6.9 cm) was recorded in control treatment. *T. viride* was less effective when used alone or with any fungicide, while *T. harzianum* reduced the colony diameter by 43.5 % under *In vitro*.

Introduction

Okra (*Abelmoschus esculentus* L.Moench) locally known as Bhindi, is one of the most important vegetable crops of Pakistan. Okra is apt to the attack of fungi such as *Macrophomina phaseolina*, *Rhizoctonia bataticola*, *R. solani*, *Fusarium solani*, *Pythium butleri*, *Phytophthora palmivora*, *Cercospora abelmoschii* and *Erysiphe cichoracearum* (Mithal, 2006). Fusarium root rot is known to decrease both quantity and quality of major crops including tomato (Parveen *et al.*, 1993) and other vegetables (Ghaffar, 1995), and soybean (Mousa, 1994). The disease is caused by *Fusarium solani* (Rahim *et al.*, 1992). Its incidence has been reported 10-80%, with a maximum (55-80%) in plants grown as kitchen / home gardening and minimum (10-45%) in the crop sown under field conditions. The infected plants are scattered or found in groups when the crop is grown on ridges in field or cultivated on flat beds as kitchen / home gardening. Severely infected plants are found dead, their roots turn dark brown and badly damaged. The fungus perpetuates in the soil or attached to infected plants. Naturally infected seeds of okra appear brown to black and show die-back and root rot symptoms (Mithal, 2006).

Trichoderma harzianum and *T. viride* have been reported to inhibit the mycelial growth of all root rot fungi. Soil infestation with each of the bio-control agents tested reduced the percentage of infected plants and severity of the disease (Mithal, 2006; Faheem *et al.*, 2010). Ghaffar (1993) used different chemicals to control root rot caused by fungi. Seed dressing with Agrosan and Furadan resulted in 38% reduction of *Fusarium* infecting okra. Combination of Benomyl and Captan was effective against all root rot fungi. Of the various bio-agents, *Trichoderma* species have been known to suppress many soil borne fungi under greenhouse and field conditions. *T. harzianum* has been found to antagonize fungal plant pathogen and parasitic nematodes (Saifullah, 1996; Saifullah & Thomas, 1997; Siddiqui *et al.*, 1999).

Trichoderma spp., are present in substantial number in nearly all agricultural soils and decaying wood (Khuls *et al.*, 1999). *Trichoderma* has multiple mechanisms for control of pathogens. It may grow towards hyphae of

other fungi and compete with them for food, space and other resources, coil around them and degrade cell walls of the target fungi, hence limiting the growth and activity and/or by direct consumption of the contents of the target pathogens. Individual strains may produce antibiotics which are harmful for the integrity of the target pathogen (Benitez *et al.*, 2004).

This paper reports the efficacy of different chemical and biological control agents for the management of Fusarium root rot in Okra.

Materials and Methods

Isolation and identification of *Fusarium solani* from dead okra roots: Infected okra seedlings were collected from Taru Jaba, Peshawar. Small pieces of the roots were surface sterilized with 0.1% solution of mercuric chloride for 15-30 seconds, rinsed with sterile distilled water to remove the disinfectant and blotted dry. The treated pieces were placed on Potato Dextrose Agar (PDA) medium in Petri dishes under aseptic conditions and incubated at 25°C for 1 week. Mycelial growth that developed after 7 days was examined. *Fusarium solani* was isolated and subcultured for further studies.

***In vitro* study:** Two species of *Trichoderma*, viz., *T. viride* and *T. harzianum*, obtained from the Plant Pathology Department, Khyber Pakhtunkhwa Agricultural University, Peshawar and three different fungicides viz., Benlate, Ridomil and Dithane M-45 (with different dosages i.e., 100, 200, 300 mg/L) obtained from Agricultural Research Institute Tarnab, Peshawar were tested against *F. solani* alone and in combination with *Trichoderma* spp. The fungicides were incorporated in the medium before pouring into plates. *F. solani* was inoculated at the centre of Petri dish having PDA medium as 1 cm² inoculum plug cut out from 2-week old culture with a sterile scalpel. Inoculum plugs of the same size from the 2-week old culture of the *Trichoderma* spp. were applied at four sites around *F. solani*. The dishes were sealed with parafilm and incubated at 25°C for 15 days. In the control treatment, *F. solani* was allowed to grow alone.

*Corresponding author email: bdulkafi.saifullah@gmail.com

The treatments were;

- T₁= Control (only *F. solani*)
- T₂= *T. harzianum* + *F. solani*
- T₃= *T. viride* + *F. solani*
- T₄= Dithane M 45 (@100 mg/L) + *F. solani*
- T₅= Dithane M 45 (@200 mg/L) + *F. solani*
- T₆= Dithane M 45 (@300 mg/L) + *F. solani*
- T₇= Benlate (@100 mg/L) + *F. solani*
- T₈= Benlate (@200 mg/L) + *F. solani*
- T₉= Benlate (@300 mg/L) + *F. solani*
- T₁₀= Ridomil (@100 mg/L) + *F. solani*
- T₁₁= Ridomil (@200 mg/L) + *F. solani*
- T₁₂= Ridomil (@300 mg/L) + *F. solani*
- T₁₃= *T. harzianum* + *F. solani* + Dithane M 45 (@100 mg/L)
- T₁₄= *T. harzianum* + *F. solani* + Dithane M 45 (@200 mg/L)
- T₁₅= *T. harzianum* + *F. solani* + Dithane M 45 (@300 mg/L)
- T₁₆= *T. harzianum* + *F. solani* + Benlate (@100 mg/L)
- T₁₇= *T. harzianum* + *F. solani* + Benlate (@200 mg/L)
- T₁₈= *T. harzianum* + *F. solani* + Benlate (@300 mg/L)
- T₁₉= *T. harzianum* + *F. solani* + Ridomil (@100 mg/L)
- T₂₀= *T. harzianum* + *F. solani* + Ridomil (@200 mg/L)
- T₂₁= *T. harzianum* + *F. solani* + Ridomil (@300 mg/L)
- T₂₂= *T. viride* + *F. solani* + Dithane M 45 (@100 mg/L)
- T₂₃= *T. viride* + *F. solani* + Dithane M 45 (@200 mg/L)
- T₂₄= *T. viride* + *F. solani* + Dithane M 45 (@300 mg/L)
- T₂₅= *T. viride* + *F. solani* + Benlate (@100 mg/L)
- T₂₆= *T. viride* + *F. solani* + Benlate (@200 mg/L)
- T₂₇= *T. viride* + *F. solani* + Benlate (@300 mg/L)
- T₂₈= *T. viride* + *F. solani* + Ridomil (@100 mg/L)
- T₂₉= *T. viride* + *F. solani* + Ridomil (@200 mg/L)
- T₃₀= *T. viride* + *F. solani* + Ridomil (@300 mg/L)

Each treatment was replicated five times in a Completely Randomized Design (CRD). The data were analyzed by Analysis of Variance (ANOVA) and means were separated by Least Significance Difference (LSD) test. Colony diameter of *F. solani* was measured twice at 7 days interval.

$$\text{Disease Incidence (\%)} = \frac{\text{Total number of infected plants}}{\text{Total number of plants}} \times 100$$

Mortality (%): The percent mortality was recorded in each treatment at the seedling stage by using the following formula;

$$\text{Mortality (\%)} = \frac{\text{Total number of dead plants}}{\text{Total number of plants}} \times 100$$

Growth Parameters and yield plant⁻¹ (g): The plant height, number of fruits and yield plant⁻¹ were recorded on four randomly selected plants. Then averages of that were calculated.

Re-isolation of *F. solani* from affected roots: The affected roots of okra plants were brought to the laboratory and were surface sterilized with 0.1% solution of mercuric chloride for 15-30 seconds, rinsed with sterile distilled water to remove the disinfectant and blotted dry. The treated pieces were then cultured on the PDA medium. Plates were regularly checked for the fungal growth. *Fusarium solani* was confirmed with a mycologist in the department of Plant Pathology.

Pot experiment: Anmol, a local variety of okra was obtained from the market. Six seeds/pot were sown in 15 cm diameter pots. These seeds of okra were treated either with three selected fungicides by dusting method or with two bioagents (*T. harzianum* and *T. viride*). Two days after emergence, each seedling was treated with 10 ml conidial suspension of *Trichoderma* spp. @ 5x10⁵ spores/ml. Inoculum of *F. solani* (one plate per pot) was added at the time of sowing. After two weeks of germination, thinning was done in pots; leaving two plants/pot.

Six treatments were used, each replicated five times in Randomized Complete Block Design (RCBD). The data were statistically analyzed by analysis of variance (ANOVA) and means were separated by Least Significance difference (LSD) test.

The treatments were;

- T₁ = Treated with *F. solani* (Control)
- T₂ = Ridomil (@ 2 g/kg) + *F. solani*
- T₃ = Dithane M 45 (@ 2 g/kg) + *F. solani*
- T₄ = Benlate (@ 2 g/kg) + *F. solani*
- T₅ = *T. viride* + *F. solani*
- T₆ = *T. harzianum* + *F. solani*

Data were recorded on:

Germination (%): Germination percentage of okra was calculated by using the following formula;

$$\text{Germination (\%)} = \frac{\text{Number of germinated seed}}{\text{Total number of seed sown}} \times 100$$

Disease incidence (%): The disease incidence (%) was recorded by using the following formula (Ajmal *et al.*, 2001);

Results and Discussion

In vitro study of fungicides and *Trichoderma* spp., against *Fusarium solani*: Different doses of the three fungicides (Dithane M 45, Benlate and Ridomil) at the selected concentrations and two bioagents (*T. harzianum* and *T. viridae*) alone as well as in combination reduced significantly the growth of *F. solani* (Table 1). Among the antagonists, *T. harzianum* significantly reduced (43.2%) the growth of *F. solani* than *T. viride* (20%).

All the fungicides also significantly affected the growth of *F. solani*. It was 57.9-100% by Dithane M 45, 17.4-100% by Benlate and 55-100% by Ridomil when applied (@100, 200 and 300 mg/L). With increase in concentration, the efficacy of all the fungicides was increased. However, Dithane M 45 showed better performance than others having colony diameter of 2.9 (unit) when applied @ 100 mg/L while Benlate and Ridomil have 5.7 and 3.1 (unit) respectively, at the same concentration. Shah *et al.*, (2006) worked on the effectiveness of different fungicides and reported that Dithane M 45 was best among the tested fungicides in controlling *Fusarium* sp., *In vitro*.

Table 1. Mean colony diameter (cm) of *F. solani* after 7 and 14 days of incubation at 25° C as affected by fungicides and *Trichoderma* spp.

Treatment	Colony diameter (cm)	Decrease than control (%)
T ₁ = Control (only <i>F. solani</i>)	6.9 A	--
T ₂ = <i>T. harzianum</i> + <i>F. Solani</i>	3.9 C	43.2
T ₃ = <i>T. viride</i> + <i>F. Solani</i>	5.5 B	20.0
T ₄ = Dithane M 45 (@100 mg/L) + <i>F. solani</i>	2.9 DE	57.9
T ₅ = Dithane M 45 (@200 mg/L) + <i>F. solani</i>	1.9 HI	71.6
T ₆ = Dithane M 45 (@300 mg/L) + <i>F. solani</i>	0.0 K	100.0
T ₇ = Benlate (@100 mg/L) + <i>F. Solani</i>	5.7 B	17.4
T ₈ = Benlate (@200 mg/L) + <i>F. Solani</i>	3.9 C	43.5
T ₉ = Benlate (@300 mg/L) + <i>F. Solani</i>	0.0 K	100.0
T ₁₀ = Ridomil (@100 mg/L) + <i>F. Solani</i>	3.1 D	55.2
T ₁₁ = Ridomil (@200 mg/L) + <i>F. Solani</i>	2.4 FGH	65.5
T ₁₂ = Ridomil (@300 mg/L) + <i>F. Solani</i>	0.0 K	100.0
T ₁₃ = <i>T.harzianum</i> + <i>F solani</i> + Dithane M 45 (@100mg/L)	1.7 IJ	76.1
T ₁₄ = <i>T. Harzianum</i> + <i>F solani</i> + Dithane M 45 (@200mg/L)	1.4 J	80.0
T ₁₅ = <i>T. harzianum</i> + <i>F. solani</i> + Dithane M 45 (@300 mg/L)	0.0 K	100.0
T ₁₆ = <i>T. harzianum</i> + <i>F. solani</i> + Benlate (@100 mg/L)	3.3 D	52.9
T ₁₇ = <i>T. harzianum</i> + <i>F.</i> + Benlate (@200 mg/L)	2.8 DEF	59.2
T ₁₈ = <i>T. harzianum</i> + <i>F. solani</i> + Benlate (@300 mg/L)	0.0 K	100.0
T ₁₉ = <i>T. harzianum</i> + <i>F. solani</i> + Ridomil (@100 mg/L)	2.4 FGH	65.5
T ₂₀ = <i>T. harzianum</i> + <i>F. solani</i> + Ridomil (@200 mg/L)	2.1 GHI	69.1
T ₂₁ = <i>T. harzianum</i> + <i>F. solani</i> + Ridomil (@300 mg/L)	0.0 K	100.0
T ₂₂ = <i>T. viride</i> + <i>F. solani</i> + Dithane M 45 (@100 mg/L)	2.9 DE	57.9
T ₂₃ = <i>T. viride</i> + <i>F. solani</i> + Dithane M 45 (@200 mg/L)	1.8 IJ	74.5
T ₂₄ = <i>T. viride</i> + <i>F. solani</i> + Dithane M 45 (@300 mg/L)	0.0 K	100.0
T ₂₅ = <i>T. viride</i> + <i>F. solani</i> + Benlate (@100 mg/L)	5.4 B	21.3
T ₂₆ = <i>T. viride</i> + <i>F. solani</i> + Benlate (@200 mg/L)	3.8 C	44.9
T ₂₇ = <i>T. viride</i> + <i>F. solani</i> + Benlate (@300 mg/L)	0.0 K	100.0
T ₂₈ = <i>T. viride</i> + <i>F. solani</i> + Ridomil (@100 mg/L)	3.3 D	52.9
T ₂₉ = <i>T. viride</i> + <i>F. solani</i> + Ridomil (@200 mg/L)	2.5 EFG	63.8
T ₃₀ = <i>T. viride</i> + <i>F. solani</i> + Ridomil (@300 mg/L)	0.0 K	100.0
LSD (0.05)	0.5	
C.V. (%)	32.7	

Means followed by different letters are significantly different from each other at 5 % level of significance

The combinations of antagonists and fungicides showed different performance (Table 1). *T. harzianum* with Dithane M 45 showed better performance and reduced the *R. solani* growth by 76.1-100% (@100, 200 and 300 mg/L) than check. This reduction was 52.9-100 and 65.5-100% when *T. harzianum* + Benlate and *T. harzianum* + Ridomil were applied. *T. viride* also showed poor performance in combination with fungicides. Dithane M 45 (@ 100, 200 and 300 mg/L) and *T. viride* reduced the growth of *F. solani* by 57-100%, while *T. viride* + Benlate and *T. viride* + Ridomil by 21.3-100 and 52.9-100. The combined effect of antagonists and chemicals was more than that of applied alone. Shaban and El-Bramawy (2011) found that the dual application of *Rhizobium* and *Trichoderma* on damping off, root rot diseases and plant growth parameters of some legumes field crop under greenhouse conditions was more effective. This could be due to the better antagonistic characteristics of *T. harzianum* i.e mycoparasitism, antibiosis, competition and production of cell wall degrading enzymes (Chet, 1987, Ghisalberti & Sivasithamparama 1991, Schirmbock *et al.*, 1994). An also of better performance of Dithane M 45 which was also reported by Shah *et al.*, in 2006.

Disease incidence and mortality (%): Keeping in view the results of *In vitro* study the best treatments were

selected and tested for *In vivo* study. Table 2 indicated that response of *Fusarium solani* to fungicides and bio-agents. Among the fungicides, Dithane M 45 was found to be highly effective in reducing the incidence and mortality percentage of *Fusarium* root rot of Okra. The least disease incidence and plant mortality (35.6 and 17.6%) were recorded in treatment where seeds were treated with Dithane M 45 @ 2 g/Kg. This was 41.3 and 69.4% lower than untreated (check). Reduction of incidence of soil borne and seed borne diseases in a number of crops by seed treatment with Vitavax 200 have been reported by Shah *et al.*, (1992). *Trichoderma harzianum* reduced the disease incidence and plant mortality by 41.9 and 27.4% respectively, more efficiently than *T. viride*. It was 30.7 and 52.3% lower than the untreated (check). These results coincide with the results of Rahim & Surrieh (1989), who reported that *Trichodeerma harzianum* was more effective in controlling the plant mortality caused by *F. solani* in Okra, while *T. viride* was less effective. Several species of *Trichoderma* have been reported (Siddiqui *et al.*, 1999) to suppress soil borne disease causing fungi including *Fusarium* sp. The highest incidence and mortality (60.5 & 57.5%) were observed in the diseased (check) plants.

Table 2. Effect of seed treatment with different fungicides and bio-agents on disease incidence and plant mortality on Fusarium root rot of okra in pot experiment.

Treatment	Disease incidence (%)	Decrease than check (%)	Plant mortality (%)	Decrease than check (%)
T ₁ . Untreated (Check)	60.5 A	--	57.5 A	--
T ₂ . Ridomil @ 2g/kg seed	41.7 B	31.1	37.5 B	34.8
T ₃ . Dithane M 45 @ 2g/kg seed	35.6 B	41.1	17.6 D	69.4
T ₄ . Benlate @ 2g/kg seed	38.3 B	36.7	37.5 B	34.8
T ₅ . <i>T. viridei</i>	44.6 B	26.3	32.6 BC	43.4
T ₆ . <i>T. harzianumi</i>	41.9 B	30.6	27.4 C	52.3
LSD (0.05)	9.1		6.7	
C.V. (%)	17.3		15.8	

Means followed by different letters are significantly different from one another at 5 % level of probability

With increase in disease incidence, the plant mortality (%) also increased. However, the lowest disease incidence was found in plants treated with Dithane M 45 followed by Benlate. Moreover, the differences between the two were non-significant. These fungicide(s) inhibit growth and multiplication of the *F. solani* (Agrios, 1997). Bioagents also played similar role in inhibiting the *F. solani*. *Trichoderma harzianum* was found more efficient than *T. viride*.

Germination (%): Results presented in Table 3 revealed that the germination percentage was significantly affected by different fungicides and antagonists. Seeds treated with Dithane M 45 showed high percentage (93.3%) of germination than others. This was 21.6% higher than

untreated (check). These results are in line with the findings of Dash & Narain (1996). They reported that pretreatment of seed with fungicide improve seed germination. Seed treatment with bio-agents i.e., *T. harzianum* and *T. viride* also showed their performance in improving germination. Both the bio-agents were non-significant between themselves in term of germination percentage.

However, *T. harzianum* and *T. viride* increased germination percentage by 13.4 and 15.1% than untreated (check). *Trichoderma* species are capable of hyperparasitising the pathogenic fungi and found to involve in protecting number of crop plants (Durrell, 1968; Barnett & Binder, 1973). With increase in disease incidence, germination percentage is decreased.

Table 3. Effect of different fungicides and bio-agents on germination of okra seeds as affected Fusarium root rot in pot experiment.

Treatment	Germination (%)	Increase over check (%)
T ₁ . Untreated (Check)	76.7 B	--
T ₂ . Ridomil @ 2g/kg seed	90.0 A	17.4
T ₃ . Dithane M 45 @ 2g/kg seed	93.3 A	21.7
T ₄ . Benlate @ 2g/kg seed	91.7 A	19.6
T ₅ . <i>T. Viridei</i>	88.3 A	15.2
T ₆ . <i>T. Harzianumi</i>	87.0 A	13.4
LSD (0.05)	10.1	
C.V. (%)	9.6	

Means followed by different letters are significantly different from one another at 5 % level of probability

Growth parameters and yield plant⁻¹: Data presented in Table 4 indicated the effect of different fungicides and bio-agents on the growth and yield of okra. Maximum plant height (77.7 cm), number of fruits (30.6) and yield (340.8 g) were recorded in plants raised in pots treated with *T. harzianum*. This was 45.8, 102.6 and 83.6% higher than untreated (check). The shortest plant (53.3 cm), lower number of fruits/plant (15.1) and low yield/plant⁻¹ (185.6 g) were recorded in treatment having no fungicide and bio-agents. Among fungicides Dithane M 45 showed better performance. Treated plants were 24.6% taller, having 73.2% higher number of fruits/plant and produced 80.2% greater yield/plant than untreated (check). However, the bio-agent (*T. harzianum*) showed

dominancy over fungicides under screen house conditions. Siddiqui *et al.*, (2000) reported that *T. harzianum* elevated plant height in okra and such results were also obtained against *F. solani* in sunflower by Waffa & Amin (2001) that *Trichoderma* species controlled *F. solani* and increased the plant growth parameters.

Bio-agents have remarkable capacity of multiplication, thus, when applied; they multiply in exponential ratio and even can overcome stress condition by forming thick walled spores. Hence, bioagents are the solution for safer environmental issues and needs proper attention for seed treatment (Bharath *et al.*, 2005).

Table 4. Effect of seed treatment with different fungicides and bio-agents on growth parameters and yield plant⁻¹ of okra in pot experiment.

Treatment	Growth parameters		
	Plant height (cm)	No. of fruits plant ⁻¹	Yield plant ⁻¹ (g)
T ₁ . Untreated (Check)	53.3 C (--)	15.1 D (--)	185.6 C (--)
T ₂ . Ridomil @ 2g/kg seed	61.9 BC (16.1)*	26.8 B (77.5)*	231.6 B (24.8)*
T ₃ . Dithane M 45 @ 2g/kg seed	66.4 B (24.6)	26.3 B (73.2)	334.4 A (80.2)
T ₄ . Benlate @ 2g/kg seed	63.5 BC (19.1)	20.1 C (33.1)	271.2 B (46.1)
T ₅ . <i>T. Viridei</i>	63.1 BC (18.4)	24.3 B (60.9)	272.1 B (46.6)
T ₆ . <i>T. Harzianumi</i>	77.7 A (45.8)	30.6 A (102.6)	340.8 A (83.6)
LSD (0.05)	10.4	3.6	42.6
C.V. (%)	13.5	12.7	12.9

Means followed by different letters are significantly different from one another at 5 % level of probability

*Percent increase over untreated (check)

References

- Agrios, G.N. 1997. Plant Pathology, 5th Edition. Academic Press, San Diego and London. pp. 922
- Ajmal, M., S. Ahmad and S. Hussain. 2001. Effect of soil moisture on black scurf disease and yield of potato. *Pak J. Biol. Sci.*, 4: 150-151.
- Barnett, H.L. and H.A. Binder. 1973. The fungal host parasite relationship. *Ann. Rev. Phytopathol.*, 11: 273-292.
- Benitez, T., A.M. Rincon., M. Carmenlimon and A.C. Condon. 2004. Biocontrol mechanisms of *Trichoderma* strains. *Int. Micro.*, 7: 249-260.
- Bharath, B.G., S. Lokesh and H.S. Shetty. 2005. Effects of fungicides and bioagents on seed mycoflora, growth and yield of watermelon. *Integ. Biol. Sci.*, 9: 75-78.
- Chet, I. 1987. *Trichoderma* – application, mode of action and potential as a biocontrol agent of soil borne plant pathogenic fungi, pp. 137-160. In: *Innovative approaches to plant disease control*. (Ed.): I. Chet. John Wiley and Sons, New York.
- Dash, S.K. and A. Narain. 1996. Efficacy of selected fungicides on seed-borne fungi and on percentage of germination of diseased seeds of crops. *Crop Res. Hisar.*, 11: 207-211.
- Durrell, L.W. 1968. Hyphal invasion by *Trichoderma viride*. *Mycopathol Mycol. Appl.*, 35: 138-144.
- Faheem, A., V.K. Razdan, F. A. Mohiddin, K. A. Bhat and S. Bandy. 2010. Potential of *Trichoderma* species as biocontrol agents of soil borne fungal propagules. *J. Phytology*, 2(10): 38-41.
- Ghaffar, A. 1993. Rhizobia as biocontrol organisms. *BNF Bulletin*, 12: 6-4.
- Ghaffar, A. 1995. Biological control of root rot and root knot disease complex of vegetables. Final Research Report, Dept. Bot. Univ. Karachi, Karachi, pp. 752-770.
- Ghisalberti, E.L. and K. Sivasithamparam. 1991. Antifungal antibiotics produced by *Trichoderma* spp. *Soil Biol Biochem.*, 23: 1011-1020.
- Khuls, K., E. Leeckfeldt., T. Borner and E. Gueho. 1999. Molecular reidentification of *Trichoderma* isolates. *Med. Mycol.* 37: 25-33.
- Mithal, M.J. 2006. Low cost and pollution free technology against root rot of okra. www.pakistan.com.
- Mousa, E.S.M. 1994. Biological management of soil-borne pathogens and root-knot nematode complexes on soybean. Proceedings of the second Afro-Asian Nematology Symposium held at Menoufiya, Egypt, 18-22 December, pp. 60-66.
- Parveen, S., S.E. Haque and A. Gaffar. 1993. Biological control of *Meloidogyne javanica* on tomato and okra in soil infested with *Fusarium oxysporium*. *Pak. J. Nematol.*, 11: 151-156.
- Rahim, A.M. and A.A. Surrieh. 1989. Biological control of *Rhizoctonia solani* the causal agent of seedling blight in okra. *Arab J. Plant Prot.*, 7: 167-171.
- Rahim, A.M., K.D. Aziza, A.M. Tarabeih and A.A.M. Hassan. 1992. Damping-off and root rot of okra and table beet with reference to chemical control. *Assiut J. Agric. Sci.*, 23: 19-36.
- Saifullah, 1996. Fungal Parasitism of young females of *Globodera rostochiensis* and *G. pallida*. *Afro-Asian J. Nematol.*, 6: 17-22.
- Saifullah. and B.J. Thomas. 1996. Studies on the parasitism of *Globodera rostochiensis* by *Trichoderma harzianum* using Low Temperature Scanning Electron Microscopy. *Afro-Asian J. Nematol.*, 6: 117-122.
- Shaban, W.I. and M.A. El-Bramawy. 2011. Impact of dual inoculation with *Rhizobium* and *Trichoderma* on damping off, root rot diseases and plant growth parameters of some legumes field crop under greenhouse conditions. *Int. Res. J. Agric. Sci. Soil Sci.*, 1(3): 098-108 May 2011.
- Shah, M.I., P. Sultan, A. Nasir, P. Williams, A. Jan, M. Sajad, S. Rehman and A.S. Shawal. 2006. *In vitro* study on effect of some fungicides viz., carbendazim, mancozeb, conjoint carbendazim, mancozeb and sulphur against *F. oxysporium*. *Res. J. Micro.*, 1(4): 360-365.
- Schirmböck, M., M. Lorito, Y.L. Wang, C.K. Hayes, A.I. Arslan, F. Scala, G.E. Harman and C.P. Kubicek. 1994. Parallel formation and synergism of hydrolytic enzymes and peptidol antibiotics: molecular mechanisms involved in the antagonistic action of *T. harzianum* against phytopathogenic fungi. *Appl. Environ. Microbiol.*, 60: 4364-4370.
- Shah, S.J., A.S. Hussain, S.A. Ali and I. Khattak. 1992. Efficacy of different seed dressing fungicides on recovery of seed borne mycoflora of soybean. *Sarhad J. Agric.*, 8: 683-687.
- Siddiqui, I.A., S.E. Haque and A. Ghaffar. 1999. Root dip treatment with *Pseudomonas aeruginosa* and *Trichoderma* spp., in the control of root rot-root knot disease complex in chili. (*Capsicum annum* L.) *Pak. J. Nematol.*, 17: 67-75.
- Siddiqui, I.A., S.E. Haque, M.J. Zaki and A. Ghaffar. 2000. Greenhouse evaluation of *Rhizobia* as biocontrol agent of root-infecting fungi in okra. *Acta Agrobotanica*, 53: 13-22.
- Waffa, M.H. and A.W. Amin. 2001. Efficiency of *Trichoderma* species on control of *Fusarium*-rot, root knot and reniform nematodes disease complex on Sunflower. *Pak J. Biol. Sci.*, 4(3): 314-318.