ORGANIC AND INORGANIC FERTILIZERS ALONG WITH THUJA ORIENTALIS L. FOR THE CONTROL OF ROOT INFECTING FUNGI

SHAHNAZ DAWAR^{1*}, MARIUM TARIQ² AND HIRA SARFARAZ¹

¹Department of Botany, University of Karachi, Pakistan ²M.A.H.Qadri Biological Research Centre, University of Karachi, Pakistan. *Corresponding author's e-mail: shahnaz_dawar@yahoo.com

Abstract

Macrophomina phaseolina (Tassi) Goid, *Rhizoctonia solani* Kuhn and *Fusarium* species Schlecht are reported to be associated with wilting and root rot diseases in different crop plants all over the world. To improve crop production, fertilizers like cow dung, goat dung, urea and Di Ammonium Phosphate (DAP) with doses of 0.01 and 0.1 % w/w were mixed in soil in combination with seed treatment with stock solution of *Thuja orientalis* L. plant at 100% (w/v). Results revealed that maximum improvement in growth and reduction in colonization of root infecting fungi like *M. phaseolina*, *R. solani* and *Fusarium* spp., were recorded when 0.1 and 0.01% goat dung was incorporated in soil in combination with seed treatment with *T. orientalis* extract on cowpea and mash bean plants.

Key words: Fertilizers with different doses, Cowpea and mash bean plants, Aqueous extract of *T. orientalis*, Root infecting fungi.

Introduction

Thuja orientalis L. belongs to the family cupressaceae, commonly known as morpankhi. Their leaves contain essential oil like camphor, fenchone, isothujone, thujone, mono and sesquiterpenes which includes carbohydrate, phenol, alcohols, ether, aldehyde and ketone are used in different activities like treatment of fungal infection, cancer, moles, as antipyretic, antitussive, astringent, diuretic, refrigerant and stomachic (Sokovic et al., 2010; Yeung, 1985). The chemical constituents of T.orientalis leaves contained rhodoxanthin, amentoflavone, hinokiflavone, quercetin, myricetin, carotene, xanthophylls, ascorbic acid and thujone which is found naturally in two diastereomeric forms: α -thujone and β - thujone (Perry *et al.*, 1999). Nowadays, plant extracts with their phytochemical constituents are getting interest due to have potential sources against microbial, viral inhibitors and for human health (Jasuja et al., 2012). Great biological activity against microorganisms have been reported due to antioxidant compounds and bioactive constituents of plants like tannins, flavonoids, saponins, terpenoids and alkaloids which provide protection against diseases of free- induced oxidative stress (Sulaiman et al., 2011; Mandalari et al., 2007; Avato et al., 2006; Funatogawa et al., 2004; Navarro & Delgado, 1999).

The most common root infecting fungi like Fusarium species, Macrophomina phaseolina, Rhizoctonia solani causes various complex root rot and wilt diseases on many leguminous and non-leguminous crops which results in the death of plants (Hassan et al., 2014; Nancy et al., 1997). Fusarium spp., causing stem rot, root rot and wilt diseases produced economic losses on a wide variety of crop plants (Ploetz et al., 2007; Larkin & Fravel, 1998). F. oxyporum have association with endomycorrhizal organisms causes wilt in banana, cotton, sweet potatoes, tomato, asparagus, muskmelon, cantaloupe and ginger (Stover, 1962). M. phaseolina produces infection in all stages of plant growth initiated from soil, seed and infected plant debris (Reuveni et al., 1983). M. phaseolina found in soil produced sclerotia which remain dormant and under suitable environmental condition produce hyphae which cause infection in the roots of host plant (Clare et al., 2010; Ammon et al.,

1974). Charcoal rot, seedling blight, pod rot, root rot and stem rot diseases are caused by *M. phaseolina* resulting in the death of seedling due to the blockage of xylem tissues (Srivastava *et al.*, 2001). *R. solani* Kuhn is also a root rot, facultative plant pathogen which causes seed rot, wilting, root rot, damping off of seedling, black scurf of potatoes, bare patch of cereals, root rot of sugar beet, belly rot of cucumber and sheath blight of rice (Bolton *et al.*, 2010; Godoy-Lutz *et al.*, 2008).

Organic manures are cheap and ecofriendly source for the improvement of crop production in low-input agriculture. Use of plant along with organic and inorganic fertilizers has gained importance in sustainable crop production (Abdullahi & Sheriff, 2013). Mixing of organic manures in soil gave positive effects on physical and biochemical properties of soil. Many research work has confirmed efficacy of organic and inorganic fertilizers mixed in soil shows promising result in improvement of plant growth and effectively suppress certain soil borne plant pathogens (Tariq *et al.*, 2008; Abdullahi *et al.*, 2014; Ikram & Dawar, 2015).

The objectives of this study was to investigate the efficacy of *T. orientalis* as seed treatment, along with different fertilizers for the control of diseases caused by *M. phaseolina, R. solani* and *Fusarium* spp., on different crop plants.

Materials and Methods

Plant material and fertilizers collection: *Thuja orientalis* L. plant was collected from Department of Botany, University of Karachi. Plant was washed with tap water followed by sterilized distilled water to avoid dusts and other unwanted materials from the natural environment. These dust free plant was allowed to dried in shade for 2 weeks and then powdered using an electric grinder. This plant powder (10 g) dissolved in 100 mL sterilized distilled water, left overnight to allow constituents to get dissolved in water. Extract was filtered using Whatman's No.1 filter paper and the residue obtained was centrifuge at 500 rpm for 15 minutes and the supernatant was taken as stock solution. In our previous work seed treatment with 100, 75 and 50% of *T. orientalis* was used, of which 100%

concentration for 10 minutes gave better result in improvement of crop plants so selected for further studies (Sarfaraz, 2016). Inorganic and organic fertilizers like Di Ammonium Phosphate (DAP), urea was purchased from local market while cow dung and goat dung obtained from cattle farm of Karachi.

Screen house experiment: Pot experiment was set in a screen house of Botany Department, University of Karachi where sandy loam soil (300 g) was placed in pots having pH ranged from 7-7.6 with moisture holding capacity (MWHC) of 23.12% (Keen & Raczkowski, 1922), total nitrogen 1.5% (Mackenzie and Wallace, 1954). Natural soil contained 4-6 sclerotia/g of M. phaseolina (Sheikh & Ghaffar, 1975), 6-9% colonization of R. solani on sorghum seeds used as baits (Wilhelm, 1955) and 3700 cfu g-¹Fusariumspp., (Nash & Snyder, 1962). Seeds of mash bean and cowpea treated with stock solution of T. orientalis for 10 minutes and dried on blotter paper. Soil was mixed with fertilizers (DAP, urea, cow dung and goat dung) at rate of 0.01 and 0.1 % w/w separately, transferred in pots containing 300 g/pot and 4 treated seeds of mash bean and cowpea were sown in each pot separately. Concentrations of fertilizers were selected on basis of previous work by different research which showed that highest dose of fertilizers gave fatal effect on plant growth (Kumar et al., 2005; Sikander et al., 2009). A set of seeds treated with sterilized distilled water was also kept as control. The experiment was performed on screen house bench in randomized complete block designed with three replicates of each treatment. For comparison, control set was also kept in which untreated seeds and soil without fertilizer were used. After thirty days of germination of seeds, plant length weight, number of nodules and number of leaves were observed. Colonization of root rot fungi was recorded by washing roots with tap water and surface sterilized with 1% sodium hypochloride and roots were placed on Petri plates containing Potato Dextrose Agar supplemented with antibiotics (Kanwal et al., 2017; Rafi et al., 2016).

Analysis of data: Data of growth parameters and colonization % of root infecting fungi were analyzed using ANOVA and means were compared by least significance difference at 5 % probability level (Sokal & Rohlf, 1995).

Results

All fertilizers used in this research gave maximum enhancement in growth parameters like shoot length, shoot weight, root length, root weight, number of nodules and leaves, while root infecting fungi were significantly reduced on mash bean plant. Shoot length was enhanced when soil was mixed with cow dung at 0.1% (w/w) in combination with seeds treatment by T. orientalis stock solution. Similarly, shoot and root weight were increased significantly (p<0.001) when soil was amended with DAP at 0.01% (w/w) along with seeds treated with T. orientalis extract. There was improvement in root length (p<0.01) and number of nodules when goat dung applied in soil in combination with seeds treatment with T. orientalis extract (p<0.01). Increased number of leaves were recorded when soil was amended with cow dung @ 0.01% (w/w) in combination with seed treatment with T. orientalis extract. Complete reduction of R. solani (p<0.05) was observed when cow dung at 0.1 % w/w was used along with seeds treated with T. orientalis extract while significant reduction of *Fusarium* spp., (p<0.05) and *M. phaseolina* were recorded when goat dung at 0.1 % w/w was used in combination with seed treatment with *T. orientalis* (Table 1).

In case of cowpea, shoot length and root length (p<0.001) were increased significantly (p<0.05) when soil was mixed with goat dung at 0.01% w/w in combination with seeds treated with T. orientalis extract. Shoot weight was significantly (p<0.01) increased when soil was mixed with DAP at 0.1% w/w and seeds treated with extract in contrast to control. Similarly, root weight was significantly (p < 0.05) increased when seeds were treated with T. orientalis extract in combination with soil amendment with urea at 0.01% (w/w). Maximum number of nodules (p<0.01) and leaves were recorded when soil was mixed with cow dung 0.1% (w/w) in addition with treated seeds as compared to control. Goat dung and cow dung at 0.01 and 0.1% w/w was observed to best in reduction of Fusarium spp., R. solani (p<0.001) and M. phaseolina (p<0.001) colonization on cowpea roots (Table 2).

Cow dung and goat dung at 0.1 %, 0.01 % gave much better results in improvement of mash bean and cowpea plants when used in combination with *T. orientalis* extract used for seed treatment followed by DAP and urea as well as reduced the colonization of root infecting fungi particularly *R. solani*.

Discussion

Goat and cow dung considered as valuable fertilizer used for centuries in providing necessary nutrients like micronutrients for crop growth. They are extensively used by growers for the production of different crops due to presence of medicinal properties according to unani or ayurvediic medicine. Presently cow dung at 0.1% and goat dung at 0.01% w/w in combination with seeds treatment with stock solution of T. orientalis extract were found to enhanced the plant growth and showed inhibition of root infecting fungi like Fusarium spp., R. solani and M. phaseolina followed by DAP and urea. In cowpea, when soil was amended with goat dung and cow dung @ 0.01% (w/w) and urea 0.01% (w/w) in combination with 100% (w/v) extract of T. orientalis, significant increment in growth parameters and reduction in the incidence of root infecting fungi were recorded. Previous observations showed improvement in growth of Zea mays, Abelmoschus esculentus, Oryza sativa, Luffa cylindrical and Lycopersicon esculentum and reduced pathogenic fungi like Colletotrichum capsici, Sclerotium rolfsii, Alternaria alternata, Penicillium species, R. solani, Phytophthora palmivora, Helminthosporium species by the use of cow dung and cow urine (Nautiyal et al., 2006). Cow dung showed much effectiveness against conidial germination and mycelial growth of Fusarium oxysporum and F. solani on cucumber plants (Basak & Lee, 2001; Basak et al., 2002). Cow dung coated seeds showed more survival, improved shoot length, dry weight and number of nodules as compared to non cow dung treated plants (Nautiyal et al., 2013). Present results showed that root pathogens were successfully reduced by the addition of cattle manure while R. solani colonization was completely inhibited in mash bean when used in combination with extract of T. orientalis. Seeds coated with cow dung and sown in the presence of mixture of wilt complex fungi treated soil could reduce cell wall degrading enzymes activities produced by plant roots against pathogens (Nautiyal et al., 2013).

ts.
an
d
an
þe
sh
ma
Ę
ž
Ĩ
Ę
Ľ0
ot
2
of
0
ntr
00
je
t
d in
Du
h a
wt
5
50
0
act
Ë,
eX
ns
eo
Ð
S a
ali
m
'n
Τ. 6
_
vith
20
U.
al
L S
ize
ij
fer
t
re
diffe
dif
of
ect
ΕŲ
ğ
ine
ē
0m
0
[able1 .
able
Ë

	Tre	Treatments			Growth parameters	rameters			Colonization	Colonization % of root infecting fungi	fecting fungi
Fertilizers	Concentrations (%)	Methods	Shoot length (cm)	Shoot weight (g)	Root length (cm)	Root weight (g)	No. of nodules	No. of leaves	F. oxysporum	R. solani	M. phaseolina
	0	 	21.64	0.44	11.68	0.05	2.33	5.66	90.66	30	30
	10.0	Treated seeds with T. orientalis	25.14	0.34	18.66	0.04	5.22	8.22	43.33	26.67	20
	10.0	Non treated seeds	24.16	0.38	16.75	0.14	6.55	4.44	76.66	6.67	13.33
Cow dung	-	Treated seeds with T. orientalis	25.21	0.17	12.17	0.02	5.55	6.33	46.67	0	26.67
	1.0	Non treated seeds	22.5	0.47	21.97	0.18	8.66	6.22	86.77	23.33	20
	10.0	Treated seeds with T. orientalis	23.11	0.45	14.51	0.06	1.88	5.77	50	30	16.67
Sand ter	10.0	Non treated seeds	24.28	0.56	19.14	0.21	6.88	6.33	80	26.67	46.67
Coat dung	-	Treated seeds with T. orientalis	23.56	0.72	16.91	0.51	9.22	6.55	33.33	16.67	10
	1.0	Non treated seeds	21.41	0.32	10.24	0.09	2.44	5.44	83.33	18.33	13.33
	10.0	Treated seeds with T. orientalis	21.78	0.46	14.61	0.08	0.33	5.44	46.67	6.67	16.67
11,000	10.0	Non treated seeds	20.58	0.52	11.07	0.31	0.77	5.38	63.33	20	20
0164	10	Treated seeds with T. orientalis	23.7	0.22	23.84	0.05	3.88	5.77	60	43.33	20
	1.0	Non treated seeds	19.1	0.41	6.03	0.14	0	5.88	66.67	6.67	16.67
	10.0	Treated seeds with T. orientalis	22.44	0.17	13.77	0.01	1.66	6.22	53.33	53.33	13.33
	10.0	Non treated seeds	22.61	09.0	15.64	0.12	4.11	7.22	73.33	36.67	33.33
DAT.	10	Treated seeds with T. orientalis	26.73	0.22	10.84	0.03	1.66	6.55	50	40	3.33
	1.0	Non treated seeds	20.0	0.57	6.48	0.17	0.11	6.77	53.33	20	20
	Concentrations		1.60	0.10	2.77	0.68	1.74	0.86	9.62	11.72	7.14
$LSD_{0.05}$	Methods		1.85	0.12	3.20	0.78	2.01	0.99	11.11	13.54	8.24
	Fertilizers		1.30	0.08	2.26	0.55	1.42	0.70	7.85	9.57	5.83

	Trea	Treatments Colonization % of root infecting			Growth p	Growth parameters			Colonization	Colonization % of root infecting fungi	fecting fungi
Fertilizers	Concentrations (%)	Methods	Shoot length (cm)	Shoot weight (g)	Root length (cm)	Root weight (g)	No. of nodules	No. of leaves	F. oxysporum	R. solani	M. phaseolina
'	0		31.91	1.31	9.3	0.19	0.88	5.88	99.96	53.33	43.33
		Treated seeds with T. orientalis	31.78	2.19	8.77	0.44	1.11	6.44	70	10	13.33
	10.0	Non treated seeds	34.73	1.80	7.61	0.45	0.88	5.11	86.66	20	19.63
Cow dung		Treated seeds with T. orientalis	37.84	2.15	10.46	0.26	2.22	6.22	73.33	3.33	6.67
	1.0	Non treated seeds	32.36	2.08	11.54	0.64	2.66	5.44	83.33	22.67	16.66
		Treated seeds with T. orientalis	37.91	2.07	16.14	0.26	5.22	7.22	50	13.33	3.33
	10.0	Non treated seeds	36.5	2.27	14.17	0.62	2.00	6.00	66.66	23.33	13.36
Goat dung	÷	Treated seeds with T. orientalis	37.53	2.18	13.45	0.29	1.77	7.00	80	20	30
	1.0	Non treated seeds	32.74	2.36	15.97	0.61	0.44	6.36	83.33	10	13.33
		Treated seeds with T. orientalis	32.95	1.97	12.62	1.51	0.77	6.44	70	3.33	16.67
	10.0	Non treated seeds	32.77	2.63	13.77	0.30	0	4.55	72.66	13.33	16.66
OICa		Treated seeds with T. orientalis	32.25	1.95	12.74	0.35	0.44	6.22	76.67	23.33	3.33
	0.1	Non treated seeds	29.62	1.54	10.84	0.37	0.55	6.22	70	20	20
		Treated seeds with T. orientalis	36.76	2.10	13.88	0.33	1.00	7.01	66.67	10	33.33
	10.0	Non treated seeds	33.71	2.27	14.83	0.82	1.33	9	50	10	10
DAF	÷	Treated seeds with T. orientalis	29.58	2.38	8.37	0.97	0.22	6.11	66.60	10.67	13.33
		Non treated seeds	28.62	1.98	8.61	0.56	0	5.11	50	16.67	26.66
	Concentrations		1.81	1.01	1.45	1.43	09.0	0.69	36.21	6.26	5.68
$LSD_{0.05}$	Methods		2.09	1.16	1.68	1.65	0.69	0.80	41.81	7.23	6.56
	Fertilizers		1.48	0.82	1.18	1.17	0.49	0.56	29.56	5.11	4.63

1256

Cattle manure is made up of digested grass, grain and rich contents of nitrogen, phosphorus and potassium which is helpful in eliminating harmful ammonia gas and pathogens. By use of manure not only moisture holding capacity improved but also helpful in addition of generous amount of organic matter in soil (Phipps, 2016; Srivastava *et al.*, 2010). Necessary nutrients and pest repellent properties are obtained from cow dung while urine contain micronutrients and increase fertility helpful in providing food which was free from hazards of chemical fertilizers and pesticides (Chauhan, 2000). According of Ebenezer *et al.*, (2012) goat dung extract with concentrations (1.0 and 5.0%) was helpful in completely reduction of *Sclerotium rolfsii* growth on oil palm plantation.

Results of the present study clearly suggest that organic fertilizers like cow and goat dung at 0.1 and 0.01% inhibited the growth of root rot pathogens. DAP and urea also reduced the fungal pathogens on roots but less activity was recorded as compared to cow and goat dung. It can be therefore be suggested that both dungs in combination with seed treatment with *T. orientalis* aqueous solution could be useful in the growth and development of crop plants with the same time enriching soil and provide valuable nutrients in growth promotion.

References

- Abdullahi, R. and H.H. Sheriff. 2013. Effect of Arbuscular mycorrhizal fungi and chemical fertilizer on growth and shoot nutrients content of onion under field condition in Northern Sudan Savanna of Nigeria. J. Agri. Veter. Sci., 3(5): 85-90.
- Abdullahi, R., H.H. Sheriff and A. Buba. 2014. Effect of biofertilizer and organic manure on growth and nutrients content of pearl millet. ARPN J. Agri. Biol. Sci., 9(10): 351-355.
- Ammon, V., T.D. Wyllie and M.F. Brow. 1974. An ultrastructural investigation of pathological alterations induced by *Macrophomina phaseolina* (Tassi) Goid. in seedlings of soybean, *Glycine max* (L.). Merril, *Physiol. Plant Pathol.*, 4: 1-4.
- Avato, P., R. Bucci, A. Tava, C. Vitali, A. Rosato, Z. Bialy and M. Jurzysta. 2006. Antimicrobial activity of saponins from *Medicago* sp. structure-activity relationship. *Phytother*. *Res.*, 20: 454-457.
- Basak, A.B. and M. W. Lee. 2001. Efficacy of cow dung in controlling root rot and *Fusarium* wilt disease of cucumber plants. Abstract published in the 2001 Korean Society of Plant Pathology Annual meeting and International Conference, held on the 25-30th October, Kyongju, Korea. pp. 49.
- Basak, A.B., M.W. Lee and T.S. Lee. 2002. *In vitro* inhibitory activity of cow urine and dung to *Fusarium solani* f. sp. *cucurbitae*. *Microbiology*, 30(1): 51-54.
- Bolton, M.D., L. Panella, L. Campbell and M.F. Khan. 2010. Temperature, moisture, and fungicide effects in managing *Rhizoctonia* root and crown rot of sugar beet. *Phytopathology*, 100: 689-697.
- Chauhan, R.S. 2000. Panchgavya dwara prakritik chikitsa. *Kheti Sansar*, 3: 21-23.
- Clare, M.M., R. Melis, J. Dereta, M. Laing and R.A. Buruchara. 2010. Identification of sources of resistance to *Fusarium* root rot among selected common bean lines in Uganda. J. *Anim. Plant Sci.*, 7: 876-891.

- Ebenezer, O., G.C. Clerk and J.K. Mensah. 2012. Effect of dung of goat in oil palm plantations on the growth of soil facultative parasite *Sclerotium rolfsii*. *Int. J. Plant Anim.& Environ. Sci.*, 2(1): 202-208.
- Funatogawa, K., S. Hayashi, H. Shimomura, T. Yoshida, T. Hatano, H. Ito and Y. Hirai. 2004. Antibacterial activity of hydrolyzable tannins derived from medicinal plants against *Helicobacter pylori. Microb. Immunol.*, 48: 251-261.
- Godoy-Lutz, G, S. Kuninaga, J.R. Steadman and K. Powers. 2008. Phylogenetic analysis of *Rhizoctonia solani* subgroups associated with web blight symptoms on common bean based on ITS-5.8S rDNA. J. Gen. Plant Pathol., 74: 32-40.
- Hassan, N., M. Shimizu and M. Hyakumachi. 2014. Occurrence of root rot and vascular wilt diseases in Roselle (*Hibiscus* sabdariffa L.) in Upper Egypt. *Microbiology*, 42(1): 66-72.
- Ikram, N. and S. Dawar. 2015. Combine effects of inorganic fertilizers and plant extracts on growth and in the control of root rot fungi on crops plants. *Int. J. Bio. Biotech.*, 12(2): 215-222.
- Jasuja, N.D., R. Saxena, S. Chandra and R. Sharma. 2012. Pharmacological characterization and beneficial uses of *Punica granatum. Asain J. Plant Sci.*, 11: 251-267.
- Kanwal, S., S. Dawar, M. Tariq and F. Imtiaz. 2017. Gamma radiation technetium-99M (TC-99M) exposure on crop seeds for management of soil borne root infecting fungi and growth promotion of mash bean (*Vigna mungo L.*), chick pea (*Cicer arietinum L.*) and sunflower (*Helianthus annuus L.*). *Pak. J. Bot.*, 49(2): 763-768.
- Keen, B.A. and H. Rakzowski. 1922. The relation between clay content and certain physical properties of soil. *J. Agric. Sci.*, 11: 441-449.
- Kumar, D., K.P. Singh and R.K. Jaiswal. 2005. Effect of fertilizers and neem cake amendment in soil on spore germination of *Arthrobotrys dactl. Microbiol.*, 33(4): 194-199.
- Larkin, R.R. and D.R. Fravel. 1998. Efficiency of various fungal and bacterial biocontrol organisms for control of *Fusarium* wilt of tomato. *Plant Disea.*, 82: 1022-1028.
- Mackenzie, H.A. and H.S. Wallace. 1954. The Kjeldahl determination of nitrogen: A critical study of digestion conditions, temperature, Catalyst and oxidizing agents. *Aust. J. Chem.*, 7:55-70.
- Mandalari, G., R.N. Bennett, G. Bisignano, D. Trombetta, A. Saija, C.B. Faulds, M.J. Gasson and A. Narbad. 2007. Antimicrobial activity of flavonoids extracted from bergamot (*Citrus bergamia* Risso) peel, a byproduct of the essential oil industry. J. Appl. Microb., 103: 2056-2064.
- Nancy, W., D.E. Mathre, B. James and S. Charles. 1997. Biological seed treatments: Factors involved in efficacy. *Hort. Sci.*, 32: 179-183.
- Nash, S.M. and W.C. Snyder. 1962. Quantitative estimation by plate counts of propagules of the bean root rot *Fusarium* in field soils. *Phytopathology*, 52: 567-572.
- Nautiyal, C.S., S. Mehta, H.B. Singh and P. Pushpangadan. 2006. Go mutra and dung based products as viable alternative for sustainable agriculture for our rural marginal farmers. *The Ind. Cow*, 2(9): 13-21.
- Nautiyal, C.S., S. Srivastava, S. Mishra, M.H. Asif, P.S. Chauhan, P. C. Singh and P. Nath. 2013. Reduced cell wall degradation plays a role in cow dung-meiated management of wilt complex disease of chickpea. *Biol. & Fertility of Soils*, 49(7): 881-891.
- Navarro, V. and G. Delgado. 1999. Two antimicrobial alkaloids from *Bocconia arborea*. J. Ethnopharmacol., 66: 223-226.
- Perry, N.B., R.E. Anderson, N.J. Brennan, M.H. Douglas, A.J. Heaney, J.A. McGimpsey and B.M. Smallfield. 1999. Essential oils from Dalmatian Sage (*Salvia officinalis* L.): Variations among individuals, plant parts, seasons, and sites. J. Agri. Food Chem., 47(5): 2048-2054.

- Phipps, N. 2016. Cow dung fertilizer: learn the benefits of cow manure compost. http://www.gardeningknowhow.com/ composting/manures/cow-manure-compost.htm
- Ploetz, R.C., A.J. Palmateer, D.M. Geiser and J.H. Juba. 2007. First report of *Fusarium* wilt caused by *Fusarium* oxysporum on roselle in the United States. *Plant Dis.*, 91: 639.
- Rafi, H., S. Dawar and M. Tariq. 2016. Combined effect of soil amendment with oil cakes and seed priming in the control of root rot fungi of leguminous and non-leguminous crops. *Pak. J. Bot.*, 48(3): 1305-1311.
- Reuveni, R., A. Nachmias and J. karikun. 1983. The role of seed borne inoculum on the development of *Macrophomina phaseolina* on melon. *Plant Dis.*, 67: 280-281.
- Sarfaraz, H. 2016. Effect of medicinal plant in the control of root rot fungi on leguminous crops. M.Sc thesis, Department of Botany, University of Karachi, pp. 94.
- Sheikh, A.H. and A. Ghaffar. 1975. Population study of sclerotia of *Macrophomina phaseolina* in cotton field. *Pak. J. Bot.*, 7: 13-17.
- Sikander, A., S. Shahnaz, M. Tariq and M.J. Zaki. 2009. Management of root diseases by combination of different soils with fertilizers. *Pak. J. Bot.*, 41(6): 3219-3225.
- Sokal, R. and F.J. Rohlf. 1995. Biometry: *The Principals and Practices of Statistical in Biological Research*, Freeman, New York. p. 887.

- Sokovic, M., J. Glamoclija, P.D. Marin, D. Brkic and L.J.L.D. Van Griensven. 2010. Antibacterial effects of the essential oils of commonly consumed medicinal herbs using an in vitro model. *Molecules*, 15: 7532-7546.
- Srivastava, .A, T. Singh, T. Jana and D. Arora. 2001. Microbial colonization of *Macrophomina phaseolina* and suppression of charcoal rot of chickpea. In: *Microbes and Plants*. (Ed.):
 A. Sinha. Vedam eBooks, New Delhi, India, pp. 269-319.
- Srivastava, R., M. Aragno and A.K. Sharma. 2010. Cow dung extract: a medium for the growth of *Pseudomonas* enhancing their efficiency as biofertilizer and biocontrol agent in rice. *Ind. J. Microbiol.*, 50: 349-354.
- Stover, R.H. 1962. *Fusarial* wilt (Panama Disease) of Bananas and other *Musa* Species. CMI, Kew, Surrey, UK.
- Sulaiman, S., D. Ibrahim, J. Kassim and L. Sheh-Hong. 2011. Antimicrobial and antioxidant activities of condensed tannin from *Rhizophora apiculata* barks. J. Chem. Pharm. Res., 3: 436-444.
- Tariq, M., S. Dawar, F.S. Mehdi and M.J. Zaki. 2008. Fertilizers in combination with *Avicennia marina* in the control of root rot diseases of okra and mung bean. *Pak. J. Bot.*, 40(5): 2231-2236.
- Wilhelm, S. 1955. Longevity of the *Verticillum* with fungus in the laboratory and field. *Phytopathlogy*, 45: 180-181.
- Yeung, H.M. 1985. Handbook of Chinese herbs and formulas. Institute of Chinese Medicine. Los Angeles, C.A.

(Received for publication 17 December 2016)