# COMPARATIVE EFFICACY OF COMMERCIAL FUNGICIDES AND TRICHODERMA HARZIANUM AGAINST FUSARIUM WILT IN STRAWBERRY

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### Abstract

Strawberry (*Fragaria ananassa*) cultivation is increasing among Pakistani farmers because of extra profit and demand in local market. This crop is attacked by multiple pathogens responsible for low yield. The current study's objective was to identify a sustainable and a better approach to managing Fusarium wilt caused by *Fusarium oxysporum* f.sp. *fragariae* (Schlecht) in the field. Firstly effectiveness of eight commercial fungicides was evaluated by the poisoned agar technique against colony growth of *F. oxysporum* f.sp *fragariae In vitro*. Among all tested chemicals, four (Score, Avito, Carbendazim and Topsin-M) were found to suppress and inhibit pathogen growth significantly with reference to control. Later, the efficacy of four strains/isolates of *Trichoderma harzianum* was also evaluated through dual culture technique against *F. oxysporum* f.sp *fragariae*. However, only one strain of *T. harzianum* was found to suppress colony diameter of pathogen. Afterwards, four fungicides and one isolate of *T. harzianum* were assessed in the greenhouse. Finally, three chemicals (Score, Carbendazim and Topsin-M) and a biocontrol isolate were applied in the field after artificial inoculation of pathogen in a randomized complete block design (RCBD). All three chemicals reduced disease severity of Fusarium wilt with varying degrees of success. The mean results of field trials indicated that Carbendazim and Score gave 72.0%, *T. harzianum* 80.0%, while Topsin-M showed 88.0% protection value against Fusarium wilt disease in the filed after 42 days. Topsin-M was most effective, followed by *T. harzianum* and can be applied to manage Fusarium wilt of strawberry confidently.

Key words: Biological management, Chemicals, Dual culture, Inoculation, Poisoned agar.

## Introduction

Cultivation of strawberry (Fragaria ananassa Duch.) has entered a critical phase of increase in Pakistan, but multiple factors like climate of the region, size of the fruit, and diseases restrict its quantity and quality. Among diseases, soil-borne fungal pathogens are the most common in a strawberry crop, viz. Phytophthora nicotinae (Mingzhu et al., 2013), P. fragariae (Wilcox et al., 1993), Verticillium dahliae (Thomas, 1932), Fusarium oxysporum (Koike et al., 2009), Rhizoctonia fragariae (Hussain & Mckeen, 1963), R. solani (Fang et al., 2013) and Pythium spp (Martin & Loper, 1999). Although many diseases have been reported but Fusarium wilt caused by Fusarium oxysporum f. sp. fragariae is one of the emerging diseases in strawberry growing areas of Pakistan (Veesar et al., 2015). The introduction of better cultural practices as acclimatized varieties and disease-free suckers can reduce disease problems in the long run, while appropriate pesticide uses are quick and effective ways to overcome disease before crop damage occurs. Several researchers have used preplanting chemicals like methyl bromide (Koike et al., 2009) and managed disease through fumigation. On the other hand, biological control organisms also showed a promising effectiveness against F. oxysporum f. sp. fragariae under controlled experimental conditions and including Bacillus Velezensis, B. subtilis and Trichoderma harzianum (Moon et al., 1995; Okayama et al., 1991; Tezuka & Makino, 1991; Nam et al., 2009; Zhang et al., 2012). The main symptoms of strawberry wilt are drying and withering of older leaves, stunting of plants, and reduction in fruit setting, wilting of foliar part and eventually plants collapsed and perished (Koike et al., 2009). Therefore, keeping in view, the frequent reports by framers of strawberry wilting in local areas, the current research was designed to manage this disease with all possible practical options presently available. The disease can be managed by applying commonly available fungicides and biological control, i.e., *Trichoderma harzianum*. The differences in cultivar susceptibility have not been recorded in Pakistan. A significant strawberry variety (Chandler) was selected and subjected to investigate. The study was initiated with the principal objective to develop a long-lasting and economical method to manage fusarium wilt in the strawberry crop. By overcoming these disease-related problems and improving the crop's quality, Pakistan can export strawberries to foreign countries, primarily Middle Eastern Countries.

## Methodology

**Collection of diseased specimen:** The symptomatic samples were gathered and examined from different locations of Lahore (Longitude :74.34 and Latitude :31.55) and Okara (Longitude: 73.459396 and Latitude: 30.808500) after complaints of strawberry growers. All samples were transported in sterile polythene bags in an icebox, and an accession number was assigned before storing at 4°C.

**Isolation, identification and preservation of pathogen:** The fungus was isolated by following the method of Joshi, 2013. Firstly, all infected leaves and lateral roots were trimmed off, leaving only the main stem and primary root. The stem was surface sterilized by dipping in 10% bleach (NaOCl) solution for 4 to 5 minutes and was dried out on paper towels. Thin wedges (2 to 4 mm) were cut from one side of the stem near the root/stem junction, including Pathogenicity test: Four-week-old seedlings of strawberry suckers (ver. Chandler) were transplanted to earthen pots containing sterilized soil. Plants were irrigated with sterile distilled water and covered with polythene bags for one day before inoculation. The conidial suspension  $(1 \times 10^8)$ spores/mL) prepared from 7-day old culture was atomized, 10 mL per plant and plants were again covered with polythene bags for two days, ensuring enough humidity for the establishment of the pathogen. Inoculated plants were then shifted to the greenhouse and periodically observed for symptom development. In case of disease, a re-isolation of the fungus from infected plants was done after two weeks, and the new isolate was compared with the original one, thus fulfilling Koch's postulates if identical.

Collection, identification and preservation of biocontrol agent: Four isolates/strains of Trichoderma harzianum were collected from different sources and vegetable growing fields in Faisalabad and Okara. Cultural characteristics including growth rate, colony colour and

appearance were recorded. These characteristics were taxonomically suitable features for Trichoderma spp. (Samuels et al., 2002). The isolates were identified on PDA following Rifai (1969) and preserved on a 2% PDA medium at 25°C. These isolates were evaluated for antagonistic potential against F. oxysporum f. sp. fragariae using a dual culture technique (Fig. 2). Mycelial growth of the fungal pathogen and Trichoderma harzianum isolates was recorded after 8, 12 and 16 days.

In vitro evaluation of chemicals: Commonly available commercial fungicides were evaluated by the poison agar method at 100, 200, 300 and 400 ppm for all formulations (Borum & Sinclair, 1968) in Department of Plant Pathology, University of Agriculture Faisalabad Pakistan. The description of fungicides, including their chemical name, active ingredient, trade name, and mode of action, is described in Table 1. The calibrated concentration of respective fungicides was mixed after autoclaving and before pouring the medium at 55°C. PDA plates without any fungicide served as control. After solidification of medium, a one-centimeter discs cut from a pure culture of the pathogen were placed in the centre of the poisoned agar plates and were incubated at 25°C for three weeks with four replications for each treatment. The mycelial growth of the pathogen was recorded in millimeters until the fungus's growth on control plates completely filled. Each fungicide's protective value was assessed by determining the percentage reduction in colony diameter over control by the following formula:

Percentage inhibition of mycelial growth =	Colony diameter in control – Colony diameter in treatment	
	Colony diameter in control	

Table 1. List of fungicides applied against Fusarium wilt in strawberry.							
S. No.	Trade name	Active ingredient	Formulation	Distributor	Dose per acre		
1.	Topsin-M	Thiophanate Methyl	70% WP	Arysta LifeScience	200 g		
2.	Aliette	Fosetyl Aluminium	80% WP	Bayer Crop Science	800 g		
3.	Sulfex Gold	Sulfur	80% WG	Jaffer Agro Services	600 g		
4.	Score	Difenoconazol	250 SC	Syngenta Pakistan	200 mL		
5.	Hexaconazole	Hexaconazole	5% EC	Arysta LifeScience	250 mL		
6.	Evito	Fluoxastrobin	480 SC	Arysta LifeScience	120 mL		
7.	Nativo	Tebuconazole + Trifloxystroben	75 WP	Bayer Crop Sciences	160 g		
8.	Carbendazim	Carbendazim	50% WP	Sungrow Agro	250 g		

Pot experiments: After the In vitro evaluation, fungicides and T. harzianum isolates with the best fungal growth inhibiting potentials were selected for greenhouse experiments. Artificial inoculation of pots was carried out with 10 mL of a freshly prepared conidial suspension (1.0 x 106 spores/mL) of F. oxysporum isolated from strawberry plants. The conidial was counted and adjusted using suspension а haemocytometer, whereas controls were kept un-inoculated. Moreover, 25 g of organic matter (semi-decomposed dhaincha leaves) was added to each pot. The experiments were conducted with a completely randomized design with 4 replications to evaluate the efficacy of selected fungicides and biocontrol isolate under controlled conditions.

under Management of disease sick field: Comprehensive field experiments were conducted in Okara during winter 2019, following randomized block

design with four treatments (Three chemicals and one bio-agent) and replicated thrice. Fresh suckers of the commonly grown variety Chandler were transplanted to 30x10 cm in an 8.0 m<sup>2</sup> size plot for each treatment/replication and NP fertilizer applied at 160:80 (kg/ha). Inoculum of F. oxysporum f. sp. fragariae was prepared on boiled chickpea grains in polythene bags (20x30 cm, autoclavable) and was applied at 500g/plot. The initial application of selected chemicals and biocontrol agents was applied together with first irrigation during transplanting, whereas further two applications were made two weeks apart before recording disease incidence.

The data were analyzed statistically. A lower than 5% level of significance for the null hypothesis of no correlation was used to interpret treatment differences using Duncan's Multiple Range (DMR) test.

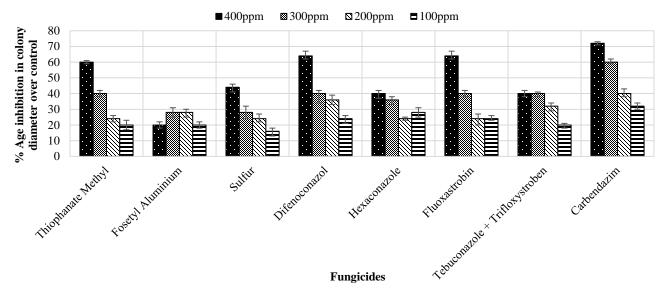


Fig. 1a. In vitro evaluation of chemicals using the poison agar technique after 8 days of incubation.

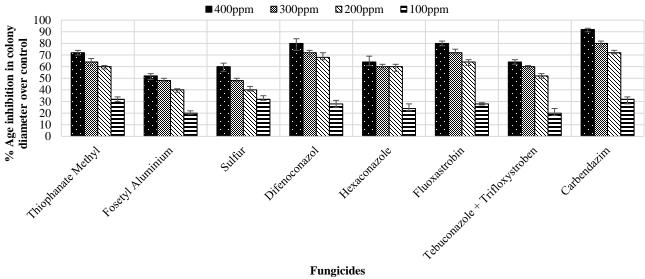




Fig.1b. In vitro evaluation of chemicals using the poison agar technique after 12 days of incubation.

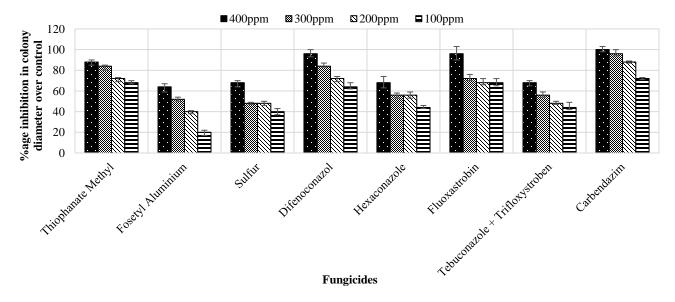


Fig. 1c. In vitro evaluation of chemicals using the poison agar technique after 16 days of incubation.

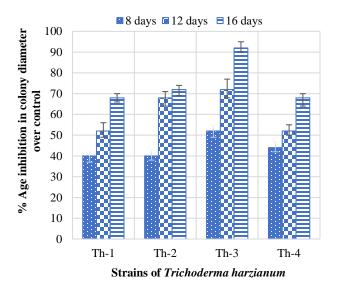
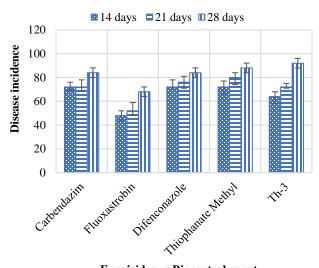
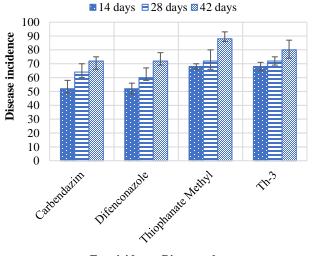


Fig. 2. In vitro evaluation of biological agent through dual culture.



**Fungicides vs Biocontrol agent** 

Fig. 3. Evaluation of fungicides and biocontrol agent under greenhouse conditions.



Fungicides vs Biocontrol agent

Fig. 4. Evaluation of chemicals and biocontrol agent under field conditions.

### **Results and Discussion**

Isolation, identification and preservation of pathogen and biocontrol agent: The isolated colonies of F. oxysporum f. sp. fragariae were pigmented with a reddishpurple colour and surrounded by a pinkish to white mycelium (Fig. 5A). The mycelia of F. oxysporum f. sp. fragariae were delicate and white with a pinkish or purple tinge (Fig. 5A). This pathogen produced three types of spores i.e., macroconidia (Fig. 5C), microconidia (Fig. 5D) and chlamydospores. The pathogen was identified by examining colony characters and shape of micro and macroconidia, according to Barnet and Hunter (1998) and Watanabe (2010). The microconidia were plentiful, oval to ellipsoid, straight and slightly curved, 5 to 12 x 2.2 to 3.5 µm, and non-septate, whereas macroconidia were sparse to abundant with 3 to 5 septations and pointed at both ends. The three-septate macroconidia measure 27 to 46 x 3 to 5  $\mu$ m while five-septate macroconidia measure 35 to 60 x 3 to 5 µm. The three-septate macroconidia were most abundant.

All isolates of *T. harzianum* were identified following Rafai (1969), where 1 to 2 concentric rings were formed with production of green conidia on PDA (Fig. 5B). The conidial production was denser in the plate's centre than at the margins. Micromorphological characteristics of two-week-old cultures of *T. harzianum* grown on PDA were examined. Conidia were globose to sub-globose, and size measured by micrometry  $(2.8x2.6 \ \mu m)$  (Fig. 5E).

In vitro Evaluation of chemicals and Trichoderma harzianum: A total of eight fungicides (Table 1) were tested with four doses for all fungicides, their effective concentration in reducing pathogen growth varied (Fig. 1a and 1b). Based on In vitro efficacy, test fungicides were divided into 2 groups. The promising group consists of Difenoconazole, Carbendazim, Fluoxastrobin and Thiophanate Methyl. These appeared more effective than the others, even at low concentrations. In the carbendazim treatments, the fungal pathogen completely failed to grow at 400 ppm, whereas 96 and 88% reduction in colony diameter of F. oxysporum was recorded at 300 and 200 respectively after 16 days of incubation (Fig. 1c). Similarly, Fluoxastrobin and Difenoconazole also suppressed the pathogen to 96% at 400 ppm, while Thiophanate Methyl inhibited the growth up to 88% at 400 ppm after 16 days of incubation. The fungicides Fosetyl Aluminium, Sulfur, Hexaconazole and Tebuconazole + Trifloxystroben, comprised the 2<sup>nd</sup> group, found less effective against F. oxysporum. Aliette was the least effective and showed 20, 40, 52 and 64% reduction at 400 ppm after 16 days, whereas its efficacy was even lower than the other at low doses. Sulfur, Hexaconazole and Tebuconazole + Trifloxystroben also showed little effect on the pathogen's growth. Despite the health hazards of chemical control, it is a fast and standard method to manage crop diseases, and eight chemicals were tested In vitro using the poison agar technique. A positive correlation was observed between concentration and inhibition of the pathogen's colony growth. Generally, high concentration of fungicides was more effective than low doses. There have been several

reports regarding the *In vitro* evaluation of fungicides against *F. oxysporum* f.sp *fragariae*. Our results confirm existing published results by Maitlo *et al.*, (2014) where carbendazim and thiophanate-methyl were most promising among 12 test chemicals against *F. oxysporum*. Moreover, similar results have also been reported by Akhtar *et al.*, (2017), Poddar *et al.*, (2004), Khan *et al.*, (2012) and Sultana & Ghafar (2010). Our investigations indicate that four fungicides efficiently inhibit the pathogen's growth (*In vitro*) and are probably effective against the same pathogen under agricultural field conditions; therefore, the four most effective fungicides (*In vitro*) were preceded further to greenhouse studies.

The dual-culture test of the biocontrol agent (Fig. 5F) T. harzianum (Th<sub>3</sub>) isolated from vegetable fields resulted in 92.0% inhibition over control after 16 days of incubation. Th<sub>3</sub>'s inhibition increased after each incubation interval, 52 and 72% after 8 and 12 days, respectively. In comparison, the isolates purified from the dung and rice field exhibited 68% of growth inhibition after 16 days. Fusarium wilt is a newly reported disease in Pakistan and can become a future threat for strawberry growers as it is a well-known soil and nursery soil-borne pathogen. The current experiments were conducted with the idea that bio-control agents may effectively be used against the Fusarium wilt and considered an excellent alternative to hazardous fungicides (Maltby et al., 2009). Although fungicides are widely used, most of these are considered hazardous to the farmer, the environment and might change soil microbial populations to the negative and should be replaced by more eco-friendly biological management (Frampton et al., 2006). Our findings were entirely in agreement with number of scientists who found that the isolates T. harzianum isolated from different rhizospheres were found to manage the plant pathogens as well (Burr et al., 1998, Postma et al., 2003 and Saikia *et al.*, 2003). Merkuz & Getachew (2012) found that Trichoderma isolates were very effective against *F. oxysporum*. Trichoderma's species were found superior to *Aspergillus niger* and *Bacillus subtilis* as biocontrol agents (Dubey *et al.*, 2007).

Four fungicides and one isolate of *T. harzianum* (Fig. 3) showing better inhibition were selected to be evaluated in greenhouse experiments where suckers were transplanted in artificially inoculated earthen pots, while controls were kept without biocontrol agents added.

The results indicated that all fungicidal treatments appeared to be effective compared to controls and minimum disease incidence was recorded. Finally, three chemicals and one isolate of *T. harzianum* were kept for field trials since Fluoxastrobin exhibited the least protection value (68%). The results revealed that Thiophanate Methyl showed 88%, *T. harzianum* 80%, while Carbendazim and Difenoconazole gave 72% protection value after 42 days in the field (Fig. 4).

Although there are many chemical control reports of Fusarium wilt in strawberry but pathogen management by T. harzianum has not been reported yet. The fungicides which efficiently reduced the pathogen growth In vitro are supposed to be effective against the same pathogen under field trials with varying degrees of success. Although carbendazim was leading to inhibit the colony growth In vitro but performance of Thiophanate Methyl was better in the field and greenhouse. Some researchers have reported that seed treatment with carbendazim reduce disease incidence significantly and increased yield (Kamdi et al., 2012). The current study concludes that fungicide applications (Thiophanate Methyl, Difenoconazole and Carbendazim) can be useful tools for Fusarium wilt control. The application of *T. harzianum* was also a worthy alternative and can be a more environmentally friendly tool for managing the soil borne plant pathogen in future.

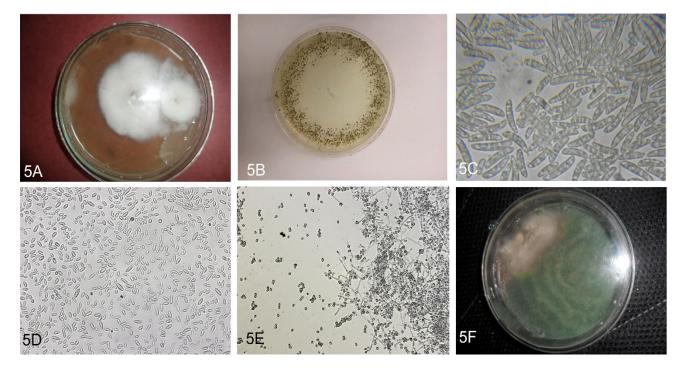


Fig. 5. (A-F) Colony of *Fusarium oxysporum* (5A) Colony of *Trichoderma. harzianum*; (5B) Macroconida of *Fusarium oxysporum* f. sp. *fragariae*; (5C) Microconidia of *Fusarium oxysporum* f. sp. *fragariae*; (5D) Conidia and mycelia of *T. harzianum* (5E) Dual Culture (5F).

#### Acknowledgement

This work was done with a start-up grant from the Higher education Commission (HEC) of Pakistan. We also thank the First Fungal Culture Bank of Pakistan University, Punjab Lahore, for providing biocontrol culture and pathogen identification.

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(Received for publication 12 May 2024)