

## **CLEYERA LIPINGENSIS VAR. XIUSHANENSIS (PENTAPHYLACACEAE): A NEW VARIETY FROM CHONGQING, CHINA**

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### **Abstract**

*Cleyera lipingensis* var. *xiushanensis* Zhe Zhang, Xin-Yun Lv & H.P. Deng var. nov. from Chongqing, China, is described and illustrated. This new variety is similar to *C. lipingensis* var. *lipingensis* and *C. lipingensis* var. *taipinensis* in leaves and fruits shape, as well as in the presence of serrate leaf margins. However, it can be distinguished by several key morphological characteristics: larger leaves (ca. 7 – 12 × 3 – 4.5 cm), more than 13 (–17) secondary veins on each side of the midrib, a usually 3 (–4) lobed style with pubescence, and a 3 (–4) loculed ovary with 8 – 10 ovules per locule. Phylogenetic analyses based on the nuclear ribosomal internal transcribed spacer (nrITS) region confirm that *C. lipingensis* var. *xiushanensis* is closely related to *C. lipingensis* var. *lipingensis*.

**Key words:** Morphology; Phylogeny; New variety; Taxonomy.

### **Introduction**

Pentaphylacaceae, traditionally recognized as a monogeneric family within Theales (Cronquist, 1981). It is now placed in Ericales, comprising 12 genera according to Weitzman *et al.*, (2004), APG III (2009), and APG IV (2016), or 14 genera according to Schönenberger *et al.*, (2005). The genus *Cleyera* Thunb. (Pentaphylacaceae), consisting of 24 species, are distributed in East, South, and Southeast Asia, as well as tropical America (Newman *et al.*, 2007; Ming & Bartholomew 2007; POWO, 2023). In China, nine species (7 endemic) have been recorded (Nakai *et al.*, 2007). *Cleyera* is sister to *Adinandra* Jack, but is distinguished by the lower number of ovules per locule and the seeds are 1-2 mm long and situated on a central axis (Keng, 1972; Ming & Bartholomew, 2007; Tsou *et al.*, 2016; Nguyen, 2017).

In 2023, during a biodiversity survey in Taiyangshan Municipal Nature Reserve, Xiushan, Chongqing, we encountered a distinctive *Cleyera* species at an altitude of 630 m. This species exhibited unique secondary vein counts, ovary chambers, and styles that distinguished it from all known *Cleyera* species. After an extensive literature review and examination of specimens in the Chinese Virtual Herbarium (CVH), we concluded that the *Cleyera* population in Taiyangshan Municipal Nature Reserve represents an undescribed variety of *C. lipingensis* var. *lipingensis* (Hand.-Mazz.) T.L. Ming (2007:444) [synonym: *C. japonica* var. *lipingensis* (Hand.-Mazz.) Kobuski (1937:127)], which is generally similar to *C. lipingensis* var. *taipinensis* (H. Keng) T. L. Ming (2007:444) (Nakai *et al.*, 2007). Consequently, we compared the new variety with these two varieties based on treatments in the literature (Keng, 1972; Nagamasu, 2006; Nguyen *et al.*, 2017; Ming & Bartholomew, 2007). In this study, we describe this new variety based on morphological characteristics and investigate its phylogenetic position using DNA sequences.

### **Materials and Methods of Phylogenetic Study**

To clarify the phylogenetic position of the new variety, we included 15 representative species of *Adinandra* and *Cleyera*, following Tsou *et al.*, (2016). *Norantea guianensis* Aubl. (1969:563) was selected as the outgroup. One nuclear region (nrITS) was used in the phylogenetic analyses. We generated sequences for 20 accessions representing 14 species of *Adinandra* and *Cleyera*. Table 1 provides voucher information and GenBank accession numbers for each taxon.

**Table 1. GenBank accession numbers of the sequence data generated in this study.**

| Species  | GenBank accession no. |
|--|-----------------------|
| <i>Adinandra hainanensis</i>                               | AF456255              |
| <i>A. latifolia</i>  | AY096023              |
| <i>A. elegans</i>  | AY626845              |
| <i>A. formosana</i>  | AY626846              |
| <i>A. lasiostyla</i>                                       | AY626847              |
| <i>A. millettii</i>  | AY626848              |
| <i>A. glischroloma</i>                                     | HM061547              |
| <i>A. bockiana</i>   | HM061548              |
| <i>A. lasiostyla</i>                                       | HM061550              |
| <i>A. millettii</i>  | KP092617              |
| <i>A. millettii</i>  | KP092618              |
| <i>A. nitida</i>   | KP092619              |
| <i>A. nitida</i>   | KP092620              |
| <i>A. bockiana</i>   | MN579515              |
| <i>A. glischroloma</i>                                     | OQ832760              |
| <b><i>Cleyera lipingensis</i> var. <i>xiushanensis</i></b> | <b>PV132331</b>       |
| <i>C. lipingensis</i>                                      | AF456257              |
| <i>C. pachyphylla</i>                                      | AY096025              |
| <i>C. japonica</i> var. <i>morii</i>                       | HM061545              |
| <i>C. japonica</i> var. <i>japonica</i>                    | HM061546              |
| <i>Norantea guianensis</i>                                 | FJ037835              |

DNA extraction, PCR amplification, sequencing, sequence editing, and sequence assembly were conducted following the protocols of Zhang *et al.*, (2013). The primers utilized for PCR amplification were as specified by Li *et al.*, (2016) and Taberlet *et al.*, (1991). The nrITS datasets were analyzed using Bayesian inference (BI) in PhyloSuite (Zhang *et al.*, 2020). The optimal substitution model (GTR+G) was selected using the Model Finder option based on the Bayesian Information Criterion (BIC) (Kalyaanamoorthy *et al.*, 2017). The Markov chain Monte Carlo (MCMC) algorithm was run for  $2.0 \times 10^7$  generations with four incrementally heated chains, starting from random trees and sampling one out of every 1,000 generations.

## Results

*C. lipingensis* var. *xiushanensis* was similar to *C. lipingensis* var. *lipingensis* and *C. lipingensis* var. *taipinensi*, but differs in its having more than 13 (–17) secondary veins on each side of middle vein, usually 3 (–4)-lobed and pubescent style, and 3 (–4)-loculed ovary

with 8–10 ovules per locule. Detailed comparison of morphological characters among this taxon and the morphologically close relatives was provided (Table 2).

The results indicated that the nrITS region of *C. lipingensis* var. *xiushanensis* is 704 bp in length. A Bayesian inference (BI) phylogeny, including BI posterior probability (BP) values, is presented in Fig. 1. As shown in Fig. 1, *C. lipingensis* var. *xiushanensis* was resolved as sister to *C. lipingensis* var. *lipingensis* (BP=0.954) and subsequently formed a sister relationship with *C. pachyphylla* (BP=1).

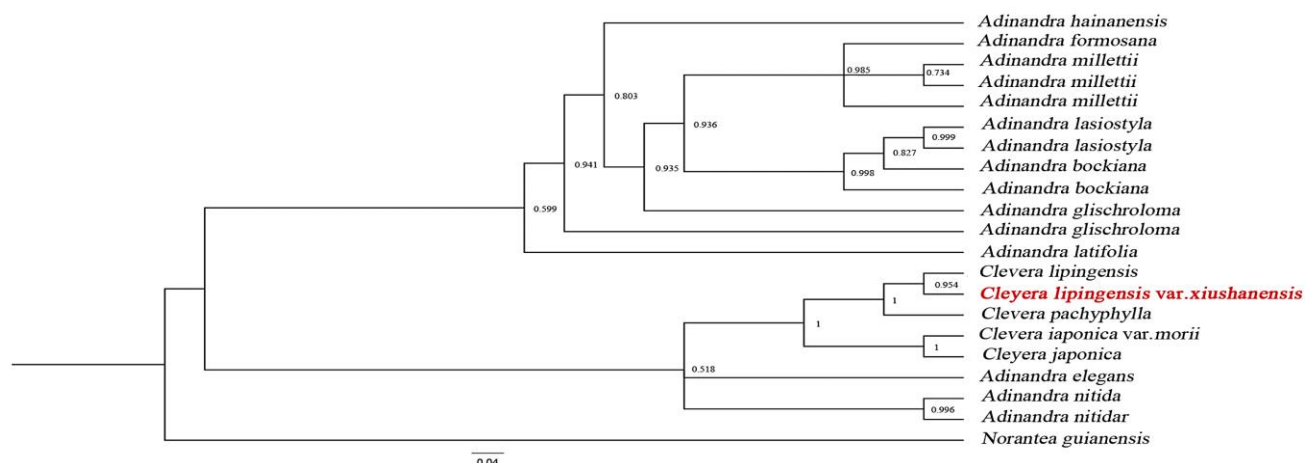
Thus, integrating both morphological and molecular evidence, we recognize *C. lipingensis* var. *xiushanensis* as a new variety of *C. lipingensis* var. *lipingensis*. The formal description is provided below:

## Taxonomic Treatment

***Cleyera lipingensis*** (Hand.-Mazz.) T.L. Ming var. ***xiushanensis*** Zhe Zhang, Xin-Yun Lv & H.P. Deng, var. nov. (秀山红淡比) (Figs. 2–4).

**Table 2. Morphological comparisons of *Cleyera lipingensis* var. *xiushanensis*, *C. lipingensis* var. *lipingensis* and *C. lipingensis* var. *taipinensi*.**

| Character              | <i>C. lipingensis</i> var. <i>xiushanensis</i>                        | <i>C. lipingensis</i> var. <i>lipingensis</i>                                 | <i>C. lipingensis</i> var. <i>taipinensi</i>                                       |
|------------------------|---|---|--|
| Shape and size of leaf | oblong to oblong-lanceolate, ca. 7–12 × 3–4.5 cm                      | oblong, oblong-elliptic, obovate, or obovate-elliptic, ca. 6–9 × 2.5–3.5 cm   | oblong-lanceolate, ovate, or obovate-spatulate, ca. 3–5.5 × 1–2 cm                 |
| Secondary veins        | over 13 (–17) on each side of midvein and visible on adaxial surfaces | 6–8 on each side of midvein and visible on both surfaces or abaxially obscure | 6–8 on each side of midvein and visible on both surfaces                           |
| Inflorescence          | Flowers axillary, (–2) 4–5 in a cluster                               | Flowers axillary, 3 (–5) in a cluster   | Flowers axillary, 2–4 in a cluster   |
| Pedicel                | ca. 1–1.5 cm  | ca. 1–2 cm  | ca. 1–2 cm   |
| Sepals                 | ovate to orbicular, 2–2.5 × ca. 3 mm, margin ciliolate, apex retuse   | ovate to orbicular, 2.5 × ca. 2.5 mm, margin ciliolate, apex rounded          | ovate to orbicular, 2–3 × ca. 2–2.5 mm, margin ciliolate, apex rounded, tip retuse |
| Petals                 | white, obovate to oblong, ca. 13 × 9 mm                               | white, obovate-oblong, ca. 8 mm   | white, obovate-oblong, ca. 8 mm  |
| Stamens                | 26–28, ca. 2–4 mm   | 25–30, ca. 4–6 mm   | 25–30, ca. 5 mm  |
| anthers                | fusiformis, ca. 1.5–2 mm, with filiform trichomes on the periphery.   | ovate to long ovate, ca. 1.5 mm, with filiform trichomes, connective exerted  | ovate, ca. 1.5 mm, with filiform trichomes, connective exerted                     |
| Style                  | ca. 5 mm, stigma usually 3 (–4) lobed with pubescent                  | ca. 6 mm, apically 2 lobed  | ca. 5 mm, glabrous, apically 2-lobed   |
| Ovary                  | ovoid, glabrous, 3 (–4)-loculed, 8–10 ovules per locule               | globose, glabrous, 2-loculed, more than 10 ovules per locule                  | ovoid, glabrous, 2-loculed, more than 10 ovules per locule                         |



**Fig. 1. Phylogenetic relationships between the genera of *Cleyera* and *Adinandra* inferred from ITS sequences. The species of *Norantea* was used as outgroup. Numbers above the branches represent Bayesian posterior probability (BP), respectively. *C. lipingensis* var. *xiushanensis* is noted in bold.**

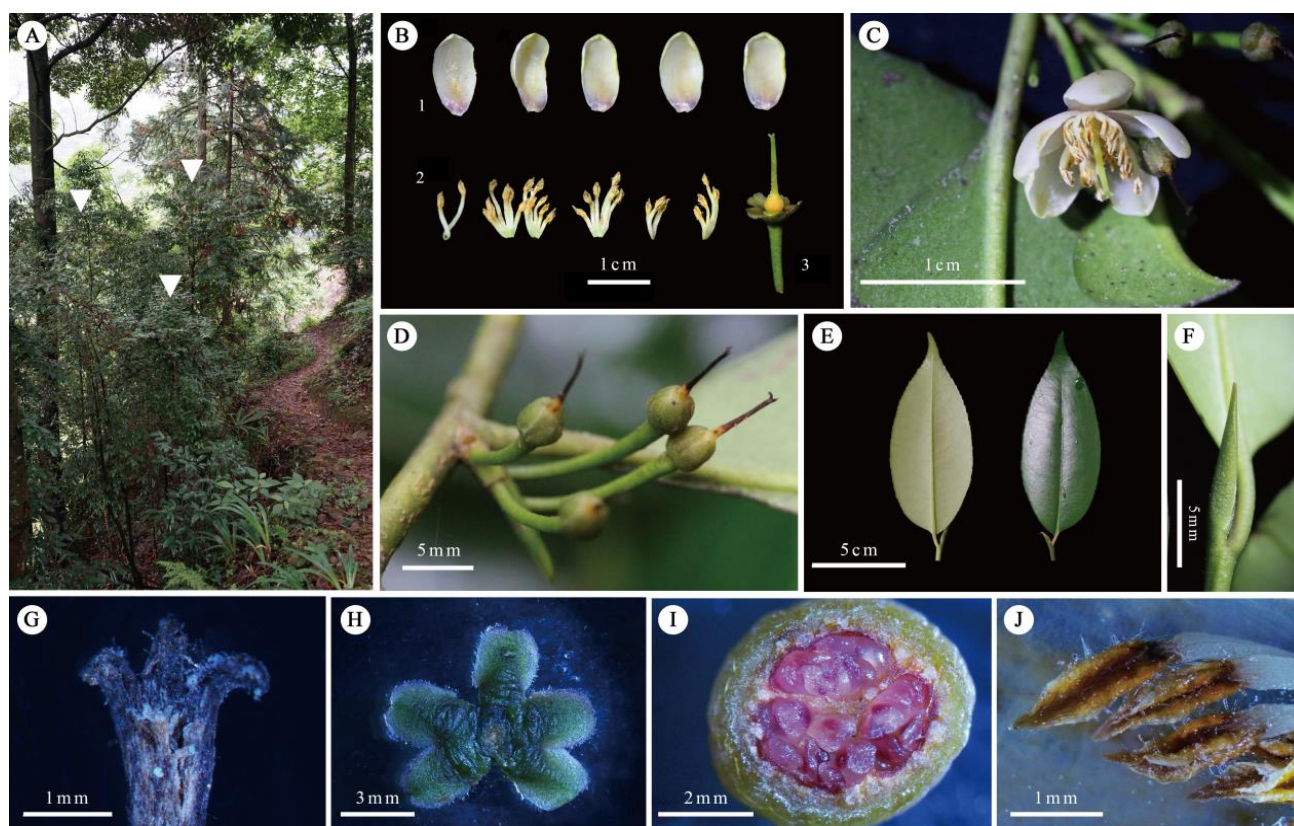


Fig. 2. *C. lipingensis* (Hand.-Mazz.) T. L. Ming var. *xiushanensis* Zhe Zhang, Xin-Yun Lv & H. P. Deng. A. habitat; B. one dissected flower, showing petals (1), stamens (2) and sepals with pistil (3); C. one opening flower; D. young fruits; E. leaves; F. bud; G. stigma; H. sepals; I. transverse section of ovary; J. anthers.

### Key to the species in Chongqing

- 1a. Young branches and terminal buds glabrous, blades margin entire, midrib slightly impressed adaxially ..... *C. japonica*
- 1b. Young branches and terminal buds pubescent, blades margin serrate, midvein adaxially raised ..... 2
- 2a. Secondary veins 6–8 on each side of middle vein and visible on both surfaces or abaxially obscure; 2 loculed ..... *C. lipingensis* var. *lipingensis*
- 2b. Secondary veins over 13 (–17) on each side of middle vein and visible on adaxial surfaces; 3 (–4) loculed ..... *C. lipingensis* var. *xiushanensis*

**Diagnosis:** The new variety resembles *C. lipingensis* var. *lipingensis* and *C. lipingensis* var. *taipinensi* but differs in its having bigger leaves ca. 7–12 × 3–4.5 cm (vs. ca. 6–9 × 2.5–3.5 cm in *C. lipingensis* var. *lipingensis* and ca. 3–5.5 × 1–2 cm in *C. lipingensis* var. *taipinensi*), young branchlets, buds, petioles and pedicels pubescent (vs. glabrous in *C. lipingensis* var. *taipinensi*), secondary veins over 13 (–17) on each side of midvein (vs. 6–8 on each side of midvein in *C. lipingensis* var. *lipingensis* and *C. lipingensis* var. *taipinensi*), usually 3 (–4) locule in ovary (vs. 2 locule in ovary), and stigma 3 (–4) (vs. 2 stigma in ovary).

**Description:** Shrubs or trees, 4–9 m tall. Young branches terete, green, sparsely pubescent, old twigs gray, glabrous; Terminal buds long-conic, 0.8–2 cm long, pubescent. Leaves alternate; blades leathery, oblong, elliptic or oblong-lanceolate, 7–12 × 3–4.5 cm, margin serrate, base cuneate, acute or acuminate at apex, abaxially pale green, both surfaces glabrous, adaxially dark green, glossy, abaxially midvein raised, over 13 secondary veins on either side of the middle vein and visible on adaxial surfaces; petioles 0.5–1 cm long, sparsely pubescent

when young, glabrescent. Flowers axillary, (–2) 4–5 in a cluster; pedicels 1–1.5 cm long, glabrous. Sepals ovate to orbicular, 2–2.5 × c. 3 mm, margin ciliolate, apex retuse. Petals white, obovate to oblong, c. 13 × 9 mm, apex rounded. Stamens 26–28, 2–4 mm long; anthers fusiformis, 1.5–2 mm long, with filiform trichomes on the periphery; filament 2–7 mm long, glabrous. Style c. 5 mm long, 3 (–4)-lobed at apex, pubescent. Ovary ovoid, 3–4 mm long, glabrous, 3 (–4)-loculed. Fruits globose, 8–10 mm in diam, 8–10 seeds per locule.

**Etymology:** The name refers to the locality where this new variety was first found.

**Geographical distribution:** *C. lipingensis* var. *xiushanensis* is only found in Xiushan county, Chongqing, China.

**Habitat and ecology:** The new variety *C. lipingensis* var. *xiushanensis* grows in secondary broad-leaved deciduous sparse forests at an elevation of 630 m. Flowering in May–June, and fruiting in October–November.



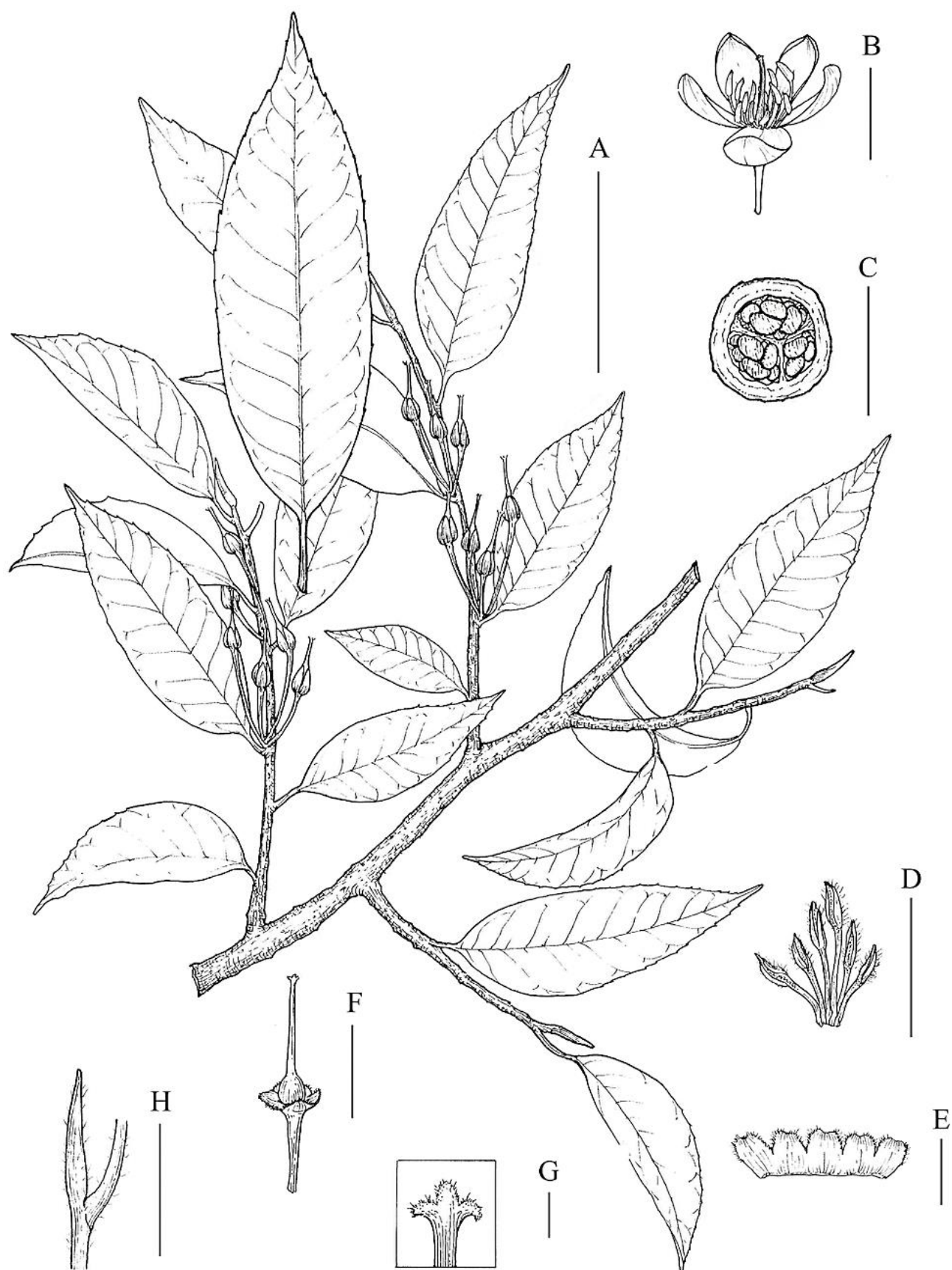


Fig. 3. *C. lipingensis* (Hand.-Mazz.) T. L. Ming var. *xiushanensis* Zhe Zhang, Xin-Yun Lv & H. P. Deng. A. Fruiting branch with young fruits (scale: 5cm); B. Flower (scale: 1cm); C. Transverse section of a young fruit with 3 loculi (scale: 5mm); D. Stamens (scale: 5mm); E. Sepals (scale: 5mm); F. Young fruit (scale: 5mm); G. Stigma (scale: 1mm); H. Bud (scale: 1cm). Drawn by Yi-fan Li.

**Type:**—CHINA. Chongqing (重庆市): Xiushan County (秀山县), Taiyang Village (太阳村), in secondary broad-leaved evergreen forest, 630 m, 4 June 2023, *Zhe Zhang, Xin-Yun Lv* ZZ0421 (holotype: HWA!; isotypes: HWA!, IBSC!, FJFC!).



Fig. 4. Holotype specimen of *C. lipingensis* (Hand.-Mazz.) T. L. Ming var. *xiushanensis* Zhe Zhang, Xin-Yun Lv & H. P. Deng.

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## References

- APG III. 2009. An update of the Angiosperm Phylogeny Group classification for the orders and families of flowering plants: APG III. *Bot. J. Linn. Soc.*, 161: 105–121.
- APG IV. 2016. An update of the Angiosperm Phylogeny Group classification for the orders and families of flowering plants: APG IV. *Bot. J. Linn. Soc.*, 181(1): 1–20.
- Cronquist, A. 1981. *An integrated system of classification of flowering plants*. Columbia University Press, New York, USA.
- Kalyaanamoorthy, S., B. Minh, T. Wong, A. Haeseler and L. Jermin. 2017. ModelFinder: fast model selection for accurate phylogenetic estimates. *Nat. Methods*, 14: 587–589.
- Keng, H. 1972. Theaceae In: (Eds.): Smitinand, T. and K. Larsen. *Flora of Thailand*, Vol. 2, The Forest Herbarium, Royal Forest Department, Bangkok. pp. 142–158.
- Li, M.H., Q. Gruss and Z.J. Liu. 2016. Nomenclature changes in *Phalaenopsis* subgen. *Hygrochilus* (Orchidaceae; Epidendroideae; Vandeae) based on DNA evidence. *Phytotaxa*, 275: 55–61.
- Ming, T. and B. Bartholomew. 2007. Theaceae. In: (Eds.): Wu, Z.Y., R.H. Raven and D.Y. Hong. *Flora of China*, vol. 12, Science Press, Beijing and Missouri Botanical Garden, St. Louis. pp. 366–478.
- Nagamasu, H. 2006. Theaceae. In: (Eds.): Iwatsuki K., D.E. Boufford and H. Ohba. *Flora of Japan*, Vol 2a, Kodansha, Tokyo. pp. 394–411.
- Nakai, S. and T. Turczaninow. 2007. *Cleyera*. In: (Eds.): Wu, Z.Y., R.H. Raven and D.Y. Hong. *Flora of China 12 (Pentaphylacaceae)*, Science Press, Beijing and Missouri Botanical Garden, St. Louis. pp. 443–446.
- Newman, M., S. Ketphanh, B. Svengsuksa, P. Thomas, K. Sengdala, V. Lamxay and K. Armstrong. 2007. *A Checklist of the Vascular Plants of Lao PDR*. Royal Botanic Garden Edinburgh, Edinburgh. p. 394.
- Nguyen, H.H. 2017. Theaceae G. Don. *Flora of Vietnam*, House for Science and Technology, Hanoi. (in Vietnamese).
- Plants of the World Online (POWO). 2023. Board of Trustees of the Royal Botanic Gardens, Kew. <https://powo.science.kew.org/>. (Accessed on 21 January 2024)
- Schönenberger, J., A.A. Anderberg and K. Sytsma. 2005. Molecular phylogenetic and pattern of floral evolution in the Ericales. *Int. J. Plant Sci.*, 166: 265–288.
- Taberlet, P., L. Gielly, G. Pautou and J. Bouvet. 1991. Universal primers for amplification of three non-coding regions of chloroplast DNA. *Plant Mol. Biol.*, 17(5): 1105–1109.
- Tsou, C.H., L. Li and K. Vijayan. 2016. The Intra-familial relationships of Pentaphylacaceae s.l. as revealed by DNA sequence analysis. *Biochem. Genet.*, 54: 270–282.
- Weitzman, A.L., S. Dressler and P.F. Stevens. 2004. Ternstroemiaceae. In: (Ed.): Kubitzki, K. *The families and genera of vascular plants*. Vol. VI. Springer, Berlin and New York. pp. 450–460.
- Zhang, D., F.L. Gao, I. Jakovlić, H. Zou, J. Zhang, W.X. Li and G.T. Wang. 2020. PhyloSuite: An integrated and scalable desktop platform for streamlined molecular sequence data management and evolutionary phylogenetics studies. *Mol. Ecol. Resour.*, 20: 348–355.
- Zhang, G.Q., K.W. Liu, L.J. Chen, X.J. Xiao, J.W. Zhai, L.Q. Li, J. Cai, Y.Y. Hsiao, W.H. Rao, J. Huang, X.Y. Ma, S.W. Chung, L.Q. Huang, W.C. Tsai and Z.J. Liu. 2013. A new molecular phylogeny and a new genus, *Pendulorchis*, of the *Aerides-Vanda* alliance (Orchidaceae: Epidendroideae). *PLoS One*, 8(4): e60097.

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