

## DISCRIMINATING SOME PROSPECTIVE CULTIVARS OF MAIZE (*ZEA MAYS L.*) FOR DROUGHT TOLERANCE USING GAS EXCHANGE CHARACTERISTICS AND PROLINE CONTENTS AS PHYSIOLOGICAL MARKERS

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### Abstract

A greenhouse experiment was conducted to discriminate maize (*Zea mays L.*) cultivars for drought tolerance using gas exchange characteristics and proline contents as physiological markers. Seven maize cultivars viz., Sahiwal-2002, Sadaf, EV-5098, Pak-Afgoyee, Agaiti-2002, Agaiti-85 and EV-1098 were grown under well watered or water deficit condition (60% of field capacity). Imposition of water deficit condition decreased the shoot and root fresh and dry weights, relative water content, photosynthetic and transpiration rates, stomatal conductance, while increased root/shoot ratio, chlorophyll *a*, *b* and chlorophyll *a/b* ratio, proline content and shoot N and K in all maize cultivars. Effect of drought stress was non-significant on shoot or root P and shoot N. Of all maize cultivars, cv. Agaiti-85 was found to be relatively better in growth, proline accumulation and gas exchange characteristics under drought stress as compared to the other cultivars.

### Introduction

Maize is one of the major world food crops. However, in areas where water availability is limited, maize grain losses may reach up to 24 million tons per year that is equivalent to 17% of well watered production in the world (Edmeades *et al.*, 1999). In Pakistan, water stress is also a major obstacle for maize production. Approximately, 65% of the maize in Pakistan has access to irrigation, while the remaining portion strictly depends on natural rain (Anon., 2007). Under mild drought stress, the reduction in maize crop is up to 10-13%, but under severe drought, the loss may increase many times. The problem becomes more alarming under arid conditions covering an area of 25-30% of major crops planted in Pakistan (Farooq *et al.*, 2007).

In view of all these problems, it is crucial to understand the mechanisms that plants use to adapt under water limited conditions (Ober & Sharp, 2003). It is now well established that, water deficit reduces plant growth by decreasing leaf area, net  $\text{CO}_2$  assimilation rate, transpiration rate, closure of stomata, chlorophyll fluorescence ( $F_v/F_m$ ), lower chloroplast activity and nutrient imbalance (Akram *et al.*, 2007; 2008; Ashraf *et al.*, 2007; Ali *et al.*, 2008). Plant growth directly depends on availability of water and water use efficiency (Edwards & Walker, 1983) and can be increased by increasing WUE which is affected by photosynthetic rate (Ehleringer & Monson, 1993). Net  $\text{CO}_2$  assimilation rate is positively correlated with net plant production and yield (Lawlor, 1995). Photosynthetic activity is directly related to dry mass production of plants and affects plant growth and yield (Natr & Lawlor, 2005). A positive correlation between photosynthetic rate and growth rate has been observed by many scientists in various crops like tomato and turnip (Makela *et al.*, 1998, 1999), *Brassica* species (Nazir *et al.*, 2001), and wheat (Ashraf & Bashir, 2003).

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Proline being the low molecular weight osmoprotectant, accumulates in larger amounts than other amino acids in water stressed plants (Pedrol *et al.*, 2000; Tamura *et al.*, 2003) and plants accumulate high concentration of proline under water deficit conditions (Parida *et al.*, 2007). Proline accumulation is thought to be a key adaptation under sever water deficit conditions (Akram *et al.*, 2007). The role of proline as a protective agent against reactive oxygen species is also very important (Hare *et al.*, 1999). High accumulation of proline is effective in chlorophyll stability (Ashraf *et al.*, 1995) and helps in stress tolerance of plants (Bartels & Nelson, 1994; Hanson *et al.*, 1994). Drought tolerance is positively correlated with high accumulation of proline in many crops like wheat and barley (Nayyar & Walia, 2003), rice (Hsu *et al.*, 2003) and *Brassica juncea* (Madan *et al.*, 1995). Transgenic tobacco plant also showed high drought resistance by having high proline content (Kishor *et al.*, 1995). Under drought stress, high accumulation of proline in the cytoplasm helps in osmotic adjustment of maize plants (Ketchum *et al.*, 1991; Voetberg & Sharp, 1991).

Maize having great economic importance, both for humans and animals as food and forage, the development of maize with high and stable yield under low moisture is an important priority for today's needs, as access to drought tolerant cultivars may be the only reasonable alternative to many small scale farmers (Tabassum, 2004). In view of the importance of maize, the principle objective of the present study was to appraise the differential resistance of 7 maize cultivars to drought stress using gas exchange characteristics and proline contents as physiological markers.

## Materials and Methods

To discriminate some prospective maize (*Zea mays* L.) cultivars for drought tolerance using gas exchange characteristics and proline contents as physiological markers, 7 maize cultivars (Sahiwal-2002, Sadaf, EV-5098, Pak-Afgoyee, Agaiti-2002, Agaiti-85 and EV-1098) were grown in a net house in the Old Botanical Garden, University of Agriculture Faisalabad, during September to November 2007. The average day and night temperatures were  $31 \pm 2^\circ\text{C}$  and  $19 \pm 3^\circ\text{C}$ , respectively. The relative humidity ranged from 32.6 to 51.4% and day length from 11-12 h.

The seed materials of maize cultivars, used in this study, were obtained from Maize and Millets Institute, Yousafwala (Sahiwal). Ten seeds were sown in each plastic pot (21 cm diameter and 25 cm depth) containing 10 kg air-dried soil. Saturation percentage of the soil used was 35.08, pH, 7.89 and ECe, 1.63 dS m<sup>-1</sup>. The plants were allowed to establish for 12 days after sowing before the start of drought stress. The plants were thinned to 6 plants per pot. Drought stress treatments were control (normal watering) or 60% of field capacity. Plants were harvested, 15 days after drought stress. The data for the following attributes were recorded.

**Plant biomass:** Two plants from each pot were uprooted carefully, washed with distilled water and fresh weights of both shoots and roots recorded. Then, plant samples were oven-dried at 65°C up to constant weight and dry weights recorded.

**Chlorophyll contents:** Chlorophyll 'a' and 'b' contents were determined according to Arnon (1949). Fresh leaves (0.5g) were ground in 80% acetone and centrifuged at 10,000 x g for 5 minutes. Absorbance of the supernatant was read at 645 and 663 nm using a spectrophotometer (Hitachi-U2001, Tokyo, Japan).

Chlorophylls *a* and *b* were calculated using the following formulae:

$$\text{Chl. } a \text{ (mg g}^{-1} \text{ f.wt)} = [12.7 \text{ (OD 663) } - 2.69 \text{ (OD 645)}] \times V/1000 \times W$$

$$\text{Chl. } b \text{ (mg g}^{-1} \text{ f.wt)} = [22.9 \text{ (OD 645) } - 4.68 \text{ (OD 663)}] \times V/1000 \times W$$

*V* = volume of the extract (ml), *W* = weight of the fresh leaf tissue (g)

**Relative water contents (RWC):** Relative water contents of fully expanded third leaf from top were determined following Turner (1986) using the following equation:

$$\text{RWC} = [\text{FW} - \text{DW}] \times 100 / [\text{TW} - \text{DW}]$$

FW = Fresh weight, DW = Dry weight, TW = Turgid weight

**Proline determination:** A fully expanded second leaf was used to determine proline according to the method of the Bates *et al.* (1973) after extraction with 3% 5-sulphosalicylic acid at room temperature. Proline concentration was calculated on fresh weight basis as follows:-

$$\mu\text{mole proline g}^{-1} \text{ fresh weight} = (\mu\text{g proline mL}^{-1} \times \text{mL of toluene}/115.5) / (\text{g of sample})$$

**Gas exchange characteristics:** An open system LCA-4 ADC portable infrared gas analyzer (Analytical Development Company, Hoddesdon, England) was used for the measurement of net  $\text{CO}_2$  assimilation rate (*A*), transpiration rate (*E*), stomatal conductance (*g<sub>s</sub>*) and sub-stomatal  $\text{CO}_2$  concentration (*C<sub>i</sub>*) of a fully expanded youngest leaf of each plant. These measurements were made from 10:15 to 11:45 a.m. with the following specifications/adjustments: leaf surface area 11.35 cm<sup>2</sup>, ambient  $\text{CO}_2$  concentration (C<sub>ref</sub>) 354.4  $\mu\text{mol mol}^{-1}$ , temperature of leaf chamber varied from 31.5 to 37.8°C, leaf chamber gas flow rate (*v*) 392.8 mL min<sup>-1</sup>, Molar flow of air per unit leaf area (U<sub>s</sub>) 404.84 mol m<sup>-2</sup> s<sup>-1</sup>, ambient pressure (P) 99.2 kPa, water vapor pressure (e<sub>ref</sub>) into chamber ranged from 20.5 to 23.1 mbar, PAR (Q<sub>leaf</sub>) at leaf surface was maximum up to 1048  $\mu\text{mol m}^{-2} \text{ s}^{-1}$ .

**Determination of mineral elements in plant tissues:** The dried ground leaf or root material (0.1 g) was digested with a digestion mixture according to the method of Allen *et al.*, (1986). Nitrogen was estimated by micro-Kjeldhal method (Bremner, 1965). Phosphorus was determined by spectrophotometer (Hitachi-220) using Barton reagent as described by Jackson (1962). Potassium was determined with a flame photometer (Jenway, PFP-7).

**Statistical analysis:** Two-way (cultivars and drought stress) analysis of variance technique was employed for the statistical analysis of the data collected (Steel & Torrie, 1980). The mean values were compared with the least significance difference test (LSD) following Snedecor & Cochran (1980).

## Results

Imposition of drought stress treatment (60% of field capacity) for a period of 15 days to 12 day-old plants of 7 maize cultivars viz., Sahiwal-2002, Sadaf, EV-5098, Pak-Afgoyee, Agaiti-2002, Agaiti-85 and EV-1098 had a significant reducing effect on shoot and root fresh and dry biomass. The difference among cultivars with respect to the above-mentioned attributes was non-significant. Cultivars differed significantly for root/shoot ratio. Cultivars Sahiwal-2002 and Agaiti-85 were higher while Sadaf and Pak-Afgoyee

lower in root/shoot ratio than the other cultivars. Root/shoot ratio increased with the imposition of water deficit conditions (60% of field capacity) in all cultivars except in Sadaf where the response was reverse (Table 1; Fig. 1).

There was a significant increase in chlorophyll *a* and *b* contents ( $p \leq 0.001$ ;  $p \leq 0.01$ , respectively) under water deficit conditions in all cultivars except Sahiwal-2002 in which the response was reverse. A maximum increase in chlorophyll *a* and *b* contents was observed in EV-5098 and Agaiti-2002 (Table 1; Fig. 1). Chlorophyll *a/b* ratio was also increased under water deficit conditions, except in cvs. Sahiwal-2002 and Pak-Afgoyee in which the ratio was decreased (Table 1; Fig. 1).

Relative water content (RWC) decreased significantly ( $p \leq 0.001$ ) in all maize cultivars due to drought stress. Cultivar Sahiwal-2002 was the highest of all the cultivars in RWC (Table 1; Fig. 2). Drought stress significantly ( $P \leq 0.001$ ) increased the proline contents of all maize cultivars. However, difference in all cultivars in proline contents was consistent in all 7 maize cultivars but no significant difference was observed among the cultivars under well watered or water limited conditions (Table 1; Fig. 2).

There was a significant decrease ( $p \leq 0.001$ ) in both photosynthetic and transpiration rates under water deficit conditions. Photosynthetic and transpiration rates were higher in cv. Pak-Afgoyee under both well watered and drought stressed conditions as compared to other cultivars, while the reverse was true for cv. Agaiti-2002 under well watered conditions (Table 1; Fig. 2).

Stomatal conductance ( $g_s$ ) and sub-stomatal  $\text{CO}_2$  concentration ( $C_i$ ) were significantly decreased in all 7 cultivars of maize under water deficit condition. Cultivars also differed significantly in the two gas exchange characteristics. However, stomatal conductance and sub-stomatal  $\text{CO}_2$  were lower in cvs. EV-5098 and EV-1098 than those in the other cultivars under water deficit conditions. In contrast, under well-watered conditions,  $g_s$  and  $C_i$  were higher in cvs. EV-5098 and EV-1098 than those in the other cultivars (Table 1; Fig. 2).

Water use efficiency ( $A/E$ ) varied significantly among the cultivars. Response of the cultivars was different under control or water stress conditions. Under non-stressed conditions, cv. EV-5098 was the highest in WUE, while the same was true for cv. Agaiti-85 under water deficit conditions (Table 1; Fig. 2).

$Ci/Ca$  ratio increased significantly in all cultivars under water limited conditions. A maximum increase in  $Ci/Ca$  ratio was observed in cv. Sahiwal-2002. The interaction between drought and cultivars for  $Ci/Ca$  ratio was also significant ( $p \leq 0.001$ ) (Table 1; Fig. 2).

Shoot N decreased significantly due to drought stress in all maize cultivars except Pak-Afgoyee, while there was no significant effect of drought stress on root N content. Cultivar EV-1098 suppressed all the cultivars in root N content under water deficit conditions (Table 1; Fig. 3).

Water deficit condition had no significant increasing or decreasing effect on shoot or root phosphorous concentration, while the cultivars showed variable behaviour in root P. Overall, cv. Sahiwal-2002 was the highest of all cultivars in root P (Table 1; Fig. 3).

There was a highly significant ( $p \leq 0.001$ ) effect of water deficit conditions on shoot and root  $K^+$  concentrations. Water deficit conditions caused a significant increase in shoot  $K^+$  in all cultivars. However, cultivar difference was not prominent for root  $K^+$  (Table 1; Fig. 3).

**Table 1.** Mean squares from analyses of variance of data for growth, chlorophyll pigments, gas exchange characteristics and mineral nutrients of seven maize cultivars when 12 day-old plants were subjected to water deficit conditions for 15 days. (n = 4)

Source of variation	Degrees of freedom	Shoot fresh weight	Shoot dry weight	Root fresh weight	Root dry weight
Cultivars (Cvs)	6	202.52ns	2.945ns	8.22ns	0.396**
Drought (D)	1	45873.6***	374.67***	820.65***	6.681***
Cvs x D	6	164.18ns	0.933ns	3.795	0.055ns
Error	42	7.31	0.50	3.21	0.113
Source of variation	Degrees of freedom	Root/ Shoot ratio	Chlorophyll <i>a</i>	Chlorophyll <i>b</i>	Chlorophyll <i>a/b</i> ratio
Cultivars (Cvs)	6	0.011*	0.221**	0.055***	0.505ns
Drought (D)	1	0.036**	1.056***	0.067**	2.322*
Cvs x D	6	0.005ns	0.274**	0.037**	0.609ns
Error	42	0.004	0.067	0.009	0.391
Source of variation	Degrees of freedom	Relative water content	Proline contents	<i>A</i>	<i>E</i>
Cultivars (Cvs)	6	1823.2***	2.542ns	180.26***	1.446**
Drought (D)	1	9880.3***	625.62***	2746.4***	39.254***
Cvs x D	6	643.21*	2.357ns	145.36***	0.332ns
Error	42	245.7	1.601	9.374	0.372
Source of variation	Degrees of freedom	<i>g<sub>s</sub></i>	<i>C<sub>i</sub></i>	<i>A/E</i>	<i>C<sub>i</sub>/C<sub>a</sub></i>
Cultivars (Cvs)	6	4996.2***	7059.5***	40.64***	0.053***
Drought (D)	1	108504.9***	105668.1***	1.865ns	0.877***
Cvs x D	6	3955.5**	8483.5***	86.92***	0.065***
Error	42	1064.7	044.7	3.906	0.009
Source of variation	Degrees of freedom	Shoot N	Shoot P	Shoot K	
Cultivars (Cvs)	6	28.888**	28.585ns	914.40***	
Drought (D)	1	823.09***	129.71ns	13801.2***	
Cvs x D	6	29.834**	69.73ns	1140.6***	
Error	42	8.683	32.59	59.36	
Source of variation	Degrees of freedom	Root N	Root P	Root K	
Cultivars (Cvs)	6	10.138***	55.761*	7.955ns	
Drought (D)	1	2.755ns	74.062ns	199.75***	
Cvs x D	6	3.779ns	66.799*	4.845ns	
Error	42	1.994	20.805	9.615	

\*, \*\*, \*\*\* = Significant at 0.05, 0.01, and 0.001 level, respectively.

ns = Non-significant

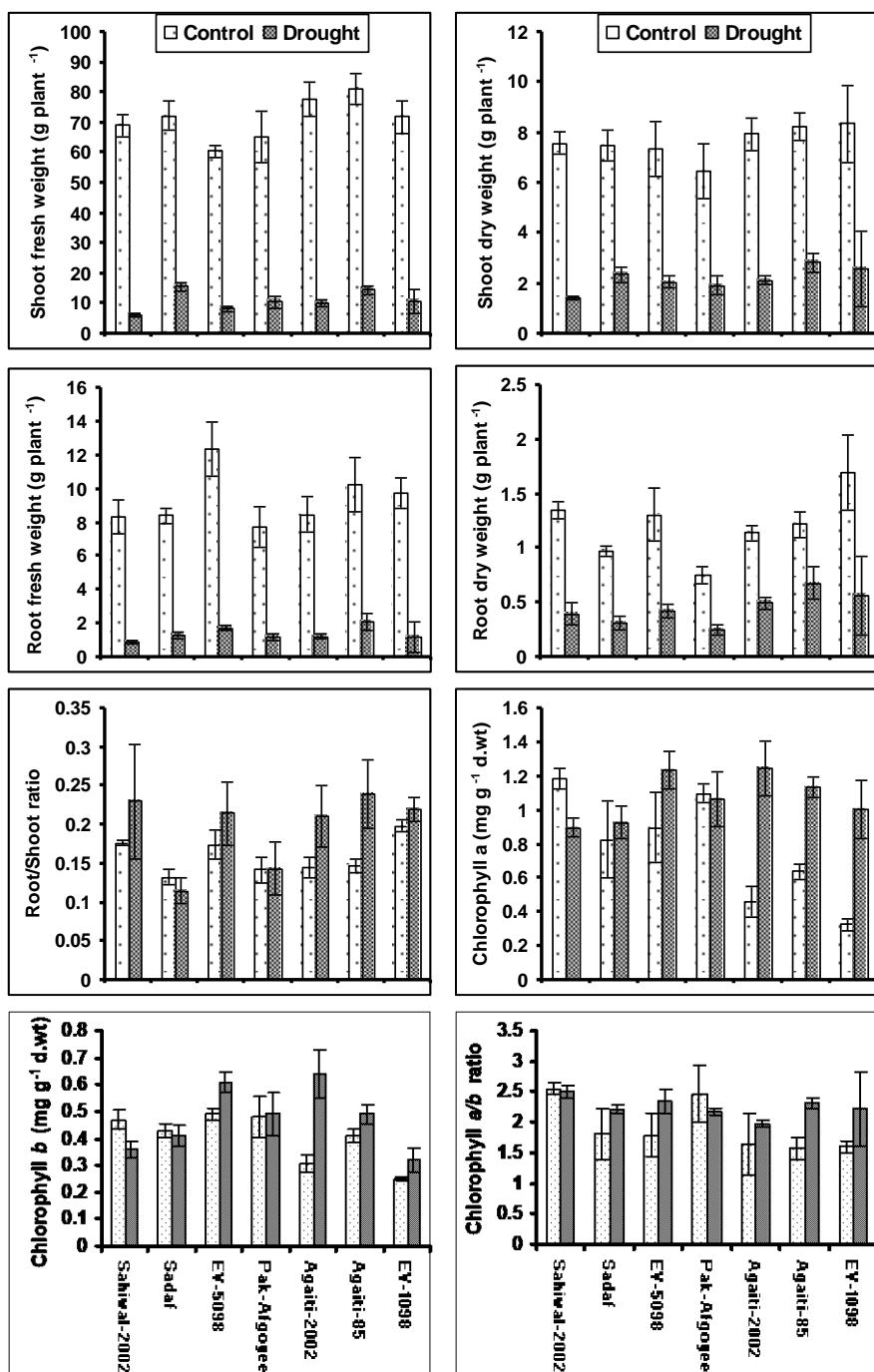


Fig. 1. Growth attributes and chlorophyll contents of 7 maize (*Zea mays* L.) cultivars when 12 day-old plants were subjected to water deficit conditions for 15 days.

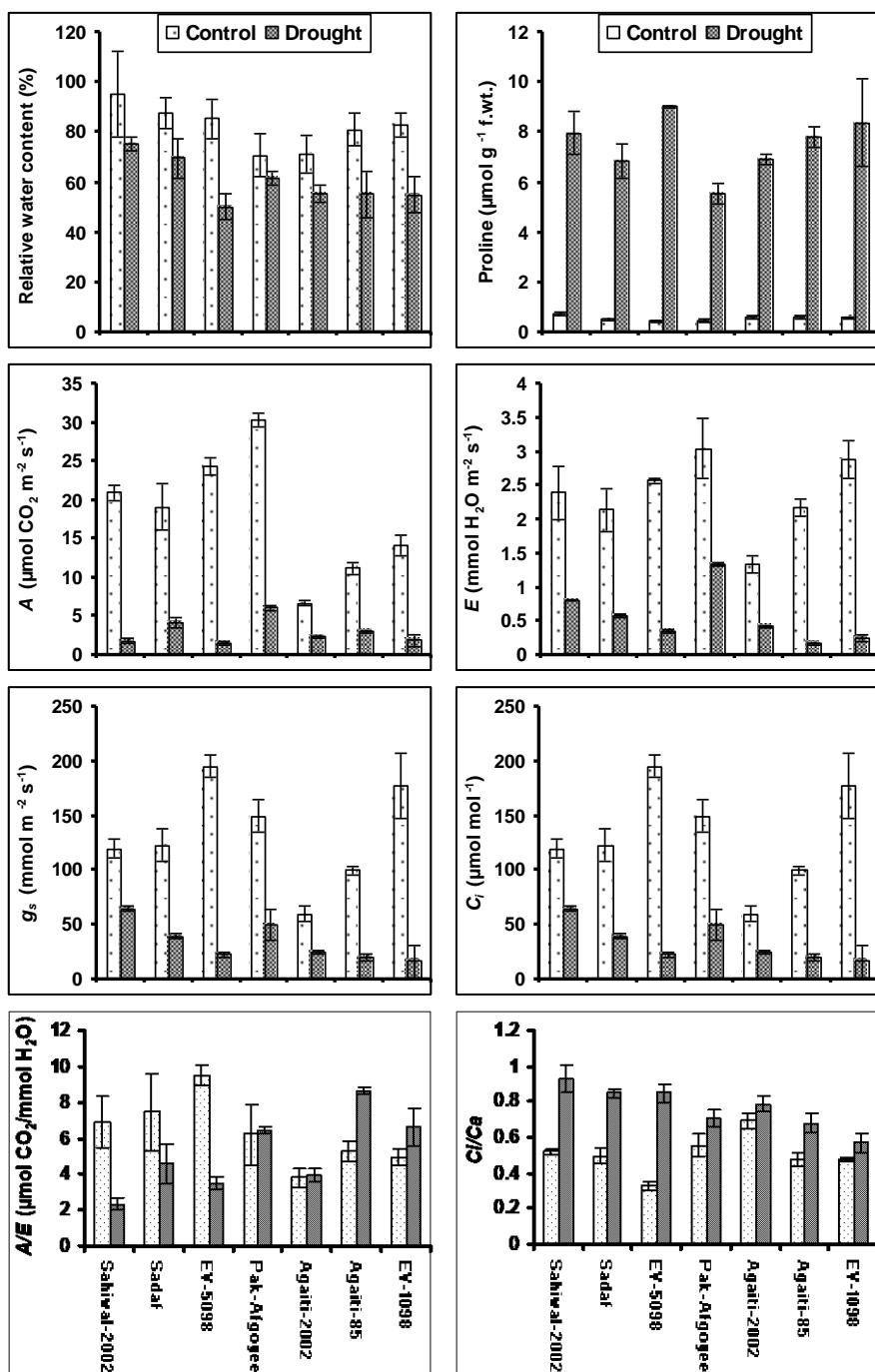


Fig. 2. Relative water content, proline content and gas exchange characteristics of 7 maize (*Zea mays* L.) cultivars when 12 day-old plants were subjected to water deficit conditions for 15 days.

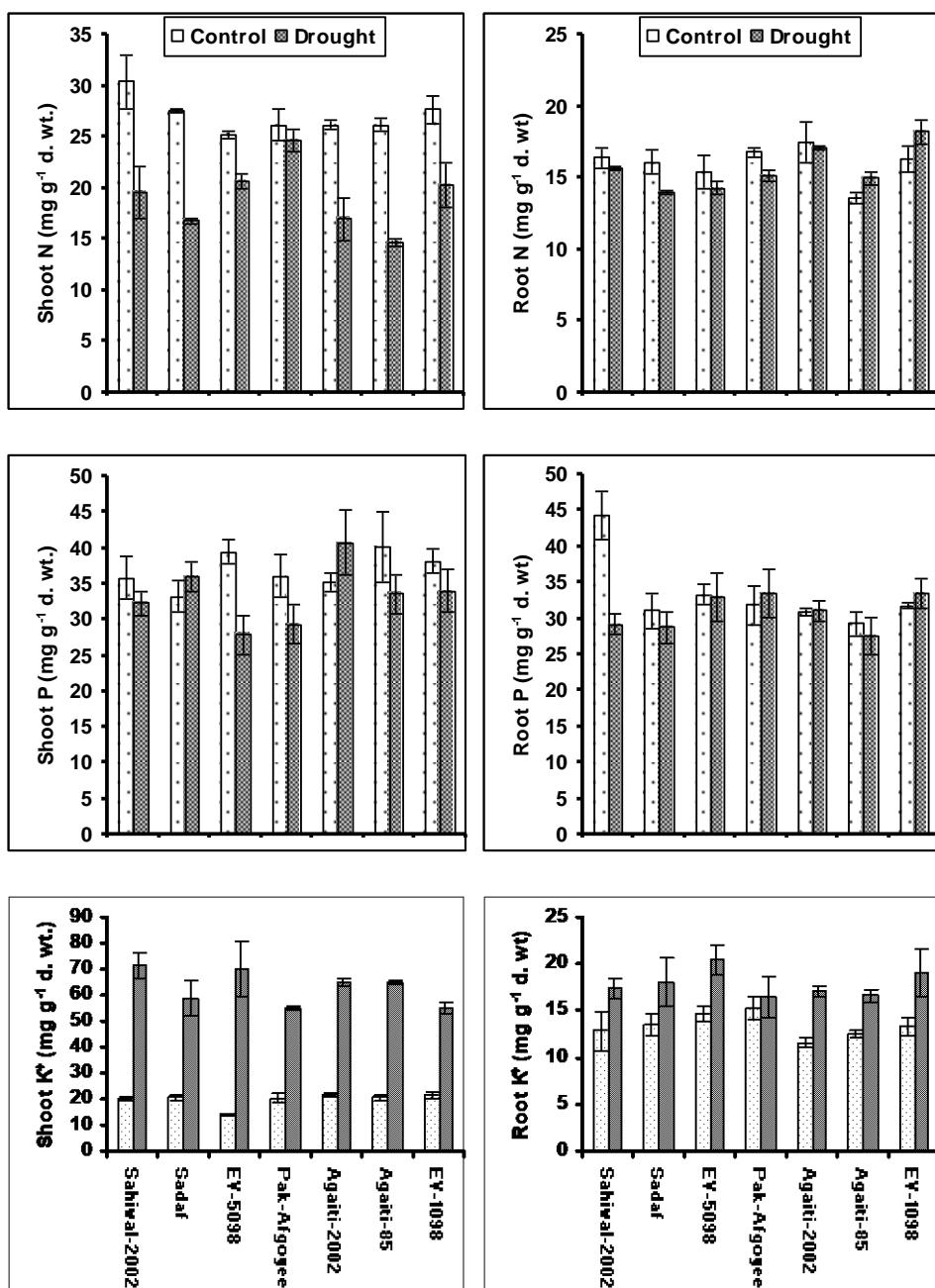


Fig. 3. Shoot and root mineral nutrients of 7 maize (*Zea mays* L.) cultivars when 12 day-old plants were subjected to water deficit conditions for 15 days.

## Discussion

Drought effects on plants are complex, variable and are expressed by a number of primary and secondary factors (Boyer, 1996). A common adverse effect of low water potential or water deficit stress is the reduction in fresh and dry biomass production in different crops such as grasses (Ashraf & Yasmin, 1995), wheat (Peschke *et al.*, 1997; Ashraf *et al.*, 1998), maize (Abrechit & Carberry, 1993) and rice (Manabendra & Baruah, 1998). Similarly, in our study water deficit conditions (60% of field capacity) imposed for a period of 15 days to 12-day-old plants had a significant detrimental effect on the growth of 7 maize cultivars viz., Sahiwal-2002, Sadaf, EV-5098, Pak-Afgoyee, Agaiti-2002, Agaiti-85 and EV-1098). These results are in agreement with some earlier studies in which it was reported that progressive drought during early growth stage suppressed the shoot dry matter production, root development and water uptake in rice (Banoc *et al.*, 2000) and wheat (Lopez *et al.*, 2003).

Huang *et al.*, (1997) were of the view that maintenance of root growth for nutrient and water uptake during drought is vital for plant tolerance to drought as water deficit stress was suggested to be the primary cause of root death under field conditions (Smucker *et al.*, 1991; Huang & Nobel, 1992) especially in the surface soil. In the present study, reduction in fresh and dry biomass production (shoot and root) was observed in all maize cultivars with the imposition of water stress treatment. However, there was an increase in root/shoot ratio under water deficit conditions (Hamblin *et al.*, 1990) that may have been due to increased accumulation of assimilates diverted to root growth (O, Toole & Bland, 1987), differential sensitivities of the roots and shoots to endogenous ABA or to a greater osmotic adjustment in roots compared with shoots (Sharp & Davies, 1989).

In our study, water stress significantly increased chlorophyll *a* and *b* contents in most of the maize cultivars under investigation. Plants under water shortage have evolved mechanisms to protect against photodamage. One such mechanism for protection entails changes in chlorophyll content in order to reduce the extent of absorbed light (Giardi *et al.*, 1996; Murchie & Horton, 1997). So the increase in chlorophyll contents under water deficit conditions is a common observation (Estill *et al.*, 1991; Hamada, 1996). In the present study, chlorophyll "a" was less affected than chlorophyll "b" under water deficit conditions. These findings are parallel to what was earlier observed in maize (Garcia *et al.*, 1987).

One of the most important responses of plants to drought and other abiotic stresses is an over-production of different types of compatible solutes (Serraj & Sinclair, 2002; Ashraf & Harris, 2004). Of these organic solutes, proline being the low molecular weight accumulates greatly in water stressed plants (Tamura *et al.*, 2003; Parida *et al.*, 2008). Moussa & Abdel-Aziz (2008) also observed that proline contents increased in maize seedlings with the application of drought stress. Proline accumulates in all parts of the maize seedlings at low water potentials (Raymond & Smirnoff, 2002). Our results for high accumulation of proline in all maize cultivars are in agreement with a number of past studies including some cited earlier. Thus, proline accumulation is not just a sign of cellular injury resulting in response to water shortage but is a marker of stress tolerance having a definite osmoregulatory role in plants subjected to stressful conditions.

Decline in the growth of many plant species subjected to a water limited environment is often associated with a reduction in photosynthetic capacity (Lawlor, 2002; Dubey, 2005). Reduction in photosynthetic rate has earlier been reported in different crops e.g., in tomato (Srinivasa-Rao *et al.*, 2000), wheat (Molnar *et al.*, 2002) and sunflower (Tezara *et al.*, 2002). With the increase in the intensity of water stress, a

biochemical decline of photosynthetic process is observed. Decline in photosynthetic rate might be due to stomatal closure, which reduces  $\text{CO}_2/\text{O}_2$  ratio in leaves and inhibits photosynthesis (Janson *et al.*, 2004; Moussa, 2006) which is in agreement with our findings that  $g_s$  decreased under water deficit conditions in all maize cultivars. Of various responses, stomatal closure is one of the earliest responses to drought, protecting the plant from extensive water loss (Chaves *et al.*, 2003). Drought stress leads to a noticeable decrease in stomatal conductance ( $g_s$ ) and mesophyll conductance, and increase in intercellular  $\text{CO}_2$  concentration along with the decrease in photosynthetic rate ( $A$ ) (Siddique *et al.*, 1999). Leaf stomatal conductance ( $g_s$ ) is also of great importance under the conditions when water stress increases (Flexas & Medrano, 2002) thereby restricting  $\text{CO}_2$  entrance into the leaf (Cornic, 1994). The extent of stomatal conductance can be used as an indicator to assess stomatal or non-stomatal limitations to photosynthesis under water deficit conditions (Flexas *et al.*, 2002a; 2002b; Medrano *et al.*, 2002).

In the present study, transpiration rate ( $E$ ) declined severely in all maize cultivars under water-limited condition. Plants can compensate water deficit condition with the closure of stomata to avoid further loss of water through transpiration (Lawlor, 1995).

Values for water use efficiency (WUE calculated as  $A/E$ ) showed significant differences among all the cultivars. It showed an increasing trend in some cultivars and decreasing in some others under water stress. Increase in WUE under reduced water availability is critical for plant survival (Chaves *et al.*, 2004). The increasing trend in WUE is in agreement to what has earlier been observed by Ashraf *et al.*, (2002) in okra (*Hibiscus esculentus*).

Leaf relative water content has been emphasized as a better indicator of water status of a plant than water potential (Sinclair & Ludlow, 1985). Our results for RWC are similar to what Moussa & Abdel-Aziz (2008) found in maize seedlings under different levels of water stress. They also reported a decline in RWC due to drought stress.

It is well established that plants subjected to water stress can accumulate inorganic solutes e.g., N, P, and K etc. Analysis of macronutrients N, P and K in the maize cultivar clearly indicates that water stress increased the shoot and root potassium ( $\text{K}^+$ ) concentrations in all maize cultivars. It is known that vacuolar cation ( $\text{K}^+$ ) could balance the cytoplasmic free proline osmotically (Weimberg *et al.*, 1982) but potassium made only a small contribution to osmotic adjustment in water stressed plants (Voetberg & Sharp, 1991). So, in the present study, the increased accumulation of potassium ( $\text{K}^+$ ) in maize seedlings might have played a significant role in plant survival under drought stress by playing an important role in osmotic adjustment.

Shoot nitrogen concentration decreased in all maize cultivars due to drought stress. In fact, soil drying induces a decrease in nutrients, particularly nitrogen with strong interactive effects on plant growth and functions (McDonald & Davies, 1996). However, the differential accumulation of root nitrogen and shoot and root phosphorous in all the cultivars under water stress cannot be related to their drought tolerance, which is in contrast to what has earlier been observed in different crops. The decrease in N concentration due to water stress has been reported in various crops including wheat (McDonald & Davies, 1996; Singh & Usha, 2003), in soybean and rice (Tanguilig *et al.*, 1987) and in maize (Premachandra *et al.*, 1990). On the other hand, Sarwar *et al.*, (1991) studied the response of different wheat varieties to water stress and reported a significant increase in N content under water stress.

In conclusion, plant biomass, photosynthetic rate, transpiration rate, stomatal conductance, relative water contents, and shoot N concentrations decreased under water deficit conditions in all maize cultivars. However, root/shoot ratio, chlorophyll *a*, *b* contents, chlorophyll *a/b* ratio, sub-stomatal CO<sub>2</sub> concentration, proline contents and potassium concentrations increased under water limited conditions. Of all maize cultivars examined in the present study cv. Agaiti-85, was found to be relatively resistant to drought stress.

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### References

Abrechit, D.G. and P.S. Carberry. 1993. The influence of water deficit prior to tassel initiation in maize growth, development and yield. *Field Crop Res.*, 31(1-2): 55-69.

Akram, N.A., M. Shahbaz and M. Ashraf. 2007. Relationship of photosynthetic capacity and proline accumulation with the growth of differently adapted populations of two potential grasses (*Cynodon dactylon* (L.) Pers. and *Cenchrus ciliaris* L.) to drought stress. *Pak. J. Bot.*, 39(3): 777-786.

Akram, N.A., M. Shahbaz and M. Ashraf. 2008. Nutrient acquisition in differentially adapted populations of *Cynodon dactylon* (L.) Pers. and *Cenchrus ciliaris* L., under drought stress. *Pak. J. Bot.*, 40(4): 1433-1440.

Ali, Q., M. Ashraf, M. Shahbaz and H. Humera. 2008. Ameliorating effect of foliar applied proline on nutrient uptake in water stressed maize (*Zea mays* L.) plants. *Pak. J. Bot.*, 40(1): 211-219.

Allen, S.E., H.M. Grimshaw and A.P. Rowland. 1986. In: *Methods in plant ecology*, (Eds.): P.D. Moore and S.B. Chapman, 2<sup>nd</sup> edition, Blackwell Sci. Publ., Oxford, pp. 285-344.

Anonymous. 2007. Pakistan Agriculture Research Council. All About/ Crop/ Maize. (Pakistan Com).

Arnon, D.T. 1949. Copper enzyme in isolated chloroplasts polyphenoloxidase in *Beta vulgaris*. *Plant Physiol.*, 24: 1-15.

Ashraf, M. and A. Bashir. 2003. Salt stress induced changes in some organic metabolites and ionic relations in nodules and other plant parts of two crop legumes differing in salt tolerance. *Flora*, 198: 486-498.

Ashraf, M. and N. Yasmin. 1995. Responses of four arid zone grass species from varying habitat to drought stress. *Biol. Plant.*, 37(4): 567-575.

Ashraf, M. and P.J.C. Harris. 2004. Potential biochemical indicators of salinity tolerance in plant. *Plant. Sci.*, 166: 3-16.

Ashraf, M., M. Arfan, M. Shahbaz, A. Ahmad and A. Jamil. 2002. Gas Exchange characteristics and water relations in some elite okra cultivars under water deficit. *Photosynthetica*, 40(4): 615-620.

Ashraf, M., S. Nawazish and H.R. Athar. 2007. Are chlorophyll fluorescence and photosynthetic capacity potential physiological determinants of drought tolerance in maize (*Zea mays* L.). *Pak. J. Bot.*, 39(4): 1123-1131.

Ashraf, M.Y., A.R. Azime, S.S.M. Nagvi and A.H. Khan. 1995. Alpha-amylase, protease activities and associated changes under water stress condition in wheat seedling. *Pak. J Sci Ind Res.*, 38: 430-434.

Ashraf, M.Y., S.A. Ali and A.S. Bhatti. 1998. Nutritional imbalance in wheat genotypes grown at soil water stress. *Acta Physiol. Plant.*, 20(3): 307-310.

Banoc, D.M., A. Yamauchi, A. Kamoshita, L.J. Wade and J.R. Pardales. 2000. Dry matter production and root system development of rice cultivars under fluctuating soil moisture. *Plant Prod. Sci.*, 3(2): 197-207.

Bartels, D. and D. Nelson. 1994. Approaches to improve stress tolerance using molecular genetics. *Plant Cell Environ.*, 17: 659-667.

Bates, L.S., R.P. Waldren and I.D. Teare. 1973. Rapid determinations of free proline for water stress studies. *Plant Soil*, 39: 205-207.

Boyer, J.S. 1996. Advances in drought tolerances in plants. *Adv. Agron.*, 56: 187-218.

Bremner, J.M. 1965. Total nitrogen and inorganic form of nitrogen. In: *Method of soil analysis*. (Ed.): C.A. Black. Amer. Soc. Agron. Madison, Wisconsin, pp. 1149-1237.

Chaves, M.M., J. Osorio and J.S. Pereira. 2004. Water use efficiency and photosynthesis. In: *Water use efficiency in plant biology*. (Ed.): M. Bacon. Oxford: Blackwell publishing, pp. 42-74.

Chaves, M.M., J.P. Maroco and J.S. Pereira. 2003. Understanding plant response to drought from genes to the whole plant. *Funct. Plant Biol.*, 30: 239-264.

Cornic, G. 1994. Drought stress and high light effects on leaf photosynthesis. In: *Photoinhibition of photosynthesis from molecular mechanism to the field*. (Eds.): N.R. Baker & J.R. Bowyer. Oxford, UK: Bios Scientific Publishers, 297-313.

Dubey, R.S. 2005. Photosynthesis in plants under stressful conditions. In: *Handbook of photosynthesis*. (Ed.): M. Passarkli. New York: Marcel and Decker., 859-875.

Edmeades, G.O., J. Bolanos, S.C. Chapman, H.R. Lafitte and M. Banziger. 1999. Selection improves drought tolerance in tropical maize populations: I. Gain in biomass, grain yield and harvest index. *Crop Sci.*, 39: 1306-1315.

Edwards, G.E. and G.A. Walker. 1983. *C<sub>3</sub>, C<sub>4</sub> mechanisms and cellular and environmental regulation of photosynthesis*. Oxford: Blackwell Scientific.

Elhrlinger, J.R. and R.K. Monson. 1993. Evolutionary and ecological aspects of photosynthetic pathway variation. *Annu. Rev. Ecol. Syst.*, 24: 411-439.

Estill, K., R.H. Delaney, W.K. Smith and R.L. Ditterline. 1991. Water relations and productivity of alfalfa leaf chlorophyll variants. *Crop Sci.*, 31: 1229-1233.

Farooq, M., T. Aziz and S.M.A. Basra. 2007. Water stress in maize cultivation, published in DAWN, the internet Ed., April 16, 2007.

Flexas, J. and H. Medrano. 2002. Drought inhibition of photosynthesis in C<sub>3</sub> plants, stomatal and non stomatal limitations revisited. *Ann. Bot.*, 89: 183-189.

Flexas, J., H. Medrano, J.M. Escalona, J. Bota and J. Gulias. 2002a. Regulation of photosynthesis of C<sub>3</sub> plants in response to progressive drought, the stomatal conductance as a reference parameter. *Ann. Bot.*, 89: 895-905.

Flexas, J., J. Bota, J.M. Escalona, B. Sampol and H. Medrano. 2002b. Effects of drought on photosynthesis in grapevines under field conditions an evaluation of stomatal and mesophyll limitations. *Funct. Plant Biol.*, 29: 461-471.

Garcia, A.L., A. Torrecillas, A. Lean and M.C. Ruiz-Sanchez. 1987. Biochemical indicators of water stress in maize seedlings. *Biol. Plant.*, 29: 45-48.

Giardi, M.T., A. Cona, B. Geiken, T. Kueera, J. Masajidek and A.K. Matoo. 1996. Long-term drought stress induces structural and function reorganization of photosystem II. *Planta*, 99: 118-125.

Hamada, A.M. 1996. Effect of NaCl, water stress or both on gas exchange and growth of wheat. *Biol. Plant.*, 38: 405-415.

Hamblin, A., D. Tennant and M.W. Perry. 1990. The cost of stress: Dry matter partitioning changes with seasonal supply of water and nitrogen to dry land wheat. *Plant Soil*, 122: 47-58.

Hanson, A.D., B. Rathinasabapathi, J. Rivoal, M. Burnet, M.O. Dillon and D.A. Gage. 1994. Osmoprotective compounds in the Plumbaginaceae: A natural experiment in metabolic engineering of stress tolerance. *Proc. Natl. Acad. Sci. U.S.A.*, 91: 306-310.

Hare, P.D., W.A. Cress, and J. Van Staden. 1999. Proline synthesis and degradation: a model system for elucidating stress-related signal transduction. *J. Exp. Bot.*, 50: 413-434.

Hsu, S.Y., Y.T. Hsu and C.H. Kao. 2003. The effect of polyethylene glycol on proline accumulation in rice leaves. *Biol. Plant.*, 46: 73-78.

Huang, B. and P.S. Nobel. 1992. Hydraulic conductivity and anatomy for lateral roots of *Agave* desert during root growth and drought induced abscission. *J. Exp. Bot.*, 43: 1441-1449.

Huang, B., R.R. Duncan and R.N. Carrow. 1997. Drought-resistance mechanisms of seven warm-season turfgrasses under surface soil drying: II. Root aspects. *Crop Sci.*, 37: 1863-1869.

Jackson, M.L. 1962. *Soil chemical analysis*. Contable Co. Ltd. London.

Janson, J.G., G.R. Thomas and D. Mason-pharr. 2004. Heat and drought influence photosynthesis, water relations, and soluble carbohydrates of two ecotypes of redbud (*Cercis canadensis*). *J. Am. Soc. Hort. Sci.*, 129(4): 497-502.

Ketchum, R.E.B., R.C. Warren, L.J. Klima, F. Lopez-Gutierrez and M.W. Nabors. 1991. The mechanism and regulation of proline accumulation in suspension cultures of the halophytic grass *Distichlis spicata* L. *J. Plant Physiol.*, 137: 368-374.

Kishor, P.B.K., Z. Hong, G.H. Miao, C.A.A. Hu and D.P.S. Verma. 1995. Overexpression of [ $\delta$ ]-pyrroline-5-carboxylate synthetase increases proline production and confers osmotolerance in transgenic plants. *Plant Physiol.*, 108: 1387-1394.

Lawlor, D.W. 1995. The effects of water deficit on photosynthesis. In: *Environment and plant metabolism*. (Ed.): N. Smirnoff. Oxford: Bios Scientific Publishers, 129-160.

Lawlor, D.W. 2002. Limitation to photosynthesis in water stressed leaves: stomata vs metabolism and role of ATP. *Ann. Bot.*, 89: 871-885.

Lopez, C.G., G.M. Banowetz, C.J. Peterson and W.E. Kronstad. 2003. Dehydrin expression and drought tolerance in seven wheat cultivars. *Crop Sci.*, 43: 577-582.

Madan, S., H.S. Nainawatee, R.K. Jain and J.B. Chowdhury. 1995. Proline and proline metabolizing enzymes in in-vitro selected NaCl-tolerant *Brassica juncea* L. under salt stress. *Ann. Bot.*, 76: 51-57.

Makela, P., M. Kontturi, E. Pehu and S. Somersalo. 1999. Photosynthetic response of drought and salt-stressed tomato and turnip rape plants to foliar applied glycine betaine. *Physiol. Plant.*, 105: 45-50.

Makela, P., R. Munns, T.D. Colmer, A.G. Condon and P. Peltonen-Sainio. 1998. Effect of foliar applications of glycinebetaine on stomatal conductance, abscisic acid and solute concentrations in leaves of salt-or drought-stressed tomato. *Aust. J. Plant Physiol.*, 25: 655-663.

Manabendra, D.K. and K. Baruah. 1998. Studies on physiological traits of rice (*Oryza sativa* L.) cultivars under moisture stress situations. *Ind. J. Ecol.*, 25: 192-196.

McDonald, A.J.S. and W.J. Davies. 1996. Keeping in touch: responses of the whole plant to deficits in water and nitrogen supply. *Adv. Botanic. Res.*, 22: 229-300.

Medrano, H., J.M. Esalona, J. Bota, J. Gulias and J. Flexas. 2002. Regulation of photosynthesis of C<sub>3</sub> plants in response to progressive drought, the stomatal conductance as a reference parameter. *Ann. Bot.*, 89: 895-905.

Molnar, I., L. Gaspar, L. Stehli, S. Dualai, E. Sarvari, I. Kiraly, G. Galiba and M. Molnar-Lang. 2002. The effects of drought stress on the photosynthetic process of wheat and *Aegilops biuncialis* genotypes originating from various habitats. *Acta Biol. Szegediensis*, 46: 115-116.

Moussa, H.R. 2006. Influence of exogenous application of silicon on physiological response of salt-stressed maize (*Zea mays* L.). *Int. J. Agric. Biol.*, 8: 293-297.

Moussa, H.R. and S.M. Abdel-Aziz. 2008. Comparative response of drought tolerant and drought sensitive maize genotypes to water stress. *Aust. J. Crop Sci.*, 1: 31-36.

Murchie, E.H. and P. Horton. 1997. Acclimation of photosynthesis to irradiance and spectral quality in British plant species: chlorophyll content, photosynthetic capacity and habitat preference. *Plant Cell Environ.*, 20: 438-448.

Natr, L. and D.W. Lawlor. 2005. Photosynthetic plant productivity. In: *Hand Book Photosynthesis*, 2<sup>nd</sup> (Ed.): M. Pessarakli, C.R.C. Press, New York, USA, pp. 501-524.

Nayyar, H. and D.P. Walia. 2003. Water stress induced proline accumulation in contrasting wheat genotypes as affected by calcium and abscisic acid. *Biol. Plant.*, 46: 275-279.

Nazir, N., M. Ashraf and R. Ejaz. 2001. Genomic relationships in oilseed Brassicas with respect to salt tolerance-photosynthetic capacity and ion relations. *Pak. J. Bot.*, 33: 483-501.

O, Toole, J.C. and W.L. Bland. 1987. Genotypic variation in crop plant root systems. *Adv. Agron.*, 41: 91-145.

Ober, E.S. and R.E. Sharp. 2003. Electrophysiological responses of maize roots to low water potentials: relationship to growth and ABA accumulation. *J. Exp. Bot.*, 54: 813-824.

Parida, A.K., V.S. Dagaonkar, M. S. Phalak, G.V. Umalkar and L.P. Aurangabadkar. 2008. Differential responses of the enzymes involved in proline biosynthesis and degradation in drought tolerant and sensitive cotton genotypes during drought stress and recovery. *Acta Physiol. Plant.*, 30: 619-627.

Parida, A.K., V.S. Dagaonkar, M.S. Phalak, G.V. Umalkar and L.P. Aurangabadkar. 2007. Alterations in photosynthetic pigments, protein and osmotic components in cotton genotypes subjected to short-term drought stress followed by recovery. *Plant Biotech Rep.*, 1: 37-48

Pedrol, N., P. Ramos and M.J. Reigosa. 2000. Phenotypic plasticity and acclimation to water deficits in Velvet grass: a long-term green-house experiment change in leaf morphology, Photosynthesis and stress induced meabolites. *J. Plant Physiol.*, 157: 383-393.

Peschke, G., C. Seidler and Yogal. 1997. Effect of drought during the growing season on Agricultural (*Triticum aestivum* L.) and forest plant canopy (*Piceae abies* L.). In: *Proc. 14<sup>th</sup> Int. Cong. Biomet.*, 2(2): Solvenia.

Premachandra, G.S., H. Saneoka, K. Eujita and S. Ogata. 1990. Cell membrane stability and leaf water relations as affected by phosphorus nutrition under water stress in maize. *Soil Sci. Plant. Nutr.*, 36: 661-666.

Raymond, M.J. and N. Smirnoff. 2002. Proline metabolism and transport in maize seedlings at low water potential. *Ann. Bot.*, 89: 813-823.

Sarwar, M., A. Nazir, G. Nabi and M. Yasin. 1991. Effect of soil moisture stress on different wheat varieties. *Pak. J. Agric. Res.*, 12(4): 275-280.

Serraj, R. and T.R. Sinclair. 2002. Osmolyte accumulation: Can it really help to increase crop yield under drought conditions? *Plant Cell Environ.*, 25: 333-341.

Sharp, R.E. and W.J. Davies. 1989. Regulation of growth and development of plants with a restricted supply of water. In: *Plants under stress*. (Eds.): H.G. Jone *et al.* Cambridge Univ. Press, Cambridge, England, pp. 71-93.

Siddique, M.R.B., A.H. Hamid and M.S. Islam. 1999. Drought stressed effect on photosynthetic rate and leaf gas exchange of wheat. *Botanical Bulletin of Academic Sinica*, 30(2): 141-145.

Sinclair, T.R. and M.M. Ludlow. 1985. Who taught plants thermodynamics? The unfulfilled potential of plant water potential. *Aust. J. Plant Physiol.*, 33: 213-217.

Singh, B. and K. Usha. 2003. Salicylic acid induced physiological and biochemical changes in wheat seedlings under water stress. *Plant Growth Regul.*, 39: 137-141.

Smucker, A.J.M., A. Nunez-Barrios and J.T. Ritchie. 1991. Root dynamics in drying soil environments. *Belowground Ecol.*, 1: 1-5.

Snedecor, G.W. and W.G. Cochran. 1980. *Statistical methods*. 7<sup>th</sup> Edition Iowa State Univ. Press, AMES, Iowa.

Srinivasa-Rao, N.K., R.M. Bhatt and A.T. Sadashiva. 2000. Tolerance to water stress in tomato cultivars. *Photosynthetica*, 38: 465-467.

Steel, R.G.D. and J.H. Torrie. 1980. *Principles and Procedures of Statistics, with special reference to Biological Science*. McGraw Hill Book Co., Inc., New York.

Tabassum, M.I. 2004. Development of maize under water stress areas. Published in DAWN, Internet Ed., November 14, 2004.

Tamura, T., K. Hara, Y. Yamaguchi, N. Koizumi and H. Sano. 2003. Osmotic stress tolerance of transgenic tobacco expressing a gene encoding a membrane-located receptor-like protein from tobacco plants. *Plant Physiol.*, 131: 454-462.

Tanguilig, V.C., E.B. Yamboo, J.C.O. Toole and S.K. De-Datta. 1987. Water stressed effects on leaf elongation, leaf water potential, transpiration and nutrient uptake in Rice, Maize and Soyabean. *Plant Soil*, 103: 155-180.

Tezara, W., V. Mitchell, S.P. Driscoll and D.W. Lawlor. 2002. Effects of water deficit and its interaction with CO<sub>2</sub> supply on the biochemistry and physiology of photosynthesis in sunflower. *J. Exp. Bot.*, 53(375): 1781-1791.

Turner, N.C. 1986. Crop water deficits, a decade of progress. *Adv. Agron.*, 39: 1-51.

Voetberg, G.S. and R.E. Sharp. 1991. Growth of the maize primary root at low water potentials. III. Role of increased proline deposition in osmotic adjustment. *Plant Physiol.*, 96: 1125-1130.

Weimberg, R., H.R. Lerner and A. Poljakoff-Mayber. 1982. A relationship between potassium and proline accumulation in salt stressed Sorghum bicolor. *Physiol. Plant*, 55: 5-10.

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