

SEASONAL CHANGES IN SOIL MICROBIAL BIOMASS CARBON UNDER A WHEAT-MAIZE CROPPING SYSTEM RECEIVING UREA AND FARMYARD MANURE IN DIFFERENT COMBINATIONS

TARIQ MAHMOOD*, REHMAT ALI, FAQIR HUSSAIN AND GHULAM RASUL TAHIR

*Nuclear Institute for Agriculture & Biology,
P.O. Box 128, Jhang Road, 38000 Faisalabad, Pakistan;*

Abstract

Seasonal changes in carbon (C) availability and soil microbial biomass carbon (C_{mic}) were studied under an irrigated wheat-maize system receiving urea at 50 or 100 kg N ha^{-1} year $^{-1}$ in combination with 8 or 16 t ha^{-1} year $^{-1}$ of farmyard manure (FYM). Treatment effects on C_{mic} were not visible during the wheat growing season, whereas fertilizer application significantly increased C_{mic} during the maize growing season. In unfertilized soil, C_{mic} was similar under two crops, whereas fertilized soils showed 11–56% higher C_{mic} under maize than under wheat. Under both crops, C availability, as assessed by aerobically mineralizable C (AMC) and total organic C, was generally higher in fertilized soils than in the unfertilized. Fertilized soils showed much higher AMC under wheat than under maize; the stimulatory effect being much more pronounced due to increasing application rate of FYM than that of urea. In fertilized soils, but not in the unfertilized, specific respiratory activity (SRA) of soil microbial biomass was twice higher under wheat as compared to that under maize. Results suggested that the soil microflora under wheat was probably dominated by 'r-strategists', which respired more C as CO_2 than that incorporated into microbial biomass. In contrast, 'k-strategists' dominated under maize, incorporating relatively more C into their biomass than that respired as CO_2 . Fertilizer application significantly increased C_{mic} turnover rate; while increasing the FYM application rate further increased the turnover rate, the increasing urea level had no effect. The overall high C_{mic} turnover rate, particularly in fertilized soils (1.69–2.29 year $^{-1}$) indicated that nutrient cycling through soil microbial biomass may be substantial under agro-climatic conditions prevailing in this region.

Introduction

Soil microbial biomass, carbon on the average, constitutes only about 1% of the soil organic carbon (Moore *et al.*, 2000), yet it may comprise up to 44% of the active pool of soil nitrogen (Liang *et al.*, 1999). Therefore, the size of soil C_{mic} pool and its turnover have significant bearing on the overall productivity of soils. Several studies indeed show a close relationship between soil microbial biomass and nutrient availability to plants (Jenkinson & Lad, 1981; Houot & Chaussod, 1995).

Application of mineral N fertilizers and manures has been reported to significantly influence the build-up and dynamics of soil microbial biomass (Bolton *et al.*, 1985; Powelson *et al.*, 1987; Mahmood *et al.*, 1997). While organic amendments generally increase C_{mic} (Mazzarino *et al.*, 1993; Goshal and Singh, 1995), there are diverse reports

*Corresponding author e-mail: mahmood114@hotmail.com

regarding the effect of mineral N fertilizers on the size and activity of soil microbial biomass. Application of mineral N fertilizers either had no effect (Moore *et al.*, 2000), or caused an increase in the content of C_{mic} (Houot & Chaussod, 1995; McCarty *et al.*, 1995; Goshal & Singh, 1995). Cropping history, type of plant cover, plant age, and climatic changes are also well known to influence soil microbial biomass and its activity, thus leading to considerable seasonal fluctuations in C_{mic} (Franzluebbers *et al.*, 1994; Xu & Juma, 1993; Mendes *et al.*, 1999; Moore *et al.*, 2000; Piao *et al.*, 2000). However, only few long-term field studies exist to ascertain the practical significance of these fluctuations on crop growth and nutrient cycling (Kaiser & Heinmeyer, 1993; Joergensen *et al.*, 1994; Goshal & Singh, 1995).

Studies on dynamics of soil microbial biomass have been particularly lacking under agro-climatic conditions prevailing in Pakistan. Previously, we reported seasonal changes in carbon availability and C_{mic} in a sandy-clay loam under a wheat-maize cropping system that had been under a long-term fertilizer trial and receiving for the past ten years 100-200 kg N ha⁻¹ year⁻¹, either as urea or as FYM (Mahmood *et al.*, 1997). However, information on soil microbial properties has been lacking for cropping systems in which mineral N and FYM are applied together in different combinations; farmers in Pakistan possessing small land holdings often follow this practice. The present report describes the seasonal fluctuations in soil microbial biomass and its activity under a wheat-maize cropping system fertilized with two urea levels in combination with two levels of FYM.

Materials and Methods

The experimental site at the Nuclear Institute for Agriculture & Biology, Faisalabad is located at 73.2° longitude, 31.4° latitude and 183m above the sea level. The area has a semiarid subtropical climate with a mean annual rainfall of 340 mm, most of which is received as heavy monsoon downpours during July and August. The hottest months are May and June, with mean maximum temperatures of 39.4° and 41.1°C, respectively, whereas January is the coldest month with a mean minimum temperature of 5°C. The soil belongs to Hafizabad series (Haplic Yermosol; Anon., 1966) and is a deep, well-drained sandy-clay loam developed in a mixed calcareous medium-textured alluvium derived from Himalayas. The soil had been under a long-term soil fertility experiment on irrigated wheat-maize cropping system receiving different urea and/or farmyard-manure (FYM) treatments for the past ten years.

Twenty experimental plots (7.5×8.5 m) were established for five treatments in a randomized complete block design, each with four replicates. The treatments included: an unfertilized control and urea applied at 50 and 100 kg N ha⁻¹ year⁻¹, each combined with either 8 or 16 t ha⁻¹ year⁻¹ of FYM. Treatment details and physicochemical properties of the soil after ten years under different fertilizer treatments are given in Table 1. Wheat (*Triticum aestivum* L. cv. Pak-81) was sown on 6th December and harvested on 5th May, whereas maize (*Zea mays* L. cv. Akbar) was sown on 29th August and harvested as fodder on 30th October. A total of six irrigations were applied during the wheat season: all were equivalent to 75 mm except the first (presowing) and the fourth which were 100 mm and 50 mm, respectively. During the maize season, five irrigations were applied: all being equivalent to 75 mm except the first (presowing) and the last, which were 100 mm and 50 mm, respectively. The soil was sampled five and three times during the wheat and maize growing season, respectively. From each replicate plot, ten soil cores (3×15 cm, diameter×depth) were randomly collected, pooled, mixed, visible roots manually removed, and the soils sieved (<2 mm). The soil moisture content at the time of sampling

was equivalent to field capacity. Soils were not stored and all analyses were carried out within six hours of collection.

Total organic carbon (TOC) of the soil was determined by an acid-dichromate method (Riehm & Ulrich, 1954). For the measurement of C_{mic} , the chloroform fumigation-incubation method of Jenkinson & Powlson (1976) was followed using 10g field moist soil. The fumigated-reinoculated and unfumigated (control) soils were incubated at 30°C for 10 days in air-tight 100-ml serum vials, and the headspace was analyzed for CO₂ on a gas chromatograph (Gasukuro Kogyo, Model 370) equipped with a thermal-conductivity detector. Microbial biomass C was calculated as: $C_{mic}=F/k_C$, where F is the flush of CO₂-C (CO₂-C evolved from the fumigated minus that from the unfumigated control) and k_C is the recovery factor, the value of which was taken as 0.41 (Voroney & Paul, 1984). Microbial biomass turnover rate was calculated by dividing the sum of losses of C_{mic} at different sampling intervals by the overall average C_{mic} (McGill *et al.*, 1986). Microbial biomass carrying capacity (MBCC) is defined as the amount of CO₂-C evolved from the fumigated-inoculated soil during a 10-day incubation at 30°C divided by a k_C factor of 0.41. Aerobically mineralizable carbon was taken as the amount of CO₂-C evolved from the unfumigated soil during a 10-day incubation at 30°C. Specific respiratory activity (SRA) of soil microbial biomass was calculated by dividing AMC by C_{mic} (Franzluebbers *et al.*, 1994). Relative mineralization rate of soil organic carbon was estimated by dividing AMC by TOC. Data were subjected to an analysis of variance, followed by Duncan's multiple range test (Steel & Torrie, 1980), and results are reported as means of four replicate plots.

Results

Seasonal changes in C availability and C_{mic} : Carbon availability showed marked temporal variation during the growing seasons of wheat and maize (date means, Tables 2 & 3). Total organic C was relatively higher during the active growth period of both crops, when its values were comparable under wheat and maize. It declined toward the crop maturity in maize but even higher values recorded near the wheat maturity indicate a contribution from the decomposing root material. The mineralization of organic C pool (expressed as AMC) was strongly influenced by the growth stage of crops; in wheat, it was highest during the active growth period, whereas it continued to increase during the maize growing period. Relative C mineralization rate (expressed as mg CO₂-C g⁻¹ TOC), was though quite low (2.9–10.2 mg CO₂-C g⁻¹ TOC; equivalent to <1.0% of TOC), yet it showed marked seasonal trend, with maximum observed during the active growth period under both crops.

Microbial biomass carrying capacity and C_{mic} showed a distinct temporal pattern, which coincided with the temporal fluctuations in C availability, i.e. maximum during the active growth stage, and declining toward the crop maturity. The ratio of C_{mic} to TOC was small, ranging from 0.46–1.53%, the higher values corresponding to the active growth period of the two crops. Under both crops, SRA of soil microbial biomass was highest at the flowering stage ($P<0.05$) indicating a fluctuating metabolic status of microbial communities during the crop growth. Averaged across treatments, the SRA values during the active growth period of wheat were significantly higher than those observed during the active growth stage of maize ($P<0.05$). Compared to that of SRA, an opposite trend prevailed for C_{mic} ($P<0.05$).

Table 4. Mineralization and turnover of soil microbial biomass carbon (C_{mic}) under wheat-maize cropping system receiving different fertilizer treatments.

Treatment	ΣC _{mic} flux (kg ha ⁻¹ year ⁻¹)	C _{mic} turnover rate (year ⁻¹) ^a
U ₅₀ +F ₈	317 a ^b	1.69 (216) ^b b
U ₅₀ +F ₁₆	265 b	2.29 (159) a
U ₁₀₀ +F ₈	252 b	1.88 (194) b
U ₁₀₀ +F ₁₆	247 b	2.22 (164) a
Unfertilized	254 b	1.04 (351) c

^aFigures in parentheses are the C_{mic} turnover time (days)^bFigures in a column followed by different letter are significantly different ($P<0.05$) by Duncan's multiple range test

Treatment effects on carbon availability and C_{mic}: Total organic carbon of the soil increased as a result of fertilization (15–29% increase under wheat, 15–23% increase under maize, $P<0.05$), except that U₁₀₀+F₈ treatment had no effect during the wheat growing season (treatment means, Table 2 & 3). Treatment effects on the relative mineralization rate of TOC were inconsistent under two crops. However, averaged across all sampling dates under wheat and maize, only U₅₀+F₈ showed slightly higher rate (9% increase over control, $P<0.05$), while other treatments slightly decreased the rate (6–12% decrease over control, $P<0.05$). Treatment effects on AMC were much more pronounced compared to other parameters. At a given FYM application rate, doubling the urea application rate either did not affect the level of AMC, or caused a little (7%) increase but only during the maize season under higher FYM application rate. However, with both urea application rates, doubling the FYM application caused an 8–18% increase in AMC ($P<0.05$). The overall carbon availability to microbes expressed as MBCC increased due to fertilization (18–36% increase under wheat, 15–29% increase under maize, $P<0.05$). Treatment effects on C_{mic} however, followed a pattern that was not consistent with that observed for C availability parameters. During the wheat growing season, the effect of fertilization on C_{mic} was observed only in U₅₀+F₈ treatment, which showed an increase of 18% over the unfertilized control ($P<0.05$). A pronounced effect of fertilization on C_{mic} was recorded during the maize growing season, showing a 15–36% increase over the unfertilized ($P<0.05$). On the other hand, treatment effects on SRA were more obvious under wheat than under maize. Averaged across sampling dates, SRA in the unfertilized soil was similar under both crops. However, in fertilized treatments it was 45–162% higher under wheat than under maize. The stimulatory effect on SRA was higher due to increasing FYM than urea application rate.

Turnover of C_{mic}: Fertilizer application substantially increased the turnover of C_{mic} (63–120% increase over unfertilized, $P<0.05$) (Table 4). Increasing urea application from 50–100 kg N ha⁻¹ had no effect, but doubling the FYM application rate produced an 18–35% increase in the C_{mic} turnover rate ($P<0.05$). Except that U₅₀+F₈ treatment showed higher values, fertilizer application had no effect on the flux of C_{mic} (Table 4). However, the flux was significantly but negatively correlated with the concurrently determined soil mineral N ($r=-0.611$, $P<0.01$; point data for soil mineral N not shown).

Discussion:

Seasonal fluctuations in different carbon availability indices and microbial properties were not related to soil temperature. This is consistent with findings of Raubuch and Joergensen (2002) but contrary to several earlier reports describing seasonal variations in

soil C as a function mainly of soil temperature (Tate *et al.*, 1993; Zak *et al.*, 1993; Carnol & Ineson, 1999). An inverse relationship has also been reported between soil temperature and C_{mic} (Piao *et al.*, 2000).

Increase in soil organic C due to organic amendments is well documented (Marschner *et al.*, 2003) and the higher TOC values observed in the present study during the active crop growth periods indicate contribution by the wheat and maize crops in the form rhizodeposits. These rhizodeposits not only served as C source for the soil microflora (as indicated by increased AMC during the active plant growth), but might also have accelerated mineralization of the native pool of soil organic C. Our contention is supported by the data on relative mineralization rate of soil organic C (mg AMC g⁻¹ TOC), which followed a strong seasonal trend i.e. being higher during the active crop growth, particularly during the wheat growing season. Soil microbial biomass is known to be triggered into activity by trace amounts of substrates like root exudates, resulting in 'priming effect' on decomposition of the native pool of soil organic matter (Nobili *et al.*, 2001).

Microbial biomass carbon was a minor fraction of soil organic matter, constituting on the average 0.91% and 1.11% of TOC during the wheat and maize growing seasons, respectively. These figures are close to those (about 1.0%) reported by Moore *et al.*, (2000). However, this minor component of the soil organic matter showed marked seasonal fluctuations, and was also strongly influenced by the application of urea and FYM. An increase in C_{mic} due to application of mineral N fertilizer, as observed during the maize growing season, agrees to some earlier reports (Mahmood *et al.*, 1997; Raiesi, 2004). The C_{mic} was not correlated with carbon availability indices such as TOC and AMC, whether utilizing point data ($n=40$), date means ($n=8$) or treatment means ($n=10$). These results are contrary to the reports of Ross *et al.*, (1995) which related temporal fluctuations in C_{mic} to fluctuations in TOC.

The temporal trend of C availability, soil microbial biomass and its activity was much more pronounced as compared to treatment effects. It seems that treatment effects on C_{mic} were ramified due to temporal variations in the microbial community structure. As evidenced from the AMC data, carbon availability to microbes was consistently higher under wheat than under maize. However, proportionally less C was incorporated into microbial biomass than that respired under wheat than under maize. This is also evident from SRA data, which showed a highly significant but negative correlation with C_{mic} (under wheat: $r=-0.579$, $n=25$, $p<0.001$; under maize: $r=-0.780$, $n=15$, $p<0.001$; under both crops: $r=-0.628$, $n=40$, $p<0.001$). Specific respiratory activity of soil microbial biomass was also negatively correlated with C_{mic} :TOC ratio (under wheat: $r=-0.532$, $n=25$, $p<0.01$; under maize: $r=-0.736$, $n=15$, $p<0.001$; under both crops: $r=-0.591$, $n=40$, $p<0.001$). A strong seasonal pattern of SRA, which was opposite to that observed for C_{mic} , has also been reported under continuous corn (Salinas-Garcia *et al.*, 1997). The higher SRA and lower C_{mic} values under wheat than under maize indicate that the rhizodeposits under wheat compared to maize crop probably favoured the microbial community that was dominated by 'r-strategists' which respire more CO₂ per unit degradable C, incorporating relatively less C into biomass (Lynch, 1984; Insam, 1990). In contrast, 'k-strategists' probably dominated during the maize growing season, respiring relatively less C as CO₂ and incorporating more into biomass. Plant age, fertilizer application, rhizosphere development and substrate loading rates are well known to cause shifts in the microbial community structure (Griffiths *et al.*, 1999; Steer & Harris, 2000;

O'Donnell *et al.*, 2001). A strong but negative correlation between SRA and C_{mic} indicates that SRA of the soil microflora was a decisive factor in determining the size of soil microbial biomass pool under the cropping system investigated.

The increased C_{mic} turnover rate due to fertilizer application rate is consistent with the results of our earlier study (Mahmood *et al.*, 1997). In addition, lack of the effect due to increasing urea application rate and a pronounced stimulatory effect of increasing FYM application rate on the C_{mic} turnover are also in agreement with our previous findings (Mahmood *et al.*, 1997). The relatively high C_{mic} turnover rates observed during the present study also concur to some earlier reports (Mahmood *et al.*, 1997; Raiesi, 2004) indicating a considerable nutrient cycling through soil microbial biomass under irrigated cropping systems in this region.

A negative relationship between C_{mic} flux and soil mineral N indicates that seasonal fluctuations in soil mineral N also contributed significantly to the buildup and decomposition of soil microbial biomass; the lower soil mineral N levels favored the decomposition of microbial biomass and *vice versa*. Considering the annual fluxes of C_{mic} (Table 4), and assuming a C_{mic} : N_{min} ratio of 6, about 40 kg N ha⁻¹ year⁻¹ (in unfertilized) to 53 kg N ha⁻¹ year⁻¹ (in U₅₀+F₈ treatment) might be released due to the death and decay of microbial biomass under the cropping system investigated. This further suggests that nutrient cycling through soil microbial biomass may be substantial under agroclimatic conditions prevailing in this region.

Acknowledgements

The authors thank Muhammad Asad Mehmood and Anwar-ul-Haq for their valuable technical assistance. Equipment grant by the Alexander-von-Humboldt Foundation, Germany is also gratefully acknowledged.

References

- Bolton, H. Jr., L.F. Elliott and R.I. Papendick. 1985. Soil microbial biomass and selected soil enzyme activities: Effect of fertilization and cropping practices. *Soil Biol. Biochem.*, 17: 297–302.
- Carnol, M. and P. Ineson. 1999. Environmental factors controlling NO_3^- leaching, N_2O emissions and numbers of NH_4^+ oxidizers in a coniferous forest soil. *Soil Biol. Biochem.*, 31: 979–999.
- Anonymous. 1966. *Guidelines for Soil Profile Description*. Soil Survey and Fertility Branch, Land and Water Development Division, Food and Agricultural Organization of the United Nations, Rome.
- Franzleubbers, A.J., F.M. Hons and D.A. Zuberer. 1994. Seasonal changes in soil microbial biomass and mineralizable C and N in wheat management systems. *Soil Biol. Biochem.*, 26: 1469–1475.
- Goshal, N. and K.P. Singh. 1995. Effects of farmyard manure and inorganic fertilizer on dynamics of soil microbial biomass in a tropical dryland agroecosystem. *Biol. Fertil. Soils*, 19: 232–238.
- Griffiths, B.S., K. Ritz, N. Ebblewhite and G. Dobson. 1999. Soil microbial community structure; effects of substrate loading rates. *Soil Biol. Biochem.*, 31: 145–153.
- Houot, S. and R. Chaussod. 1995. Impact of agricultural practices on the size and activity of the microbial biomass in a long-term field experiment. *Biol. Fertil. Soils*, 19: 309–316.
- Insam, H. 1990. Are soil microbial biomass and basal respiration governed by the climatic regime? *Soil Biol. Biochem.*, 22:525–532.
- Jenkinson, D.S. and J.N. Lad. 1981. Microbial biomass in soil: Measurement and turnover. In: *Soil Biochemistry* (Eds.): E.A. Paul & N. Ladd, Vol. 5, Marcel Dekker, New York, pp. 415–471.

Jenkinson, D.S. and D.S. Powlson. 1976. The effects of biocidal treatments on metabolism in soil—V. A method for measuring soil biomass. *Soil Biol. Biochem.*, 8: 209–213.

Joergensen, R.G., B. Meyer and T. Muller. 1994. Time-course of the soil microbial biomass under wheat; A one year field study. *Soil Biol. Biochem.*, 26: 987–994.

Kaiser, E.A. and O. Heinmeyer. 1993. Seasonal variations of soil microbial biomass carbon within plough layer. *Soil Biol. Biochem.*, 25: 1649–1655.

Liang, B.C., A.F. Mackenzie and E.G. Gregorich. 1999. Changes in ^{15}N abundance and amounts of biologically active soil nitrogen. *Biol. Fertil. Soils*, 30: 69–74.

Lynch, J.M. 1984. Interactions between biological processes, cultivation and soil structure. *Plant Soil*, 76: 307–318.

Mahmood, T., F. Azam, F. Hussain and K.A. Malik. 1997. Carbon availability and microbial biomass in soil under an irrigated wheat-maize cropping system receiving different fertilizer treatments. *Biol. Fertil. Soils*, 25: 63–68.

Marschner, P., E. Kandeler and B. Marschner. 2003. Structure and function of soil microbial community in a long term fertilizer experiment. *Biol. Fertil. Soils*, 35: 453–461.

Mazzarino, M.J., L. Szott and M. Jimenez. 1993. Dynamics of soil total C and N, microbial biomass, and water-soluble C in tropical agroecosystems. *Soil Biol. Biochem.*, 25: 205–214.

McCarty, G.W., J.J. Meisinger and F.M.M. Jenniskens. 1995. Relationships between total-N, biomass-N and active-N in soil under different tillage and fertilizer treatments. *Soil Biol. Biochem.*, 27: 1245–1250.

McGill, W.B., K.R. Canon, J.A. Robertson and F.D. Cook. 1986. Dynamics of soil microbial biomass and water soluble organic C in Breton L after 50 years of cropping to two rotations. *Can. J. Soil Sci.*, 66: 1–19.

Mendes, I.C., A.K. Bandick, R.P. Dick and P.J. Bottomley. 1999. Microbial biomass and activities in soil aggregates affected by winter cover crops. *Soil Sci. Soc. Am. J.*, 63: 873–881.

Moore, J.M., S. Klose and M.A. Tabatabai. 2000. Soil microbial biomass carbon and nitrogen as affected by cropping systems. *Biol. Fertil. Soils*, 31: 200–210.

Nobili, M.D., M. Contin, C. Mondini and P.C. Brookes. 2001. Soil microbial biomass is triggered into activity by trace amounts of substrate. *Soil Biol. Biochem.*, 33: 1163–1170.

O'Donnell, A.G., M. Seasman, A. Macrae, I. Waite and J.T. David. 2001. Plants and fertilizers as drivers of change in microbial community structure and function in soils. *Plant Soil*, 232: 135–145.

Piao, H.C., Y.T. Hong and Z.Y. Yuan. 2000. Seasonal changes of microbial carbon related to climatic factors in soils from karst areas of southwest China. *Biol. Fertil. Soils*, 30: 294–297.

Powlson, D.S., P.C. Brookes and B.T. Christensen. 1987. Measurement of soil microbial biomass provides an early indication of changes in total soil organic matter due to straw incorporation. *Soil Biol. Biochem.*, 19: 159–164.

Raiesi, F. 2004. Soil properties and N application effects on microbial activities in two winter wheat cropping systems. *Biol. Fertil. Soils*, 40: 88–92.

Raubuch, M. and R.G. Joergensen. 2002. C and net N mineralization in a coniferous forest soil: the contribution of temporal variability of microbial biomass C and N. *Soil Biol. Biochem.*, 34: 841–849.

Riehm, H. and B. Ulrich. 1954. Quantitative colorimetrische Bestimmung der organischen Substanz im Boden. *Landwirtsch. Forsch.*, 6: 173–176.

Ross, D.J., T.W. Speir, H.A. Kettles and A.D. Mackay. 1995. Soil microbial biomass, C and N mineralization and enzyme activities in a hill pasture: influence of season and slow-release P and S fertilizer. *Soil Biol. Biochem.*, 27: 1431–1443.

Salinas-Garcia, J.R., F.M. Hons, J.E. Matocha and D.A. Zuberer. 1997. Soil carbon and nitrogen dynamics as affected by long-term tillage and nitrogen fertilization. *Biol. Fertil. Soils*, 25: 182–188.

Steel, R.G.D. and J.H. Torrie. 1980. *Principles and Procedures of Statistics*. McGraw Hill, New York.

Steer, J. and A. Harris. 2000. Shifts in the microbial community in rhizosphere and non-rhizosphere soils during the growth of *Agrostis stolonifera*. *Soil Biol. Biochem.*, 32: 869–878.

Tate, K.R., D.J. Ross, B.J. O'Brien and F.M. Kelliher. 1993. Carbon storage and turnover, and respiratory activity in the litter and soil of an old-growth southern beech (*Nothofagus*) forest. *Soil Biol. Biochem.*, 25: 1601–1612.

Voroney, R.P. and E.A. Paul. 1984. Determination of k_C and k_N in situ for calibration of the chloroform fumigation-incubation method. *Soil Biol. Biochem.*, 16: 9–14.

Xu, J.G. and N.G. Juma. 1993. Above- and below-ground transformation of photosynthetically fixed carbon by two barley (*Hordium vulgare* L.) cultivars in a typic cryoboroll. *Soil Biol. Biochem.*, 25: 1263–1272.

Zak, D.R., D.F. Grigal and L.F. Ohrmann. 1993. Kinetics of microbial respiration and nitrogen mineralization in great lake forest. *Soil Sci. Soc. Am. J.*, 57: 1100–1106.

(Received for publication 25 August 2004)